

STABILITY-INDICATING RP-HPLC METHOD AND ITS VALIDATION FOR ANALYSIS OF METFORMIN HYDROCHLORIDE & SAXAGLIPTIN IN BULK AND PHARMACEUTICAL DOSAGE FORM

Naidu Srinivasa Rao*, Dr. K. Venkataramana, and P.Anitha

Department of Pharmacy, Acharya Nagarjuna University, Andhra Pradesh, India.

Article Received on
11 Nov 2014,

Revised on 06 Dec 2014,
Accepted on 31 Dec 2014

***Correspondence for
Author**

Naidu Srinivasa Rao
Department of Pharmacy,
Acharya Nagarjuna
University, Andhra
Pradesh, India.

ABSTRACT

A simple, rapid, precise, sensitive and reproducible reverse phase high performance liquid chromatography (RP-HPLC) method has been developed for the quantitative analysis of Metformin hydrochloride & Saxagliptin in pharmaceutical dosage forms. Chromatographic separation of MET & SAXA was achieved on Inertsil C8, 150mm x 4.6mm, 5 μ m and the mobile phase containing P^H 2.5 buffer & Acetonitrile in the ratio of 70:30 v/v. The flow rate was 1.0 ml/min, detection was carried out by absorption at 229nm using a photodiode array detector at ambient temperature. The RT of Metformin hydrochloride and Saxagliptin is found to be 2.8 and 5.2 min. The drugs were exposed to thermal, photolytic, hydrolytic, acid, alkali, and

oxidative stress and the stressed samples were analyzed by use of the proposed method & chromatograms from the stressed samples, obtained by use of the photodiode-array detector. The linearity of the method was excellent over the range 50-740 μ g/ml and 0.5-7.5 μ g/ml for MET & SAXA respectively. The correlation coefficient was 0.999. The proposed method was validated according to ICH guidelines. And it was found to be suitable and accurate method for quantitative analysis of Dosage form and study of its stability.

KEYWORDS: High performance liquid chromatography, forced degradation, Metformin hydrochloride, Saxagliptin.

INTRODUCTION

Saxagliptin (rINN) is (1S,3S,5S)-2-((2S)-Amino(3-hydroxytricyclo(3.3.1.1^{3,7})dec-1-yl)acetyl)-2-azabicyclo (3.1.0) hexane-3-carbonitrile an orally active hypoglycemic (anti-

diabetic drug) of the new dipeptidyl peptidase-4 (DPP-4) inhibitor class of drugs and Metformin hydrochloride is N,N-dimethyl imidocarbonyl diamide is used in the treatment of type 2 diabetes. Structures are shown in fig 1 and 2 respectively. Saxagliptin works to competitively inhibit the enzyme dipeptidyl peptidase 4 (DPP-4) affecting the action of natural hormones in the body called incretins they are able to potentiate the secretion of insulin and suppress the release of glucagon by the pancreas. This drives blood glucose levels to normal. Metformin hydrochloride activates AMP-activated protein kinase (AMPK), a liver enzyme that plays an important role in insulin signaling, whole body energy balance, and the metabolism of glucose and fats; activation of AMPK is required for Metformin hydrochloride inhibitory effect on the production of glucose by liver cells.

Metformin is official in IP^[11] and USP,^[12] while Sitagliptin is not official in any pharmacopoeias. Literature survey reveals, UV,^[1] HPLC^[2] methods for analysis of Metformin as single and combined dosage forms with other drugs and UV,^[3] HPLC^[4] methods for analysis of Sitagliptin as single component systems. Few methods are reported for simultaneous determination of Metformin hydrochloride and Saxagliptin by reverse phase HPLC^[5-10] in pharmaceutical dosage forms. As one or two stability HPLC methods have been reported for the determination of Saxagliptin and Metformin hydrochloride an attempt was made to report a simple, sensitive, validated and economic method for the determination of Saxagliptin and Metformin hydrochloride

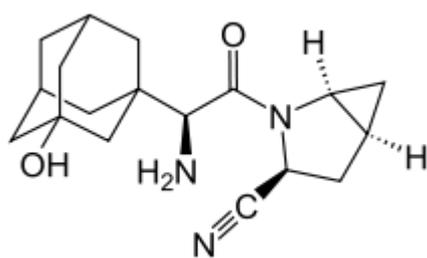


Fig:1 Saxagliptin

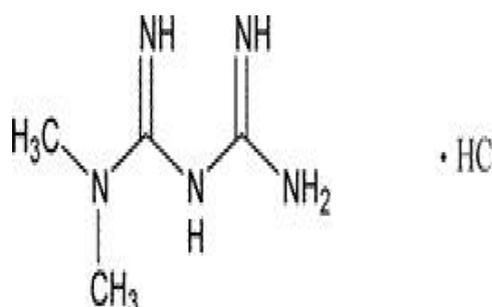


Fig:2 Metformin hydrochloride

MATERIALS AND METHODS

Instruments Used

All analytical works performed on Waters model LC-20AD dual pump, a Waters model DGU-20A degasser, Waters model SPD-M20A photo diode array (PDA) detector and a

Waters model SIL-20A HT auto injector, Empower2 solution version software, Inertsil C8 column (150 X 4.6mm, 5 µm particle size) as stationary phase, a calibrated electronic single pan balance Shimadzu (AUX-220), a pH meter of Elico (LI-120) and ultrasonic cleaner (SONICA) were also used during the analysis.

Reagents and chemicals

Analytically pure Metformin hydrochloride and Saxagliptin were obtained as gift samples from Hyderabad, India. Tablet (Kombiglyze) was purchased from the local market. Buffers, ACN and all other chemicals were analytical grade.

Method Development

Preparation of Buffer: Weigh 1.50gm of Sodium Hexane-1-sulphonic acid is dissolved into 1lt Water adjust pH-2.5 with OPA.

Mobile phase: Prepare a mixture of Buffer and ACN in the ratio of (70:30). Filter and degas.

Chromatographic condition Use suitable High Performance Liquid Chromatography equipped with UV-visible detector.

Column : Inertsil C8, 150mm x 4.6mm, 5µm.

Wavelength : 229 nm

Injection Volume : 10µL

Column Temperature : Ambient

Flow rate : 1.0 mL/min.

Retention time of Metformin hydrochloride is about 3.0-4.0 min and Saxagliptin is about 5.0-6.0min.

Preparation of Diluent: Used mobile phase as diluents.

Preparation of standard solution of Metformin hydrochloride: Weigh accurately about 500 mg of Metformin hydrochloride working standard is taken into 100ml volumetric flask. Add 70 mL of diluent, sonicate to dissolve and dilute to volume diluent. Further dilute 5 mL to 50 mL with the diluent.

Preparation of standard solution of Saxagliptin: Weigh accurately about 10mg of Saxagliptin working standard are taken into 200ml volumetric flask. Add 70 mL of

diluent, sonicate to dissolve and dilute to volume diluent. Further dilute 5mL to 50mL with the diluent.

Preparation of Sample solution: Weigh 10tablets and crush the tablets weigh powder then take tabletpowder equivalent to metformin and saxagliptin of sample into a 200 mL volumetric flask. Add 70 mL of diluent, sonicate to dissolve and dilute to volume diluent. Further dilute 5 mL to 50 mL with the diluent. Filter through 0.45 μ Nylon syringe filter.

Procedure Inject 10 μ l of Standard preparation five times and Sample preparation in the Chromatograph. Record the chromatograms and measure the peak responses for Metformin hydrochloride & Saxagliptin. The System suitability parameters should be met. From the peak responses, calculate the content of Metformin hydrochloride & Saxagliptin in the sample. The results are shown in “Fig:3” & “Fig:4” and “Table:1”

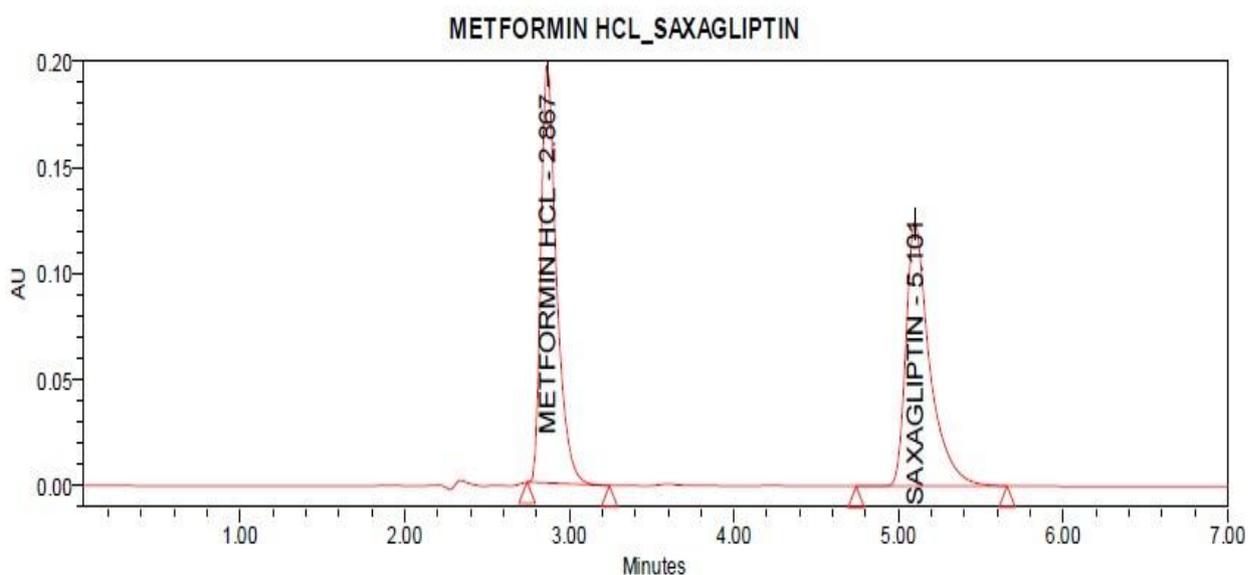


Fig: 3 Standard chromatogram of Metformin & Sitagliptin.

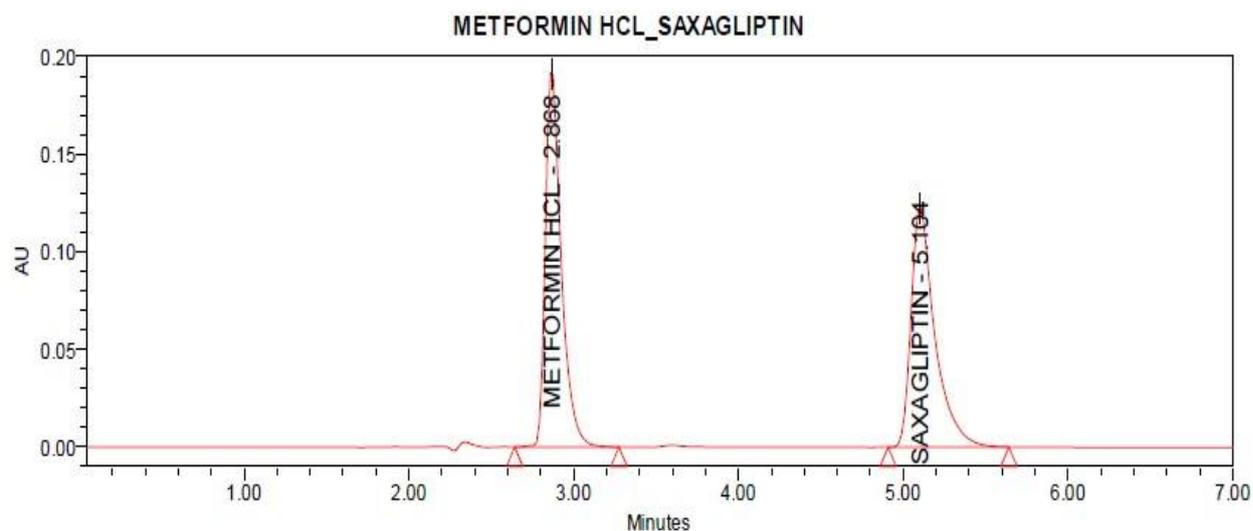


Fig: 3 Sample chromatogram of Metformin & Sitagliptin.

Table 1. Assay results of Metformin hydrochloride and Saxagliptin in combined dosage form.

Drug	Label claim	% Drug found \pm SD	% RSD
Metformin hydrochloride	500 mg	99.8	0.18
Saxagliptin	5 mg	99.6	0.23

Evaluation of system suitability

1. Relative Standard Deviation of five replicate injections of Standard preparation for Metformin hydrochloride & Saxagliptin peaks should not be more than 2.0%.
2. The tailing factor for Metformin hydrochloride & Saxagliptin peaks should be more than 2.0 and plate count will be not less than 3000.

The results are shown in “Table:2”.

Table: 2 System suitability parameters.

Parameter	Metformin hydrochloride	Saxagliptin
USP tailing factor	1.50	1.65
Theoretical plates	3481	6692
%RSD for Areas	0.48	0.335
%RSD for RT	0.10	0.13

Method validation

This method described above had been validated as per the ICH guidelines for the parameters like accuracy, linearity, precision, detection limit, quantitation limit and robustness. And the results were summarized below.

System suitability

The system suitability was assessed using five replicate analyses of drugs at concentration of 500µg/mL for MET and 5µg/mL for TEL by increasing the injection volumes 10-50µL.

Specificity

Specificity studies were carried for both pure drug and drug product by comparing the plots with blank and placebo. Peak purity tests were also carried out to show that the analyte chromatographic peak is not attributable to more than one component as the impurities are not available by purity index data.

Forced Degradation studies

Forced degradation study was carried out by treating the sample under the following conditions. Sample Stock solution is from Method Precision sample flask.

a) Acid degradation

5 ml of the above stock solution was transferred into 100ml volumetric flask and added 60ml of diluent, treated with 5.0ml of 5N hydrochloric acid and heated at 60°C for 10 minutes, and cooled, neutralized with 5ml of 5N sodium hydroxide and diluted to volume with diluent and was analyzed as per the test method.

b) Alkali degradation

5 ml of the above stock solution was transferred into 100ml volumetric flask and added 60ml of diluent, treated with 5.0ml of 5N sodium hydroxide and heated at 60°C for 10 minutes, and cooled, neutralized with 5ml of 5N hydrochloric acid and diluted to volume with diluent and was analyzed as per the test method.

c) Peroxide degradation

5 ml of the above stock solution was transferred into 100ml volumetric flask and added with 60ml diluent was treated with 5 ml of 30% v/v solution of hydrogen peroxide and heated at 60°C for 10 minutes, cooled and diluted to volume with diluent and was analyzed as per the test method.

d) Reduction

5 ml of the above stock solution was transferred into 100ml volumetric flask and added with 60ml diluent was treated with 5 ml of 1N solution of sodium bicarbonate and heated at 60°C for 10 minutes, and cooled, made to volume with diluent and was analyzed as per the test method.

e) Photolytic degradation

Sample was exposed to 1.2 Million lux hours of light and analyzed the exposed sample as per test procedure.

f) Thermal degradation

Sample was kept in hot air oven at 60°C for 1 hour. Treated sample was analyzed as per the test method.

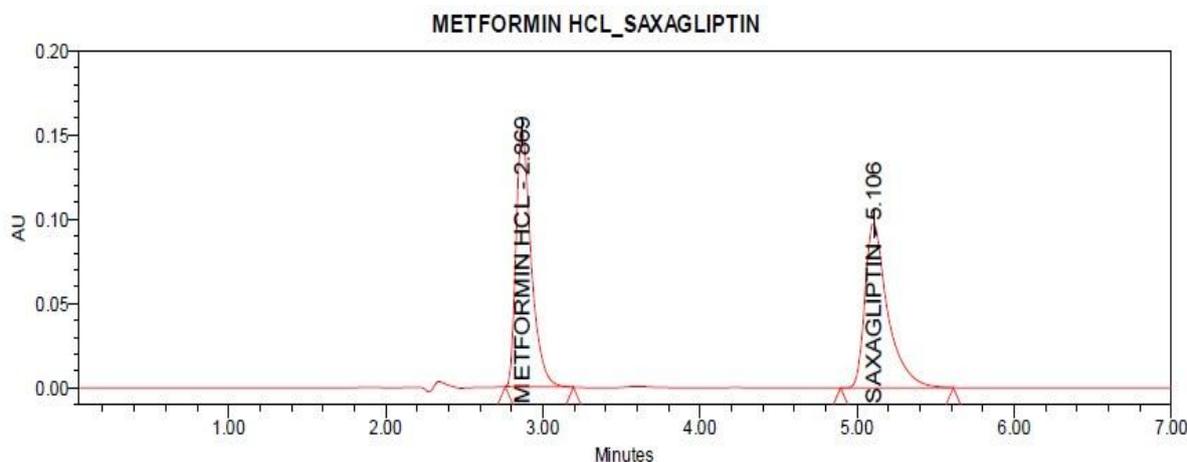
The results are shown in “Fig:5” to “Fig:10” and “Table:3” & “Table:4”.

Table: 3 Data for forced degradation studies of Metformin Hydrochloride.

Parameter	Area count	%label claim	% degradation
Control	1289594	100.4	-
Acid	1021542	79.5	20.9
Alkali	1025578	79.3	21.1
Peroxide	1066442	80.2	20.2
Reduction	1037231	80.5	20.5
Thermal	1029321	80	20
Photolytic	1031751	80.3	20.7

Table: 4 Data for forced degradation studies of Saxagliptin.

Parameter	Area count	%label claim	% degradation
Control	1232568	100.7	-
Acid	981325	80.7	20
Alkali	969984	80.6	20.1
Peroxide	1037923	80.5	20.2
Reduction	980952	72.6	28.1
Thermal	983006	79.4	21.3
Photolytic	985308	79.8	20.9

Chromatograms for forced Degradation Studies**Fig:5 Acid Degradation Chromatogram for Metformin Hcl & Saxagliptin.**

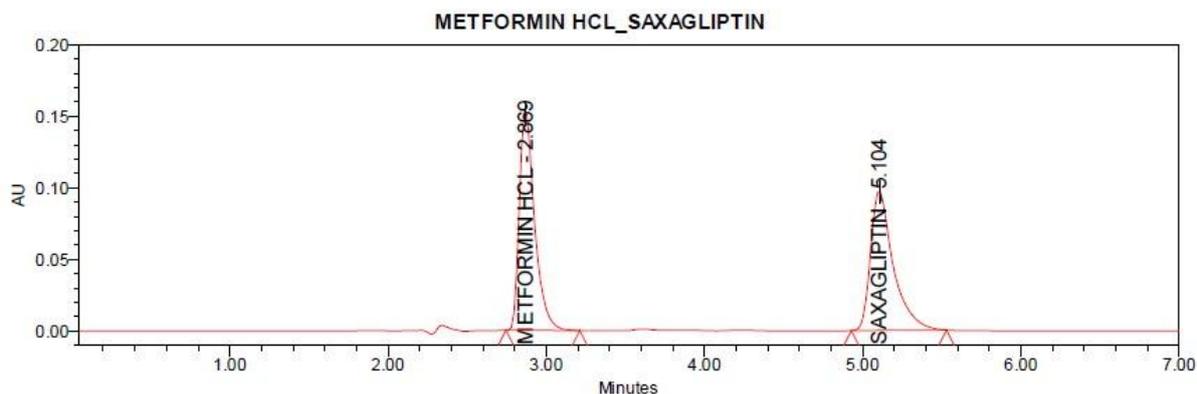


Fig:6 Alkali Degradation Chromatogram for Metformin Hcl & Saxagliptin.

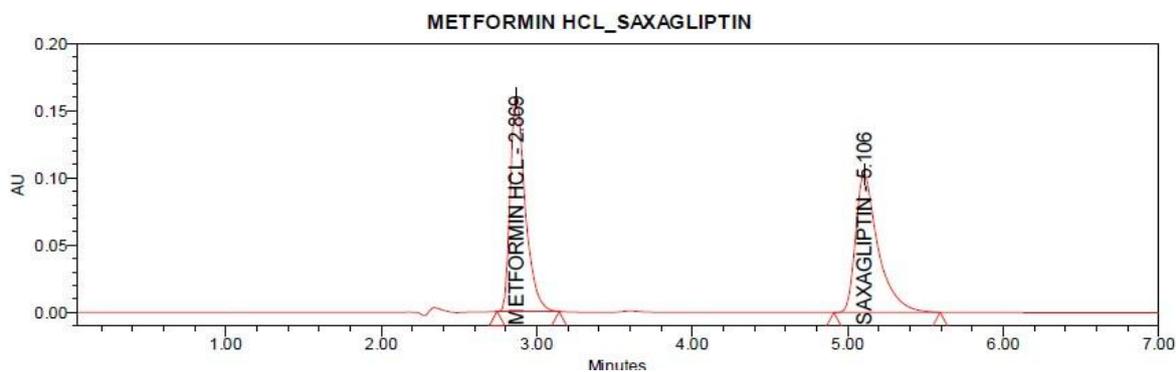


Fig:7 Peroxide Degradation Chromatogram for Metformin Hcl & Saxagliptin.

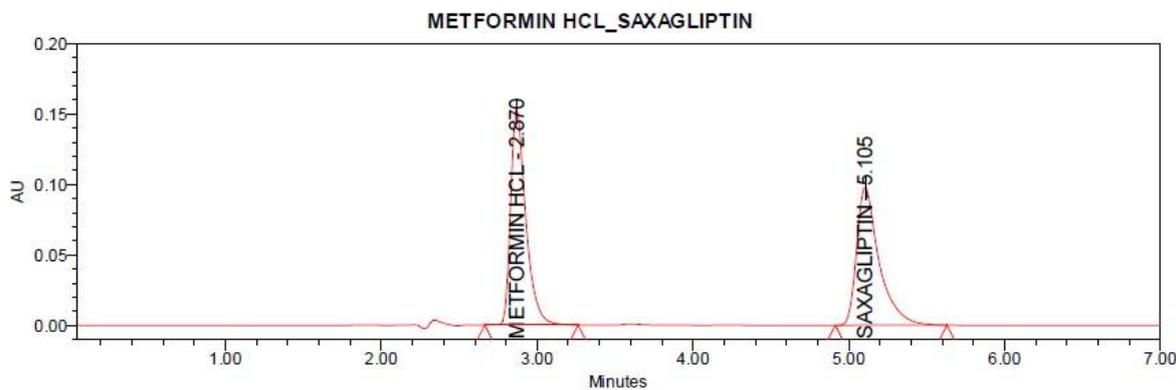


Fig:8 Reduction Degradation Chromatogram for Metformin Hcl & Saxagliptin.

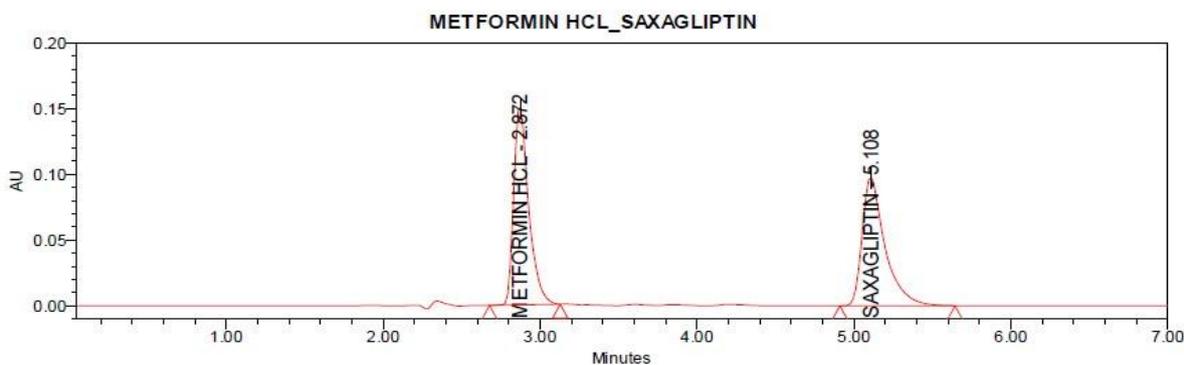


Fig:9 Thermal Degradation Chromatogram for Metformin Hcl & Saxagliptin.

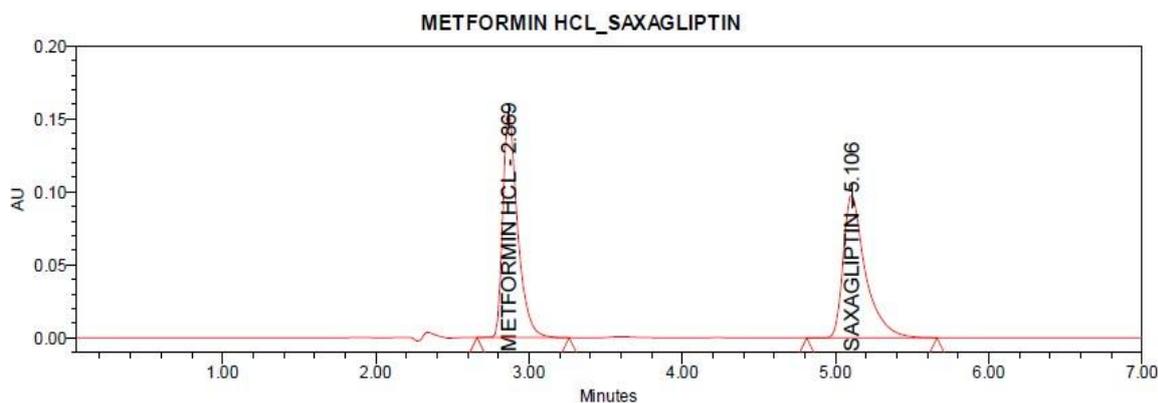


Fig:10 Photolytic Degradation Chromatogram for Metformin Hcl & Saxagliptin.

Linearity

The linearity responses in the concentration range of 80-730 $\mu\text{g/mL}$ for MET and 8-70 $\mu\text{g/mL}$ for SITA was determined. And the co-relation coefficient was NLT 0.99.

Table 5: Linearity of Metformin hydrochloride.

Concentration ($\mu\text{g/ml}$)	Area
50	122309
100	257527
200	495746
261	648447
300	760044
400	972351
500	1240768
600	1512721
740	1835172

Table 6: Linearity of Saxagliptin.

Concentration ($\mu\text{g/ml}$)	Area
0.5	110969
1.0	260032
2.0	477758
2.5	621817
3.0	728045
4.0	933008
5.0	1196868
6.0	1463862
7.5	1772508

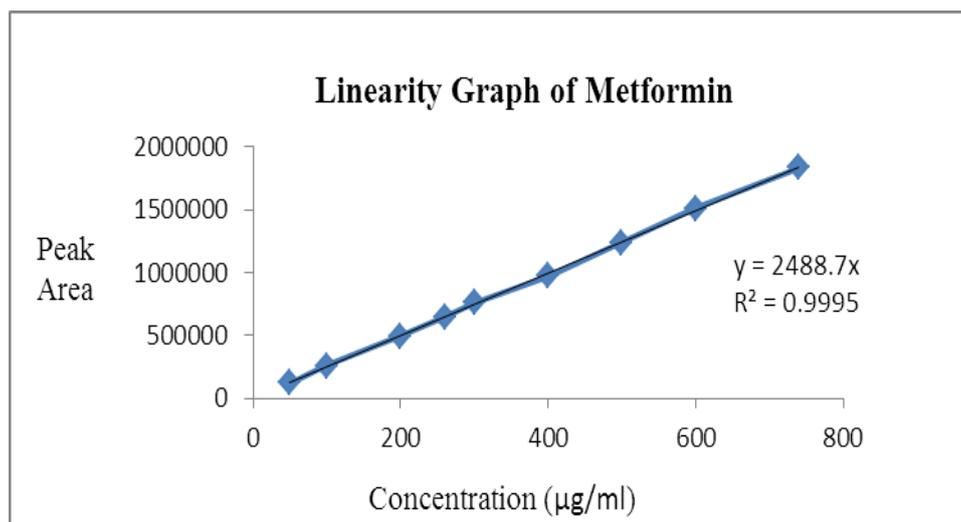


Fig: 11 Linearity graph of Metformin Hcl.

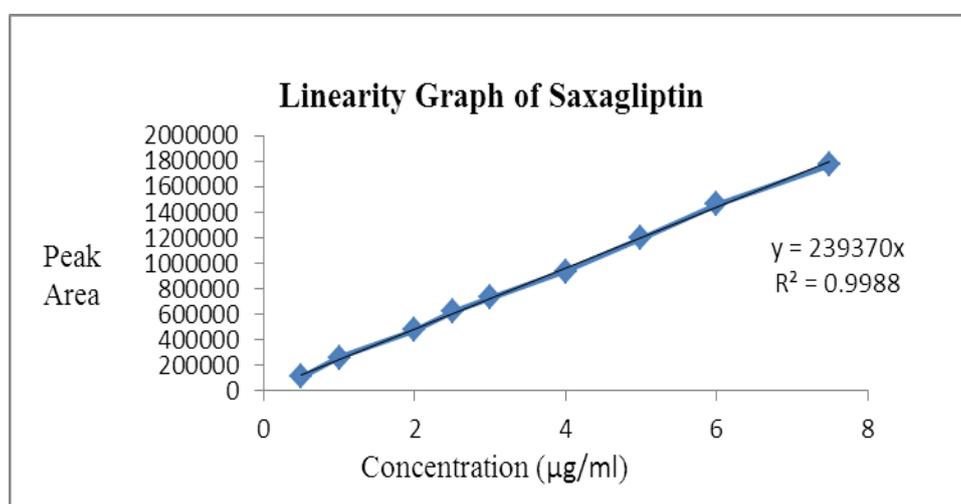


Fig:12 Linearity graph of Saxagliptin.

Precision

Precision was measured in terms of repeatability of application and measurement. Study was carried out by injecting six replicates of the standard at a concentration of 500 $\mu\text{g/mL}$ for MET and 5 $\mu\text{g/mL}$ for SITA. And the RSD calculated from replicates of assay values NMT 2.0%.

Table 7: Precision data of the proposed method.

Drug	Concentration added, $\mu\text{g mL}$	Intra-day precision		Inter-day precision	
		Mean amount found, $\mu\text{g mL}$	% RSD (n = 6)	Mean amount found, $\mu\text{g mL-1}$	% RSD (n = 6)
Metformin hydrochloride	500	499.66 \pm 0.72	0.32	499.81 \pm 0.27	0.52
Saxagliptin	5	4.9.25 \pm 0.54	0.24	4.87 \pm 0.53	0.38

Accuracy

Accuracy (Recovery) of the method was determined by spiking 50, 100 and 150% of working standard at a concentration of 500 μ g/mL for MET and 5 μ g/mL for SAXA. Samples were injected in triplicate across its range according to the assay procedure. The RSD calculated from replicates of assay values NMT 2.0% and the percentage recovery was in between 99% to 102%.

Recovery

S.NO	Level	% Recovery for Metformin hydrochloride	%Recovery for Saxagliptin
1	50%	99.8	99.6
2	100%	100.1	98.9
3	150%	99.5	99.6

Detection and quantitation limits

The LOD and LOQ values were determined by the formulae $LOD = 3.3 s / m$ and $LOQ = 10 s/m$ (Where, *s* is the standard deviation of the responses and *m* is mean of the slopes of the calibration curves).

Robustness

Robustness of the method was determined by making slight changes in the chromatographic conditions, such as flow rate (1 ± 0.1 mL/min), wavelength (± 1 nm), organic phase ($\pm 10\%$) and pH (± 0.2)

RESULT AND DISCUSSION

A reversed-phase column procedure was proposed as a suitable method for the simultaneous determination of Saxagliptin and Metformin hydrochloride in combined dosage forms. The chromatographic conditions were optimized by changing the mobile phase composition, pH, and buffers used in the mobile phase. Different ratios were experimented to optimize the mobile phase. Finally a mixture of P^H2.5 buffer & Acetonitrile in the ratio of 70:30 v/v was used. A typical chromatogram obtained by using the above mentioned mobile phase from 10 μ l of the assay preparation is illustrated below. The retention times of Saxagliptin and Metformin hydrochloride were 5.1 and 2.8 min, respectively. The linearity of the method was tested from 0.5-7.5 μ g/ml for Saxagliptin and 50-750 μ g/ml for Metformin hydrochloride. Correlation coefficients for the regression line were 0.9965 and 0.9998 for Saxagliptin and Metformin hydrochloride

respectively. The accuracy of the method was studied by recovery experiments. The recovery was determined at three levels, viz. 50%, 100%, and 150% of the selected concentrations. Three samples were prepared for each recovery level. The recovery values for Saxagliptin and Metformin hydrochloride ranged from 99.5- 100.4%, respectively. The precision of the method was determined from one lot of combined dosage form. To determine the robustness of the developed method experimental conditions were purposely altered and RSD of the peak areas of Saxagliptin and Metformin hydrochloride were found not greater than 2.0 illustrates the robustness of the method.

CONCLUSION

A simple specific stability-indicating HPLC method has been developed for the quantification of Metformin Hydrochloride and Saxagliptin. This method has been validated and found to be specific, precise, accurate, linear, robust, and linear for the detection and quantification of Metformin Hydrochloride and Saxagliptin. This method exhibited an excellent performance in terms of sensitivity and speed and it helps in simultaneous estimation of Metformin Hydrochloride and Saxagliptin in pharmaceuticals i.e., in combination drugs. This method is suitable for routine analysis and quality control of pharmaceuticals.

ACKNOWLEDGEMENTS

The authors are thankful to the management of Acharya Nagarjuna University for providing the essential facilities for successful completion of the Research work.

REFERENCES

Journal References

1. Arayne MS, Najma S, Zuberi MH, Siddiqui FA. Spectrophotometric Quantitation of Metformin in bulk drug & Pharmaceutical formulation using multivariate technique. *Indian J Pharm Sci*, 2009; 71: 331-35.
2. Mousumi K, Choudary PK. HPLC method for estimation of Metformin hydrochloride in formulated microspheres & tablet dosage form. *Indian J Pharm Sci*, 2009; 2: 318-20.
3. Kalaichelvi R, Jayachandran E, Validated Spectroscopic method for the estimation of Saxagliptinin pure and from tablet formulation, *Int J Pharm Pharm Sci*, 2011; 3: 179-180.
4. Srikanth Inturi, Ravikanth Inturi, Israel kumar Tagaram, validated novel lc determination of saxagliptin in pure bulk and pharmaceutical dosage forms, *ijprd*, november 2011; 3(8).

5. R. Pravin Cumar *et al.*, Validated RP-HPLC For Simultaneous Estimation Of Saxagliptin And Metformin Hydrochloride In Tablet Dosage Form, *Rasayan J.Chem.*, April-June 2012; 5(2): -137-141,
6. Nyola Narendra *et al.*, RP-HPLC Method Development and Validation for simultaneous estimation of Saxagliptin and Metformin HCl in Tablet Dosage Form. *AJPRHC*, 4(3): 70-77.
7. Aruna Kumari .Bondu *et al.*, Method Development and Validation of Saxagliptin and Metformin Hydrochloride by RP-HPLC in Tablet Dosage Form. *World Journal of Pharmaceutical Research*, 2014; 3(4).
8. Sena Caglar *et al.*, Stability Indicating RP-HPLC Method Development And Validation For Simultaneous Estimation Of Metformin And Saxagliptin In Tablet Dosage Forms. *Analytical & Bioanalytical Techniques Special*, 2014; 12.
9. P.Srisowmya *et al.*, Simultaneous Determination of Saxagliptin Monohydrate and Metformin Hydrochloride in Tablets by HPLC Method *IJP*, Oct-Dec 2014; 3(4).
10. N.V.M.S. Bhagavanji *et al.*, Method Development And Validation of Stability Indicating HPLC Method for Simultaneous Determination Of Saxagliptin And Metformin Hydrochloride In Bulk And Tablet Dosage Form. *Int. Res J Pharm. App Sci.*, 2013; 3(5): 37-42

Books References

11. Government of India, Ministry of health and family welfare, *Indian Pharmacopoeia*, Vol-I & II, The Controller of Publication, New Delhi, 2010; 1657-60.
12. The United States pharmacopoeia, USP 34 NF 29, Vol- III *United States pharmacopoeia convention, Inc*, Rockville, 2011; 3442.
13. ICH, Q2A Text on validation of analytical procedures, international conference on Harmonization, Oct, 1994.
14. ICH, Q3B validation of analytical procedures: methodology, international conference on Harmonization, Nov, 1996.