

**DETERMINATION OF ANTIOXIDANT ACTIVITY, TOTAL PHENOLIC CONTENT IN DIFFERENT PARTS OF MEDICINAL FERN, *BLENCHUM ORIENTALE***

**Hind S. Jasim<sup>1\*</sup>, Nissreen J. Hayawi<sup>2</sup>, Mushrifah Idris<sup>3</sup>, Aminah Abdullah<sup>4</sup>**

<sup>1</sup>Department of Biology, College of Science, Thi-Qar University, Nassiriya, Iraq.

<sup>2</sup>Department of Chemical, College of Science, Thi-Qar University, Nassiriya, Iraq.

<sup>3</sup>Tasik Chini Research Centre, Universiti Kebangsaan Malaysia, 43600 UKM Bangi, Selangor Malaysia.

<sup>4</sup>School of Chemical Sciences and Food Technology, Universiti Kebangsaan Malaysia, 43600 Bangi, Selangor, Malaysia.

Article Received on  
09 Oct 2015,

Revised on 30 Oct 2015,  
Accepted on 19 Nov 2015

**\*Correspondence for**

**Author**

**Hind S. Jasim**

Department of Biology,  
College of Science, Thi-  
Qar University, Nassiriya,  
Iraq.

**ABSTRACT**

Medicinal value of pteridophytes has been known to man for more than 2000 years. *B. orientale* is an edible fern that is used as a food and in traditional medicine. This study assessed the total phenol content (TPC) and antioxidant activity by two different assays (DPPH and FRAP) of the 50% aqueous acetone extracts of *B. orientale* parts sampled from two selected location (UKM Fern Garden and Chini Forest). The extracts analyses were prepared from the young fronds, mature fronds, rhizome and fiddlehead of the sampling fern. ANOVA test at  $P < 0.05$  determines significant differences between various parts extract. It was found the TPC of young frond extract of *B. orientale* had the highest contents (1360 and 1643 mg GAE/100g DW)

in UKM Fern Garden and Chini Forest respectively. While the antioxidant activity of plant extracts by DPPH and FRAP assay value for all parts of *B. orientale* was very high at both locations. It illustrated that there is strong antioxidant activity for all parts of plant, especially for Chini Forest and that correlate to diversity of climatic and geomorphological features. These findings provide scientific evidence to support its traditional medicinal importance which is potentially rich sources of natural antioxidants.

**KEYWORDS:** Medicinal Ferns, Total Phenolic Content, *Blenchum orientale*.

## 1. INTRODUCTION

Over the recent years, the effects of antioxidants on illnesses have been investigated. Antioxidants inhibit and scavenge radicals, thereby protecting humans against infections and degenerative diseases. Antioxidants can be defined as any substrate that significantly delays or prevents oxidation when present at low concentration compared with an oxidizable substrate (lipid, protein, carbohydrate, or DNA) (Halliwell 1999). Numerous studies have also established the relationship between consumption of antioxidant-rich plants and prevention of human diseases (Rathore et al. 2011). A natural antioxidant from medicinal plant is safer for the human body because it is a less harmful alternative to a synthetic antioxidant.

Malaysia is known for its green tropical vegetation and forest, and its diverse nature is believed to possess medicinal values. Ferns have been used by humans as food and medicine since ancient times (Ghosh 2004; Lee & Shin 2010). Ferns are a group of non-flowering plants known as Pteridophytes. The fern species in Malaysia are estimated to be 1136 (Bidin & Jaman 1999). As the nutritional contents of the fern are comparable or even superior to those of some common leafy vegetables and medicines, we explore the potential of ferns as a low-cost functional medicine particularly for developing countries (Chang et al. 2010).

*Blechnum orientale* is an important medicinal fern belonging to the family Blechnaceae, is a widespread fern in Southeast Asian with a rapid growth rate and large aboveground biomass. Figure 1 showed the whole plant. This fern has been used in traditional Chinese, Indian, and Malay medicine, as well as a food source plant since ancient times (Benjamin & Manickam 2007). The fern is used in the treatment of diarrhea and stomach problems (Vasudeva 1999). It is considered a cure for intestinal worms and bladder complaints in India and Polynesia, and as a diaphoretic and aromatic in the Philippines (Dixit & Vohra 1984). The rhizome is used as an anthelmintic in China. The diuretic properties of this fern have been put to good use in treating swellings.

The information and knowledge gained from this study are expected to increase the awareness of using natural antioxidants besides synthetic antioxidants. Despite the varied uses of ferns in traditional medicine, no published report exists on their antioxidant activity. Therefore, the dire need of the hour is to discover or identify medicinal plants, rich in antioxidants. Medicinal ferns can be economic, natural and easily affordable by all the people. Thus, The present study was taken up for explore scientifically the antioxidant

potential and total phenolic content of different parts of *B. orientale* fern that were collected from two different growth location. All the assays were carried out in triplicate and the average value was considered.



**Figure 1: *B. orientale* fern: (a) fiddlehead, (b) whole plant.**

## **2. MATERIALS AND METHODS**

### **2.1 Samples Collection**

The fern samples in this study were randomly collected from two study sites, the first site was from, UKM Fern Garden (Taman Paku Pakis), and the second site was Chini Forest reserve. Healthy and green plants were selected for the collection of plant parts. The plant was identified based on taxonomical parameters and their vernacular names. In the present study, the plants used are similar to those generally used in medical preparations in traditional Malay medicine. Fern samples (three replicate) were collected and stored in polyethylene plastic bags and washed gently with deionized distilled water for approximately 3minutes. Plant sample divided into rhizome, mature fronds, young fronds and fiddlehead. It was oven dried at temperature 40°C for 48 h. The dried sample was then pulverized using a mechanical grinder (sharp EM-II, Malaysia) and passed through a standard 20 mesh size sieve (particle size 0.5 mm) The homogenized oven-dried ground material was kept in air-tight container labeled and stored at 4°C until required for analyses.

### **2.2 Extraction of Antioxidant**

The fine powder of fern parts was extracted using 50% of aqueous acetone. Acetone-water solvent was a good solvent to extraction of less polar phenolics. About one gram of each sample (mature frond, young fronds, rhizome and fiddlehead) were weighed in universal bottles and 10 ml solvent was added at room temperature and then swirled with a magnetic

stirrer at a speed of 1,000 rpm for 30 min. All extracted samples were centrifuged using a table top centrifuge (MLX 210, Thermo-line, China) at 4750 g for 15 min. All extractions were carried out in three replicates. The supernatant was collected and kept in  $-20^{\circ}\text{C}$  until the experiment commenced.

### 2.3 Total Phenol Content (TPC) Assay

Total phenolic concentration in the different plant parts extract were quantified by Folin-Ciocalteu procedure which is considered as one of the best methods for the determination of TPC. According to (Musa et al. 2011), about  $10\mu\text{L}$  parts extract of *B. orientale* a part extracts were added to 0.5 ml diluted Folin-Ciocalteu reagent. The samples (Ferns extracts with Folin-Ciocalteu reagent) were left for 5 min before 1 ml 7.5% sodium carbonate (w/v) was added. The absorbance were taken at 765 nm wavelength with a spectrophotometer (Epoch, Biotek, USA), after 2 hours in the dark place. These data were used to estimate the total phenolic content using a standard calibration curve prepared by plotting absorbance against five points concentration: 20, 40, 60, 80, 100 ppm of Gallic acid in methanol to estimate the activity capacity of samples. The result was means of three reading expressed as mg of Gallic acid equivalents per 100 g of dry sample (mg GA/100 g of DW).

### 2.4 DPPH Radical Scavenging Activity (DPPH) Assay

The free radical scavenging activities by antioxidant in the plant extracts were evaluated using DPPH (2,2-diphenyl-1-picrylhydrazyl) radicals following the procedure described by (Brand-Williams et al. 1995), with slight modification. This method widely used to predict the ability of compounds to act as free radical scavengers or hydrogen donors to radicals is based on the free radical 2,2-diphenyl-1-picrylhydrazyl (DPPH•) (Cos et al. 2002).

A stock solution was prepared by dissolving 40 mg DPPH in 100 ml methanol and kept at  $20^{\circ}\text{C}$  until used. About 350 ml stock solution was mixed with 350 ml methanol to obtain the absorbance of  $0.70\pm 0.01$  unit at 517 nm wavelength by using spectrophotometer (Epoch, Biotek, USA). About  $10\mu\text{L}$  of *B. orientale* extracts with 1 ml methanolic DPPH solution prepared were kept 30 min for scavenging reaction in the dark. The difference in absorbance between the test sample and control (DPPH) expressed as percentage inhibition is taken as antioxidant activity AOA was determined as follows:

$$\text{Inhibition (\%)} = \frac{A_{\text{blank}} - A_{\text{sample}}}{A_{\text{blank}}} \times 100.$$

Where, A is the absorbance. All determinations are performed in triplicate.

### 2.5 Ferric Reducing Antioxidant Power (FRAP) Assay

The determination of antioxidant activity through FRAP was carried out according to the method of (Musa et al. 2011) was followed with slight modification. FRAP reagent was prepared fresh as using 300 mM acetatebuffer, pH 3.6 (3.1 g sodium acetate trihydrate, plus 16 ml glacial acid made up to 1:1 with distilled water); 10 mM TPTZ (2,4,6-tris (2-pyridyl)-s-triazine), in 40 mM HCL; and 20 mM FeCl<sub>3</sub>•6H<sub>2</sub>O mixed in a ratio of 10:1:1 and then incubated at 37°C for 10 minutes prior to the analysis to give the working reagent. About 1 mL FRAP reagent was added to 10µL parts extract of *B. orientale*. After adding FRAP reagent, plates were incubated for 30 minutes at room temperature. The absorbance of blue complex (ferrous tripyridyltriazine), formed from the reaction was measured at 595 nm wavelength with spectrophotometer. The antioxidant activity in this study was expressed as milligram Trolox equivalents per hundred grams of plant material on dry basis (mgTE/100g). Linear standard calibration curve of Trolox ranging from 0-100 mM Trolox was set up to estimate the activity capacity of samples. The higher absorbance of the reaction mixture indicates higher reductive potential.

### 2.6 Statistical Analysis

All data were expressed as mean ± standard deviation and were done in triplicate independent analyses. Data were analyzed using one-way ANOVA using SPSS version 20 (SPSS Inc., Chicago, Illinois, USA). analysis of variance (ANOVA) followed by Duncan's test for comparison, as a post hoc test to analyze the different parts of the plant while independent samples.

## 3 RESULTS AND DISCUSSION

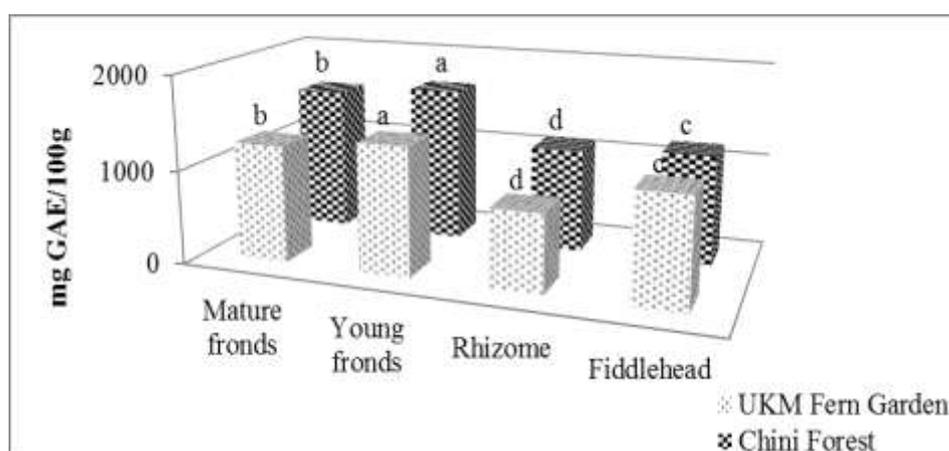
### 3.1 Total Phenolic Content (TPC)

The plants were initially characterized by the amount of phenolic compound they contained. Results are calculated as Gallic acid equivalent (GAE) of a sample using a standard curve. A linear calibration curve was constructed within the range of 20–100mg/ml with an R<sup>2</sup> value of 0.9963. The contents of phenolic compounds in 50% aqueous acetone extracts of mature fronds, young fronds, rhizomes and fiddleheads of the wild fern, *B. orientale* are shown in figure 2. The young fronds of *B. orientale* contained higher values of TPC than the plant's mature fronds, rhizomes, and fiddleheads. The young fronds of *B. orientale* from Chini Forest and UKM Fern Garden exhibit maximum TPC values (1643 and 1360 mg GAE/100g DW), followed by the mature frond (1545 and 1244mg GAE/100g DW), fiddlehead (1170

and 1152 mg GAE/100g DW) and rhizome had the lowest content (1098 and 825.53 mg GAE/100g DW) respectively, which is comparable with the TPC obtained in ethanolic extracts of other wild edible plants, such as *Conyza sumatrensis* (Stem and leaves) possess 1566, *Artemisia dubia* (Stem and leaves) 1424, *Dolichandroneserrulata* (Flower) 1325 and *Vacciniumsprengelii* (Leaves) 954.20 mg of GAE/100g DW which were reported by (Phomkaivon & Areekul 2009).

Interestingly, comparison among different parts of *B. orientale* showed their different phenolic contents. This finding could be attributed to the variation in the nature and distribution of phenol contents within different parts of the same plant. Previous studies showed that the developmental stage of a plant may affect the biosynthesis pathways of phenolic compounds, thereby affecting the total phenolic and flavonoid contents (Križman *et al.* 2007). The plants sampled from the remote area of Chini Forest demonstrated higher TPC than those from the urban area of UKM Fern Garden. Differences in growing conditions, such as climate and soil state, increase in organic matter, rainfall rate, and topography of the land in the two different locations significantly affected bioactive compounds and could be cause of the significant differences between the TPC values of the investigated fern.

The results strongly suggest that phenolics are very important components of the plant and some of its pharmacological effects could be attributed to the presence of its valuable constituents. The results from this study showed that the constituents of this plant are having antioxidant and pharmacological effects.



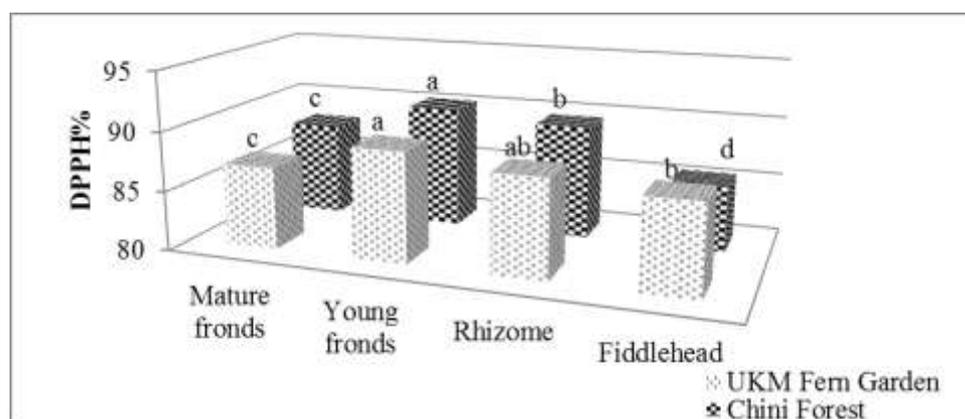
**Figure 2: Total phenolic content of different parts of *B. orientale* from Chini Forest and UKM Fern Garden in 50% aqueous acetone (Mean value, n=3).**

Note: Means within samples with different alphabet letters are significantly different

### 3.2 Antioxidant Substances

#### 3.2.1 DPPH Radical Scavenging Activity (DPPH)

DPPH is a stable nitrogen-centered free radical compound that is widely used to assess the free radical scavenging activity ability of various chemicals, including single compounds, food, and plant extracts (Yamaguchi *et al.* 1998). Figure 3 shows the scavenging effect of *B. orientale* extracts of different parts have high values for all samples at both locations. These values of radical scavenging activities in *B. orientale* extracts collected from Chini Forest were ranked as; young fronds > rhizome > mature fronds > fiddlehead with 90.24%, 89.56%, 87.88% and 85.52%, respectively. The young fronds of *B. orientale* in UKM Fern Garden obtained the highest scavenging activity (89.22%) followed closely by rhizome (88.21%) with no significant difference between them, and fiddlehead with no significant difference with rhizome (87.54%) and mature frond had the lowest (86.87%). This is consistent with the result by (Naik *et al.* 2013) reported that the methanol extract of *B. orientale* showed good antioxidant properties against DPPH with (80%) and this provides an empirical scientific evidence to support its traditional medicinal importance. The present study found that the DPPH value of the plant sampled from Chini Forest was significantly higher ( $p < 0.05$ ) than that sampled from UKM Fern Garden. The high radical-scavenging activity of *B. orientale* extracts may be related to the high amounts of phenolic compounds as the plant sample from Chini Forest contained higher phenolic content than the plant from UKM Fern Garden.



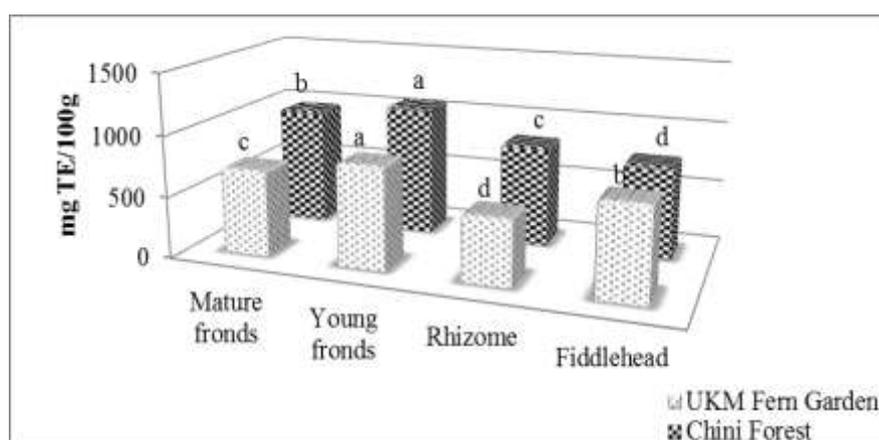
**Figure 3: DPPH of different parts of *B. orientale* from Chini Forest and UKM Fern Garden in 50% aqueous acetone (Mean value, n=3).**

Note: Means within samples with different alphabet letters are significantly different, means with same alphabet letters within each location are not significantly different ( $p > 0.05$ ).

### 3.2.2 Ferric Reducing Antioxidant Power (FRAP)

The FRAP assay was originally developed by (Benzie & Strain 1996), to measure reducing power in plasma, but the assay has also been adapted and used to assess the “antioxidant power” of food and biological samples and plant extracts by its ability to reduce ferric ions (Pellegrini *et al.* 2003). In the present study, we evaluated the ferric reducing potential of *B. orientale* plant parts, expressed as Trolox equivalent capacity (TE). Analysis of the different parts extracts showed that the plant extracts exhibited a significantly different ( $p < 0.05$ ) reducing ability to ferric compounds.

As depicted in figure 4, the ferric compound was significantly reduced by *B. orientale* sampled from Chini Forest, as the Ferric reducing abilities recorded were 1,077 mg TE/100g DW (young fronds), 978.13 mg TE/100g DW (mature frond), 836.82 mg TE/100g DW (rhizome) and 763.56 mg TE/100g DW (fiddlehead). Similarly, *B. orientale* sampled from UKM Fern Garden showed 838.91 mg TE/100g DW (young fronds), 758.17 mg TE/100g DW (fiddlehead), 707.28 mg TE/100g DW (mature fronds) and 537.71 mg TE/100g DW (rhizome). These values are in agreement with the results obtained in a previous study (Lai *et al.* 2009), in which the *B. orientale* contained (1,098 mg GAE/100g) in methanol extract. furthermore, (Naik *et al.* 2013) showed that the frond of *B. orientale* presented 534.05 mg equivalent of ascorbic acid at dry weight in methanol extract. The origin of the plants collected was significantly affected ( $p < 0.01$ ) FRAP value in both plants.



**Figure 4: FRAP value of different parts of *B. orientale* from Chini Forest and UKM Fern Garden in 50% aqueous acetone (Mean value, n=3).**

Note: Means within samples with different alphabet letters are significantly different, means with same alphabet letters within each location are not significantly different ( $p > 0.05$ )

## 1. CONCLUSION

This study has provided an empirical evidence of the role of DPPH, FRAP and TPC assays in providing essentially identical information in regard to the antioxidant capability of fern extracts. The results obtained demonstrated that young fronds of *B.orientale* had the highest total phenolic content and antioxidant activity compared to other parts but all the different parts of ferns contained high antioxidants that are beneficial to the human health. The TPC and total antioxidant content of different parts of *B. orientale* were significantly affected at the locations where the plant was growing. The plant collected from Chini Forest had higher TPC and antioxidant activity compared with the plant collected UKM Fern Garden, and this could be related to the different growing conditions such as soil, geographical and environmental conditions.

## REFERENCES

1. Benjamin, A. & V. Manickam 2007. Medicinal pteridophytes from the Western Ghats. Indian Journal of Traditional Knowledge, 6(4): 611-618.
2. Benzie, I. F. & J. Strain 1996. The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": the FRAP assay. Analytical biochemistry, 239(1): 70-76.
3. Bidin, A. A. & R. Jaman 1999. The Pteridophytes of Tawau Hills Park, Sabah. ASEAN Review of Biodiversity and Environmental Conservation (ARBEC).
4. Brand-Williams, W., M. Cuvelier & C. Berset 1995. Use of free radical method to evaluate antioxidant activity. Lebensm Wiss Technology, 28: 25-30.
5. Chang, H.-C., S. K. Gupta & H.-S. Tsay. 2010. Studies on Folk Medicinal Fern: An Example of "Gu-Sui-Bu". Dlm. (pnyt.). Ed. Working with Ferns hlm., 285-304. Springer.
6. Cos, P., P. Rajan, I. Vedernikova, M. Calomme, L. Pieters, A. J. Vlietinck, K. Augustyns, A. Haemers & D. V. Berghe 2002. In vitro antioxidant profile of phenolic acid derivatives. Free radical research, 36(6): 711-716.
7. Dixit, R. D. & J. N. Vohra 1984. dictionary of the Pteridophytes of India.
8. Ghosh, S. 2004. Pteridophytic flora of eastern India.
9. Halliwell, B. 1999. Food-derived antioxidants. Evaluating their importance in food and in vivo. Food science and agricultural chemistry.
10. Križman, M., D. Baričević & M. Prošek 2007. Determination of phenolic compounds in fennel by HPLC and HPLC-MS using a monolithic reversed-phase column. Journal of pharmaceutical and biomedical analysis, 43(2): 481-485.

11. Lai, H. Y., Y. Y. Lim & S. P. Tan 2009. Antioxidative, tyrosinase inhibiting and antibacterial activities of leaf extracts from medicinal ferns. *Biosci Biotechnol Biochem*, 73(6): 1362-6.
12. Lee, C. H. & S. L. Shin. 2010. Functional activities of ferns for human health. *Dlm. (pnyt.)*. Ed. Working with Ferns hlm., 347-359. Springer.
13. Musa, K. H., A. Abdullah, K. Jusoh & V. Subramaniam 2011. Antioxidant activity of pink-flesh guava (*Psidium guajava* L.): effect of extraction techniques and solvents. *Food Analytical Methods*, 4(1): 100-107.
14. Naik, D. J., P. T. Ramappa, K. Maddappa & N. Somalapura<sup>1</sup> 2013. Antioxidant Activities of *Blechnum orientale* L. *international journal of biological and pharmaceutical research*, 4(2): 105-108.
15. Pellegrini, N., M. Serafini, B. Colombi, D. Del Rio, S. Salvatore, M. Bianchi & F. Brighenti 2003. Total antioxidant capacity of plant foods, beverages and oils consumed in Italy assessed by three different in vitro assays. *The Journal of nutrition*, 133(9): 2812-2819.
16. Phomkaivon, N. & V. Areekul 2009. Screening for antioxidant activity in selected Thai wild plants. *Asian Journal of Food and Agro-Industry*, 2(4): 433-440.
17. Rathore, G. S., M. Suthar, A. Pareek & R. Gupta 2011. Nutritional antioxidants: A battle for better health. *Journal of Natural Pharmaceuticals*, 2(1).
18. Vasudeva, S. 1999. Economic importance of pteridophytes. *Indian Fern J.*, 16(1-2): 130-152.
19. Yamaguchi, T., H. Takamura, T. Matoba & J. Terao 1998. HPLC method for evaluation of the free radical-scavenging activity of foods by using 1, 1-diphenyl-2-picrylhydrazyl. *Bioscience, biotechnology and biochemistry*, 62(6): 1201-1204.