

ANTIMICROBIAL ACTIVITY OF MORINGA OLEIFRA**Mukesh Kanesh, H. C. Khatria and Sheeraz Ahmad Waagy***

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Corresponding Author*Sheeraz Ahmad Waagy**Govt. Geetangali College
Bhopal.**ABSTRACT**

The aim of the present study is to investigate the antimicrobial activity and phytochemical analysis of Methonal extract of *Moringa Oleifra* whole plant against the wound isolates. Two different solvents such as ethanol and methanol were used to extract the bioactive compounds from the whole plant of *Moringa Oleifra* and screened for their antimicrobial activity against the isolated wound pathogens under well diffusion method. The maximum antibacterial activity was observed in crude Ethanolic extract of *Moringa Oleifra* against *Pseudomonas aeruginosa*. Qualitative analysis of phytochemical screening reveals

the presence of Flavonoids, Tannin, Alkaloids and Terpenoids.

KEYWORDS: Moringa oleifra, Antimicrobial activity and Phytochemical analysis.

INTRODUCTION

A wound is a disruption in the normal anatomical structure and function of living tissue that can be caused by physical, chemical, Microbiological or immunological injury. Wounds represent a significant burden on the patients and health care professionals worldwide. Current estimates indicate that worldwide nearly 6 million people suffer from chronic wounds (Kumar et al., 2007).

Chronic wounds may even lead to multiple organ failure or death of the patient. Wound infections often contain multiple organisms, including both aerobic and anaerobic gram-positive cocci and gram- negative bacilli and yeast. Organisms like Streptococcus sp, Staphylococcus aureus, Escherichia coli, Pseudomonas aeruginosa, Proteus sp, Klebsiella, Enterobacter, Clostridium, Peptostreptococcus, Fusobacterium and Aeromonas are highly predominant. (Henry, John, 2001). Many antibiotics used to treat wound pathogens but they also cause undesirable side effects. Bacterial resistance to antibiotic is a major therapeutic problem (Russell, 2002). Search towards safety medicines is must, scientists turn towards the herbal medicines to overcome side effects.

According to World Health Organization medicinal plants with various life sustaining constituents would be the best source to obtain a variety of potential, safe and novel drugs. (Natarajan *et al.*, 2003). Bioactive compounds are exploited on a large scale because of their more systemic and no toxic effects. *Moringa Oleifra* L. (apocyanaceae) also known as Vinca Rosea, is native to the Caribbean Basin and has historically been used to treat a wide assortment of diseases. European\ herbalists used the plant for conditions as varied as headache to a folk remedy for diabetes. It has more than 400 known alkaloids, some of which are approved as antineoplastic agents to treat leukemia, Hodgkin's disease, malignant lymphomas, neuroblastoma, rhabdomyosarcoma, Wilms' tumor, and other cancers.

Its vasodilating and memory-enhancing properties have been shown to alleviate vascular dementia and Alzheimer's disease .it also act as a wound healer (Fischhof *et al.*, 1996 and Hindmarch *et al.*, 1991). In our investigation an attempt was made to analyze the antibacterial activity of *Moringa Oleifra* against the clinical wound isolates.

MATERIAL AND METHODS

Plant Materials

Matured leaves of *Moringa Oleifra* were collected from Outside of Vidishi (M.P) and identified by experts of Institute.

Leaves were surface sterilized with 70% ethyl alcohol followed by 0.1% mercuric chloride. *Moringa Oleifra* leaves were shade dried and powdered with mechanical blender. The air dried plants powders (100 g) were successively extracted with water and alcohol. The extracts were dried in vaccum desicator and were stored in a sterile container for further use (Kelmanson *etal*, 2000).

Determination antibacterial activity

Collection of Samples

Hundred Pus samples were collected from the Wound of patients admitted in Bhopal Hospital. Pus samples were collected from wall of an abscess with the help of sterile swab. Samples were transported to the laboratory for microbiological analysis.

Isolation and identification of wound samples

For the isolation of causative agents, the wound samples were inoculated in Blood agar and MacConkey agar. Plates were incubated at 37°C for 24-48 hrs. Colonies were analyzed by physiological and biochemical test for conformation (Koneman*etal.*, 1998).

Agar Well Diffusion Assay

Agar well diffusion method was followed by using Muller-Hinton Agar (MHA). The plates were seeded with 18 hours old culture of the isolates. The organic fractions were dissolved in Dimethyl sulfoxide (DMSO) and sterilized by using sortorius syringe filter of pore size 0.22µm. Various concentrations of the extracts (250µl, 500µl, 750µl and 1000µl) were added into the sterile 8mm diameter well. Incubation was made at 37°C for 24hrs. Antibacterial activity was assayed by measuring the diameter of the inhibition zone formed around the well using standard (Hi-Media) scale. The experiment was repeated thrice and the average values were calculated for antibacterial activity (Perez *et al.*, 1990).

Phytochemical screening

Methanolic extracts were analyzed for the presence of alkaloids, saponins, triterpenes, and/or steroids, flavonoids, and tannins according to standard methods (Harborne, 1973).

Thin layer chromatography Preparation of TLC Plates

30x10 cm glass plates were washed with distilled water followed by smearing with Methonal. After drying the plates were placed on the template in row. The slurry of silica gel G prepared with glass distilled water in the ratio 1:2 (w/v) was poured in the applicator. The glass plates were immediately coated with a layer of silica gel in 500µm thickness. The coated plates were activated at 80°C for 3 hours. Then the plates were stored in a plate chamber for further study. In that study chloroform and methanol (solvent) was used in 96:4 ratio.

Loading of substances

The concentrated plant extract of 2.5 mg was loaded on the TLC plates just above 2 cm from the bottom using a capillary tube. The plates were reserved in a developing jar containing the solvent mixture. After, the solvent front reached approximately 18cm height. The plates were removed and allowed at room temperature for 30 min. Then the plates were also observed under UV light (240 and 300 nm) and recorded the R_f value of fluorescence substances (Anushia *et al.*, 2009).

RESULTS AND DISCUSSION

Antibacterial activity

Different concentrations (250µg to 1000µg) of all the four extracts were recorded. Crude ethanolic fractions of *Moringa Oleifera* were tested against all the isolates. *Pseudomonas aeruginosa* (29mm), were highly sensitive to the ethanol fraction followed by *Staphylococcus aureus* (25mm), *Escherichia coli* (24mm), *Klebsiella pneumoniae* (18mm) and *Streptococcus pyogenes* (15mm). In

case of Crude methanolic extract *Staphylococcus aureus* (25mm), *Klebsiella pneumoniae* (24mm), *Escherichia coli* (21mm), *Pseudomonas aeruginosa* (20mm), *Streptococcus pyogenes* (16mm) also shows their sensitivity. Soxhlet methanolic extract shows highest sensitivity against *Staphylococcus aureus* (16mm) followed by *Escherichia coli* (13mm), *Klebsiella pneumoniae* and *Streptococcus pyogenes* (12mm) *Pseudomonas aeruginosa* (9mm). Under Soxhlet ethanolic extraction sensitivity was recorded in *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Klebsiella pneumoniae*, *Escherichia coli* and *Streptococcus pyogenes* as 22mm, 21mm, 18mm, 15mm and 12mm respectively (Table 1).

Phytochemical analysis

Phytochemical such as Tannin, Flavonoids, Alkaloids, saponin and Terpenoids were reported from *Catharanthus roseus*. Alkaloids and flavonoids found to be present in all the four extracts whereas Terpenoids present in crude ethanol. Tannin shows their presence in Crude methanol and ethanol. Saponin present in soxhlet ethanolic and methanol. Piovan and Fillipini also reported the presence of various alkaloids, viz. Vincristine, Vinblastine, Yohimbine is an alkaloid and another flavonoids hirsutidin in *Moringa Oleifera* (Table 2).

Thin layer chromatography

Crude ethanol (Rf 0.9) and soxhlet ethanol (0.77) shows brown colour spots whereas yellow spots were identified in crude methanol extracts. Olive green (Rf 0.15) spot was observed in soxhlet methanol extract (Table 3).

Table 1: Antibacterial activity of *Moringa Oleifra* against isolated wound Pathogens.

Sr. No	Organisms	Crude methanolic extract (ul)	Concentration of Extracts/Zone of inhibition in mm															
1.	Escherichia coli	300	550	800	1000	250	500	750	1000	250	500	750	1000	250	500	750	1000	
2.	Klebsiella pneumoniae	15	15	19	22	16	19	22	25	6	8	11	14	10	11	13	16	
3.	Pseudomonas aeruginosa	15	16	19	21	17	19	24	30	-	6	8	10	10	16	19	23	
4.	Staphylococcus	15	18	22	26	20	21	24	26	6	8	10	18	9	15	18	22	
5.	Streptococcus pyogenes	9	13	15	17	10	11	13	16	6	8	11	13	-	-	7	13	

Table 2: Photochemical screening of *Moringa Oleifra* extracts.

Sr. No	Phytochemicals	Crude methanolic	Crude ethanol	Soxhlet methanolic	Soxhlet ethanol extract
1.	Allcaloids	+	+	+	+
2.	Flavanoids	+	+	+	+
3.	Steroids	-	-	-	-
4.	Glyeosides	-	-	-	-
5.	Terpanoids	+	+	-	+
6.	Tannins	+	+	-	-
7.	Quinine	-	-	-	-
8.	Coumarin	-	-	-	-
9.	Starch	-	-	-	-
10.	Saponins	-	+	-	+
11.	Phenols	-	-	-	-

Table 3: Rf value of various extracts of *Moringa Oleifra* Leaves.

Sr. No	Extracts	Observation of sports	Rf value
1.	Crude Methanol	Yellow	0.5
2.	Crude Ethanol	Brown	0.9
3.	Soxhlet	Olive green	0.16
4.	Soxhlet Ethanol	Brown	0.78

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