STUDY ON PHYTOCHEMICAL SCREENING, TEST FOR INORGANIC ELEMENTS, QUALITATIVE AND QUANTITATIVE CHARACTERIZATION OF PHYTOCONSTITUENTS, TOTAL PHENOL, TOTAL FLAVONOID CONTENT AND ANTI-OXIDANT POTENTIAL OF METHANOL EXTRACT OF SCINDAPSUS AUREUS LEAVES

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ABSTRACT

The aim of the present study was to identify the antioxidant potential of methanol extract of Scindapsus aureus leaves. The extract was examined for DPPH free radical scavenging activity, reducing power capacity and for phenol and flavonoid content. Scindapsus aureus is plant belonging to family Araceae. To give a scientific basis for usage of this plant, the leaf extract were appreciated for its antioxidant activity. In the present study we estimate the qualitative analysis and the phytochemical compounds such as steroids, carbohydrates, triterpenoids, coumarins, phenolic compounds, flavonoids, saponins, nthroquinones, Proteins and lipids were screened in leaves of Scindapsus aureus. Quantitative analysis of phenols, flavonoids and Triterpenoids was further performed. Preliminary phytochemical screening of leaves were by methanolic extracts which identified 11 major functional groups, amongst them flavonoid, phytosterols, saponins and coumarins. Additionally inorganic elements like iron, chloride and sulphate were identified by total ash analysis. The extract showed a dose dependent radical scavenging effect in DPPH assay. IC50 for free radicals achieved by the extract is 144.72 μg/ml. The extract showed significant reducing power activity as compared to ascorbic acid and proportionally increased with the increasing concentration of the extract. Increase in
absorbance of the reaction mixture indicates the increase in the reducing power of the extract. Total phenolic and total flavonoid content has been present in the extract and the amounts are $16.42 \pm 0.48 \text{ mg Gallic acid/g dry weight}$ and $20.12 \pm 0.13 \text{ mg Quercetin/g dry weight}$ on dried extract respectively.

**KEYWORDS:** *Scindapsus aureus*, phenol, flavonoid, antioxidant, DPPH, lignins, stilbenes, tannins, amines, betalains, flavonoids, quinones, coumarins, alkaloids.

**INTRODUCTION**

Plants are furnished with various phytochemical molecules such as terpenoids, phenolic acids, vitamins, lignins, stilbenes, tannins, amines, betalains, flavonoids, quinones, coumarins, alkaloids, and other metabolites, which are rich in antioxidant activity. (Zheng et al., 2001; Cai et al., 2003) Studies have revealed that lot of these antioxidant compounds have anti-inflammatory, antiatherosclerotic, antitumor, antimutagenic, anticarcinogenic, antibacterial and antiviral activities (Sala et al., 2002; Rice-Evans et al., 1995). The treatment with natural antioxidants has been connected with reduced risks of cancer, cardiovascular disease, diabetes, and other diseases related to ageing. (Ashokkumar, et al., 2008; Veerapur, et al., 2009) and in current years, there has been a worldwide movement towards the exercise of the natural phytochemicals present in oilseeds, beans, fruits berry crops, teas, herbs and vegetables (Kitts, et al., 2000; Muselík, et al., 2007; Wang, et al., 2007). In current years, phytochemicals (secondary metabolites) with unknown pharmacological activities have been comprehensively studied as a source of therapeutic agents (Krishnaraju et al., 2005). Since there is a hurdle in use of traditional medicines worldwide due to lack of quality and quantity safety and efficacy information on traditional medicines. The lack of research data are not only due to lack of methodologies for the evaluation of herbal medicines but also due to health policies. The plant contains lots of active chemicals and therapeutically constituents. Hence in modern systems of medicine it important to study quality control of herbal medicines (WHO). General Guidelines for Methodologies on Research and Evaluation of Traditional Medicines; 2001) herbal medicine is compulsory (Chaudhury., 1999; Kokate, et al., 2005; Raina, et al., 2003; Raven, et al., 1999; Ran, et al., 1999).

An expanding human population, global environment change and the change of terrestrial food resources for energy needs in recent times have elevated serious global food security concerns (Kumar et al., 2011). There has been a quest to explore and use foods from diverse sources to enhance and supplement the nutritional quality of human foods. Non timber forest
products used in diet have recently gotten an increasing interest as they constitute a developing source of food. Organic and natural compounds have been reported to have own antioxidant properties, bioactivities and applications of preparations isolated from herb, flower, vegetable, which most frequently include berries, fruits, fresh fruit and vegetables, medicinal, aromatic plants, herbs and other botanicals have been well documented (Biapa et al., 2011; Choumessi et al., 2012; Dimo et al., 2001). Polyphenols are bioactive compounds extensively spread in plants and they are generally also significant constituents of the human diet (Pauline et al., 2013). Plants are considered as the key sources of antioxidants, which constitute a rich diversity of substances such as flavonoids (anthocyanins, flavonols, flavones) as well as some classes of non-flavonoids (phenolic stomach acids, lignins, stilbenes, terpenoids and many others). These compounds vary in structure, the quantity of phenolic hydroxyl groups and the location, leading to variance in their antioxidative and biological potential (Erkan et al., 2011). Some culinary herbs and spices have been proven to be more effective antioxidants than common food additives (butylated hydroxyanisole, butylated hydroxytoluene and propyl gallate) and vitamins (ascorbic acid) (Suhaj, 2006). Therefore herbs and spices rich in anti-oxidants and other phyto-compounds are able to prevent oxidative stress and its related disorders such as persistent diseases (Soory, 2009; Tapsell et al., 2006).

Hence, investigations in naturally occurring antioxidants has considerably increased and natural products have regained prominence in the recent past with increasing understanding of their biological significance such as antioxidant, radical scavenging activities and increasing recognition of the origin and function of their structural diversity (Deng et al., 2006; Gul et al., 2011; Hogan et al., 2010; Zhang et al., 2007). It then becomes necessary to search new source for noble antioxidants, especially those that would be safe and cheap and thus easily affordable by all population.

In this study, in vitro antioxidant activities of the leaf extract of *Scindapsus aureus* were determined by qualitative, quantitative analysis, total antioxidant, DPPH, Reducing power capacity, total phenolic content and total flavonoid content.

2. MATERIAL AND METHOD

2.1 Plant material

Fresh leaves of *Scindapsus aureus* were collected at Lal-bagh.
2.2 Preparation of Extract

The leaves were dried for a period of 10 days under shade and ground. The ground leaves (450 gm) were soaked in sufficient amount of methanol for one week at room temperature with occasional shaking and stirring then the whole mixture was filtered and the filtrate thus obtained was concentrated using a water bath to get a viscous mass. The viscous mass was kept at room temperature under a ceiling fan to get a dried extract (yield value, 5.3%). The extract prepared was for pharmacological screening.

2.3 Qualitative phytochemical analysis

Qualitative phytochemical analysis of methanol extracts of leaves was conducted using following standard procedures. (Khandelwal et al., 2006) Elemental analysis of Ash for detection of inorganic elements (Calcium, Iron, Magnesium, Potassium, Sulphate, phosphate, chloride, carbonate and Nitrate) was performed by specific tests.(Khadbadi et al., 2006).

2.4 Quantitative phytochemical analysis

The phytochemicals are present in the methanol extract of leaves and were determined by standard procedures.(Khadbadi et al., 2006).

2.5 In vitro Antioxidant Activity

2.5.1 DPPH free radical scavenging activity

DPPH scavenging activity was carried out using the method of Braca et al., 2001. Different concentrations (400, 200, 100, 50, 25 and 12.5 μg/mL) of Scindapsus aureus extract were dissolved in methanol and placed in different test tubes, and 3 mL of a 0.004% w/v methanol solution of DPPH was added to each test tube. Absorbance at 517 nm was determined after 30 min against a blank and the percent inhibition activity was calculated from \[ \frac{(A_0 - A_1)}{A_0} \times 100 \], where \( A_0 \) is the absorbance of the control and \( A_1 \) is the absorbance of the sample. Ascorbic acid was used as a reference standard and dissolved in methanol to make the stock solution with the same concentration. The control sample was prepared containing the same volume without any extract or reference drug. Methanol served as a blank. The inhibition curves were prepared and the half maximal inhibitory concentration (IC50) values were calculated using linear regression analysis.

2.5.2 Reducing power capacity

The reducing power of the extract was evaluated by the established method described by Oyaizu (Oyaizu, 1986) with slight modification. Different concentrations of leaf extract of \( B. \)
platyphylla (125, 250, 500 and 1000 μg/mL) in 1 mL of distilled water were mixed with phosphate buffer (2.5 mL, 0.2 M, pH = 6.6) and potassium ferricyanide \([K_3Fe(CN)₆]\) (2.5 mL, 1% w/v). The mixture was incubated at 50°C for 20 min. After incubation, 2.5 mL of 10% trichloroacetic acid solution was added to each tube and the mixture was centrifuged at 3000 rpm for 10 min. Subsequently, 5 mL of the upper layer solution was mixed with 5 mL of distilled water and 1 mL of ferric chloride solution (0.1% w/v) and the absorbance was measured at 700 nm. The reducing power of the extract was linearly proportional to the concentration of the sample. Ascorbic acid was taken as a reference standard. Phosphate buffer (pH 6.6) was used as a blank solution.

2.5.3 Determination of total phenolic content

Total phenolic content of the extract was evaluated with Folin-Ciocalteu method (Uddin et al., 2015). Samples containing polyphenols are reduced by the Folin-Ciocalteu reagent thereby producing blue colored complex. The phenolic concentration of extracts was evaluated from a gallic acid calibration curve. To prepare a calibration curve, 0.5 mL aliquots of 12.5, 25, 50, 100, 200 and 400 μg/mL methanolic gallic acid solutions were mixed with 2.5 ml Folin-Ciocalteu reagent (diluted ten-fold) and 2.5 ml (75 g/l) sodium carbonate. After incubation at 25°C for 30 min, the quantitative phenolic estimation was performed at 765 nm against reagent blank by UV Spectrophotometer 1650. The calibration curve was constructed by plotting the value of absorbance vs. concentration. A similar procedure was adopted for the extract as above described in the preparation of calibration curve. All determinations were performed in triplicate. Total phenolic content was expressed as milligrams of gallic acid equivalent (GAE) per g of extract.

2.5.4 Determination of total flavonoid content

Total flavonoid content of ethanol extract was evaluated with method of Jiao (Jiao & Wang, 2000). One mL of Scindapsus aureus extract or standard of different concentrations was taken in a test tube and 3 mL of methanol was added. Then 200 μl of 10% aluminium chloride solution was added into the same test tube followed by the addition of 200 μl of 1M potassium acetate. Finally, 5.6 mL of distilled water was mixed with the reaction mixture. The reaction mixture was then incubated for 30 min at room temperature to complete the reaction. Then the absorbance of the solution was measured at 415 nm using a spectra photometer against blank. Methanol served as blank. The Total content of flavonoid compounds in Scindapsus aureus was expressed in mg/g quercetin equivalent (QE).
2.6 STATISTICAL ANALYSIS

All results are expressed as mean ± standard error of the mean (SEM). The results were statistically analyzed using repeated measures analysis of variance with Dunnett’s multiple comparison when compared against negative control in all in vivo model of Sedative and Anxiolytic activities. P<0.05, P<0.01 and P<0.001 were considered as statistically significant. Statistical programs used were SPSS (Statistical Package for Social Science, version 22.0, IBM Corporation, NY). GRAPHPAD PRISM® (version 6.00; Graphpad Software Inc., San Diego, CA, USA) was used for graphical presentation.

3. RESULTS

Phytochemical investigation showed the presence of bioactive compounds like Cardioglycosides, Flavonoid, Quinones, Terpenoids, Alkaloids and Steroids and inorganic elements like magnesium, iron, sulphate, phosphate, chloride and fluoride were reported in leaves part. While leaves of plant shows the presence of carbohydrate, protein, saponin, coumarin and flavonoid and inorganic elements like iron, sulphate and chloride. Hence leaves of plants can also be used as source of medicine for treatment of many diseases.

3.1 Qualitative phytochemical analysis

Leaf extracts of Scindapsus aureus was carried out by using various solvents of increasing polarity and then the methanolic extract was subjected to preliminary phytochemical screening for the identification of active major functional groups. Additionally the ash of powdered pods was utilized for the detection of inorganic elements, which shows the medicinal importance of the plant. The results are given in Table 01 and 02.

Table 01: Showing results of phytochemical analysis Phytochemical analysis.

<table>
<thead>
<tr>
<th>Sl. No.</th>
<th>Test</th>
<th>Inference</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Carbohydrate</td>
<td>+</td>
</tr>
<tr>
<td>2</td>
<td>Protein</td>
<td>+</td>
</tr>
<tr>
<td>3</td>
<td>Glycoside</td>
<td>-</td>
</tr>
<tr>
<td>4</td>
<td>Saponin</td>
<td>+</td>
</tr>
<tr>
<td>5</td>
<td>Coumarin</td>
<td>+</td>
</tr>
<tr>
<td>6</td>
<td>Flavonoid</td>
<td>+</td>
</tr>
<tr>
<td>7</td>
<td>Anthraquinone glycoside</td>
<td>-</td>
</tr>
<tr>
<td>8</td>
<td>Phytosterol</td>
<td>-</td>
</tr>
<tr>
<td>9</td>
<td>Phenol</td>
<td>-</td>
</tr>
<tr>
<td>10</td>
<td>Alkaloids</td>
<td>-</td>
</tr>
<tr>
<td>11</td>
<td>Lipid</td>
<td>+</td>
</tr>
</tbody>
</table>

(+): present  (-): absent.
Table 02: Detection of inorganic elements. Phytochemical analysis.

<table>
<thead>
<tr>
<th>Sl. No.</th>
<th>Test</th>
<th>Inference</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Calcium</td>
<td>+</td>
</tr>
<tr>
<td>2</td>
<td>Iron</td>
<td>+</td>
</tr>
<tr>
<td>3</td>
<td>Magnesium</td>
<td>+</td>
</tr>
<tr>
<td>4</td>
<td>Potassium</td>
<td>+</td>
</tr>
<tr>
<td>5</td>
<td>Sulphate</td>
<td>-</td>
</tr>
<tr>
<td>6</td>
<td>Phosphate</td>
<td>+</td>
</tr>
<tr>
<td>7</td>
<td>Chloride</td>
<td>+</td>
</tr>
<tr>
<td>8</td>
<td>Carbonate</td>
<td>+</td>
</tr>
<tr>
<td>9</td>
<td>Nitrate</td>
<td>+</td>
</tr>
</tbody>
</table>

(+) present (-) absent.

3.2. In Vitro Antioxidant Activity

3.2.1. DPPH radical scavenging activity

Results for the free radical scavenging activity of methanol extract of *Scindapsus aureus* are shown in Figure 1. The extract showed a dose dependent radical scavenging effect in DPPH assay. The half inhibition concentration (IC50) for free radicals achieved by the extract is 144.72 μg/ml which is statistically significant compared to that (IC50 8 μg/ml) of reference antioxidative agent ascorbic acid.

![Figure 1: DPPH radical scavenging activity of Scindapsus aureus leaves.](image)

Ascorbic acid.

Methanol extract of *Scindapsus aureus*.

3.2.2. Reducing power capacity

The extract showed significant reducing power activities as compared to ascorbic acid and proportionally increased with the increasing concentration of the extract, which is shown in Figure 2. Increase in absorbance of the reaction mixture indicates the increase in the reducing power of the sample.
Ascorbic acid.

Methanol extract of *Scindapsus aureus*.

Figure 2: Reducing capacity of the methanol extract of *Scindapsus aureus* leaf.

3.2.3. Quantitative determination of phytochemical contents

Data for total phenolic and total flavonoid content has been summarized in Table 1. Data shows that moderate amount of total phenols and total flavonoids are present in the extract and the amounts are 18.44 ± 0.45 mg Gallic acid/g dry weight and 21.12 ± 0.23 mg Quercetin/g dry weight of dried extract respectively.

3.2.3. Quantitative determination of phytochemical contents

Data for total phenolic and total flavonoid content has been summarized in Table 3. Data shows that moderate amount of total phenols and total flavonoids are present in the extract and the amounts are 18.44 ± 0.45 mg Gallic acid/g dry weight and 21.12 ± 0.23 mg Quercetin/g dry weight of dried extract respectively.

<table>
<thead>
<tr>
<th>Phytochemicals (mg/gm)</th>
<th>Scindapsus aureus (methanol)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total Phenol (mg Gallic acid /g)</td>
<td>16.42 ± 0.48</td>
</tr>
<tr>
<td>Total Flavonoid (mg Quercetin/g)</td>
<td>20.12 ± 0.13</td>
</tr>
</tbody>
</table>

Values are the mean of triplicate experiments and represented as mean ± SEM (n=3).

4. DISCUSSIONS

From best of our knowledge, present study was the first make an effort to evaluate the ability of the leaves extract of *Scindapsus aureus* to act as antioxidant agents. The most natural antioxidants are multifunctional. Consequently, a trusted antioxidant analysis process
requires different antioxidant activity assessments to account various mechanisms of antioxidant action. In this study, several techniques have been used to determine the in vitro antioxidant activity to let quick screening of substances.

DPPH radical scavenging model is widely used method to evaluate antioxidant activity of natural compound and plant extracts. The degree of discoloration indicates the scavenging potential of the antioxidant extract, which is due to the hydrogen donating ability (Barreira et al., 2008). The experimental data revealed that methanol extracts of leaves have the effects of scavenging free radicals and a dose dependent relationship in the DPPH radical scavenging activity. The involvement of free radicals, especially their increased production leads to the development of cardiovascular diseases and cancer. Thus, the consumption of *Scindapsus aureus* leaves can be beneficial in preventing oxidative stress related numerous chronic diseases.

The reducing power of MEBP was determined by direct electron donation in the reduction of ferri cyanide \([\text{Fe(CN)}_6]^{3-}\) to ferro cyanide \([\text{Fe(CN)}_6]^{4-}\). The product was visualized by addition of free \(\text{Fe}^{3+}\) ions after the reduction reaction, by forming the intense Prussian blue colour complex, \((\text{Fe}^{3+})_4[\text{Fe}^{2+} (\text{CN})_6]^{3-}\) and quantified by absorbance measurement at 700 nm (Saha et al., 2013). The presence of reductants (i.e. antioxidants) in *Scindapsus aureus* leaves cause the reduction of the \(\text{Fe}^{3+}\) /ferricyanide complex to the ferrous form which was monitored by measuring the formation of Perl’s Prussian blue at 700 nm. Figure 2 shows the reductive capabilities of MEBP compared to ascorbic acid. Therefore, like the DPPH radical scavenging activity, the observed reducing power of leaves and stem bark was in agreement with the chemical constituents in the extracts of *Scindapsus aureus* leaves.

Polyphenols were found in the extracts of leaves. The obtained results for DPPH are in agreement with the phenol contents determined for each sample. Plant polyphenols are produced from phenylalanine or from its precursor shikmic acid. These phenolics are important dietary antioxidants because they have ideal structural chemistry for free radical scavenging activities and have been shown to be more effective antioxidants in vitro than vitamins E and C on a molar basis (Ribeiro et al., 2008). Polyphenols exhibit a wide range of biological effects such as protection of LDL oxidation in vivo with significant consequences in atherosclerosis and also protect DNA from oxidative damage with important consequences in the age-related development of some cancers (Reddy et al., 2012). Our findings suggested
that leaves of *Scindapsus aureus* rich in phenolic and flavonoid contents which are the major contributor to scavenge the free radicals in oxidation pathways.

The results obtained from correlation between polyphenols (phenol and flavonoid) and DPPH scavenging suggested that phenolic compounds are dominant contributors to the antioxidant activity of the extract.

5. CONCLUSIONS
The *Scindapsus aureus* screened for phytochemical constituents seemed to have the potential to act as a source of useful medicines and also to improve the health status of the consumers as a result of the presence of various compounds that are vital for good health. Hence, *Scindapsus aureus* can be utilized in herbal drug formulation.

The present study indicated that *Scindapsus aureus* contains considerable amount of total polyphenols and flavonoids and exhibited good antioxidant activity by effectively scavenging various free radicals. The antioxidant and biological activities might be due to the synergistic actions of bioactive compounds present in them. However, it is still unclear which components are playing vital roles for this activity. Therefore, further studies are still needed to elucidate mechanistic way how the plant contributes to this property.

6. RESULT
The extract showed a dose dependent radical scavenging effect in DPPH assay. IC50 for free radicals achieved by the extract is 144.72 μg/ml. The extract showed proportionally increased with the increasing concentration of the extract. Increase in absorbance of the reaction mixture indicates the increase in the reducing power of the extract. Phenol content was 16.42 ± 0.48 mg gallic acid/g and flavonoid content was 20.12 ± 0.13 mg quercetin/g.

7. CONCLUSION
Our current results emerged that *Scindapsus aureus* act as an antioxidant agent due to its free radical scavenging and cytoprotective activity. So, the plant may be further pursued to find out for its pharmacological active natural products.

8. ACKNOWLEDGMENT
As a prefatory exercise, I owe my heartiest gratitude and unflinching thankfulness to Deenadayalan K, HOD of Biology, Sri Jagadguru Renukacharya Rajajinagar College, Bengaluru - 560 010 for his guidance and planned execution throughout our study.
9. REFERENCES


