

HYDROGEN PEROXIDE INDUCED DNA DAMAGE: PROTECTION BY ANTIOXIDANT COLEUS AROMATICUS

Dinesha Ramadas¹, Sachidananda Gurumahadevaiah², Harsha Ramakrishna³,
Subhas Chandrappa Mundasada^{2*}

¹Adichunchanagiri Biotechnology & Cancer Research Institute, B.G. Nagara

²Shridevi Institute of Medical Science & Research Hospital, Tumkur

³Novozymes R & D, Bangalore

Article Received on
16 May 2014,

Revised on 10 June 2014,
Accepted on 04 July 2014

*Correspondence for
Author

Subhas Chandrappa
Mundasada

Shridevi Institute of Medical
Science & Research Hospital,
Tumkur

ABSTRACT

The antioxidant properties of boiling water extract of *Coleus aromaticus* were evaluated using lipid peroxidation inhibition activity, hydroxyl radicals scavenging activity, 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical scavenging effect and Superoxide radical scavenging activity. The extracts of *Coleus aromaticus* effectively protects H₂O₂ (100µM) induced cell death in human peripheral lymphocytes. Further *Coleus aromaticus* extract prevents H₂O₂ (1mM) mediated calf thymus DNA damage as evidenced by agarose gel electrophoresis. The present study reveals that, the boiling water extract of *Coleus aromaticus* showed significant antioxidant and DNA damage

protectant activity when compared to antioxidants like BHA, Curcumin and α-tocopherol.

KEY WORDS: Antioxidants, *Coleus aromaticus*, Cytotoxicity, DNA damage, H₂O₂.

INTRODUCTION

Reactive oxygen species (ROS) cause wide oxidative damage to cellular biomolecules like DNA, proteins and lipids and they contribute to the oxidative stress-related diseases^[1,2], though several promising synthesized antioxidants available, but their toxicity and side effects avoids their recommendation^[3]. Hence, there is great interest in the use of naturally occurring antioxidants towards inhibition or prevention of oxidative stress-related diseases^[4]. The administration of an antioxidant source comprising of multiple components could offer protection against cancer^[5,6] and combat oxidative stress-induced physiological malfunctions^[7]. According to President of Foundation for innovation in Medicine, if a food or

its part is having medical /health benefits and resulting in the prevention /cure of diseases then it is called as Nutraceutical. *Coleus aromaticus* is a large succulent herb, aromatic, branched with distinctive smelling leaves. The taste of this leaf is pleasant with refreshing odour. The leaves are strongly flavoured and make an excellent addition to meat dishes. It is also used as a vegetable, also as herb in the food trade. It has traditional medicinal properties and hence used for the treatment of coughs, sore throats and nasal congestion, infections, rheumatism and flatulence. In traditional medicine, it is used to treat malarial fever, hepatopathy, renal and vesical calculi, cough, chronic asthma, hiccup, bronchitis, helminthiasis, colic, convulsions, and epilepsy henoy and others^[8,9,10,11].

MATERIALS AND METHODS

Coleus aromaticus leaves obtained from Sasyakashi, maintained by Adichunchanagiri Maha Samsthana Mutt, Bellur cross, Nagamangala Taluk, Mandya District, Karnataka state, India. DPPH (2,2-Diphenyl-2-picrylhydrazyl), Superoxide dismutase (SOD), NBT (Nitro blue tetrazolium), DMSO (Dimethyl sulphoxide), Bovine serum albumin (BSA), Calf thymus DNA, Linolenic acid and Gallic acid from Himedia Private Ltd India. Hydrogen peroxide was purchased from SRL, India. All other chemicals unless otherwise mentioned were of analytic grade and procured from Merck (Darmstadt, Germany). Solvents were distilled before use.

Boiling water extract of *Coleus aromaticus* leaves

Ten gram of the cleaned, fresh selected *Coleus aromaticus* leaves crushed with 100 ml of hot water and boiled for five minutes. It was vortexed for 2 hrs at room temperature. The solution was centrifuged at 6000 rpm for 20mins at room temperature. The supernatant was filtered using Whatman No. 1 filter paper, followed by 0.22m Sartorius microbial filter. Then the supernatant was freeze dried and lyophilized and stored at 4°C for further studies. The efficacy of the extract reported here was quantified based on the dry weight of the whole extract per volume of assay solution.

Proximate analysis

The proximate analysis such as protein^[12], total sugars^[13], polyphenols^[14], flavonoids^[15] was done for the *Coleus aromaticus* extract.

Antioxidant activity

Antioxidant activity of aqueous and boiling water extract of *Coleus aromaticus* was studied in different model system to study their antioxidant activities.

1,1-Diphenyl-2-picrylhydrazyl (DPPH) radical scavenging activity

DPPH radical scavenging activity was assessed according to the method of Shimada et al. 1992^[16]. The α -tocopherol at various concentrations ranging from 0 to 100 M was mixed in 1 ml of freshly prepared 0.5 mM DPPH ethanolic solution and 2 ml of 0.1 M acetate buffer pH 5.5. The mixtures were incubated at 37C for 30 min and measured at 517 nm. α -tocopherol (400 μ M) was used as positive control under the same assay conditions. Negative control was without any inhibitor or extract of *Coleus aromaticus* extract. Lower absorbance at 517 nm represents higher DPPH scavenging activity. The % DPPH radical scavenging activity of *Coleus aromaticus* extract was calculated.

Superoxide scavenging activity by alkaline DMSO method

The Superoxide radical ($O_2^{\cdot-}$) scavenging activity of *Coleus aromaticus* was measured according to the method of Lee et al. 2002^[17]. The reaction mixture containing 100 μ l of 30mM EDTA (pH 7.4), 10 μ l of 30mM hypoxanthine in 50mM NaOH, and 200 μ l of 1.42mM nitro blue tetrazolium (NBT) with or without *Coleus aromaticus* extracts. SOD serving as positive control at various concentrations ranging from 50-300 μ g. After the solution was pre-incubated at ambient temperature for 3min, 100 μ l of xanthine oxidase solution (0.5U/ml) was added to the mixture and incubated for one hour at 37°C, and the volume was made up to 3ml with 20mM phosphate buffer (pH 7.4). The solution was incubated at room temperature for 20 minutes, and absorbance was measured at 560 nm. Appropriate controls were included to rule out the artifacts induced reaction. The control was without any inhibitor. Inhibitory effect of *Coleus aromaticus* on superoxide radicals was calculated as follows.

$$\% \text{ Superoxide radical scavenging activity} = \frac{\text{Absorbance of control} - \text{Absorbance of test sample}}{\text{Absorbance of control}} \times 100$$

Cyto-toxicity study of the *Coleus aromaticus* extract

Human peripheral lymphocytes were isolated from 10ml of venous blood drawn from young, healthy donors. Blood was collected in ACD (85mM citric acid-71mM trisodium citrate-165mM D-glucose) in the ratio of 5:1.4 volumes of solution A (hemolysing buffer-150mM NH_4Cl in 10mM Tris buffer, pH 7.4) was added, mixed well, incubated at 4°C for 30 min.

Centrifuged at 1200 rpm for 12 min, the supernatant (hemolysate) was discarded, pellet was washed again with 5ml of hemolyzing buffer and the pellet containing cells were washed thrice with 10 ml of solution B (250mM m-inositol in 10mM phosphate buffer pH 7.4) and suspended in same solution. The cell viability was determined by Trypan dye blue exclusion method^[18]. 10µl of lymphocyte sample added 10µl of Trypan blue (0.02%) and the cells were charged to Neuberg's chamber and the cell number was counted. The dead cells being permeable to Trypan blue appear blue against white color of the viable cells. The survival rate of lymphocytes was determined at time intervals 20th, 40th and 60th minutes of incubation. Viability was tested by Trypan blue exclusion and exceeded 96% in each isolation. Percentage viability was calculated by the formula.

$$\% \text{ viability} = \frac{\text{Total no. of viable cells}}{\text{Total no. of viable cells + dead cells}} \times 100$$

Protective effect of *Coleus aromaticus* extract on H₂O₂ induced DNA damage

The DNA damage induced by H₂O₂ was analyzed on 0.6% submarine agarose gel according to the method of Sultan et al, 1995^[19]. Calf thymus DNA (10 mg) was mixed well in one ml of 20 mM Potassium phosphate buffer, at pH 7.4, 150 mM NaCl and store at 4C. The sheared 15 µg of calf thymus DNA was treated with 1mM of H₂O₂ with or without *Coleus aromaticus* and Ascorbic acid (400µM) in 100µl potassium phosphate buffer (20mM, pH 7.4). The reaction mixture was mixed with 10 µl of sample loading buffer (0.5% bromophenol blue, 50% glycerol in water) and then the reaction mixture was incubated at 37C for 30 min and then placed on ice for 10 min to stop the reaction. 20µl of the reaction mixture (15 µg of DNA) was run on 0.6% agarose with ethidium bromide (1 µg/ml). The electrophoresis was carried out using 20mM TBE buffer (40 mM Tris, 20 mM sodium acetate, 2 mM EDTA, 18 mM NaCl, pH 8) at 60 V for 3 h and DNA was visualized under a UV transilluminator.

Statistical analysis

Statistical analysis was done using student's t-test and a p < 0.05 was considered as statistically significant when compared with controls and results refer to mean ± SD.

RESULTS

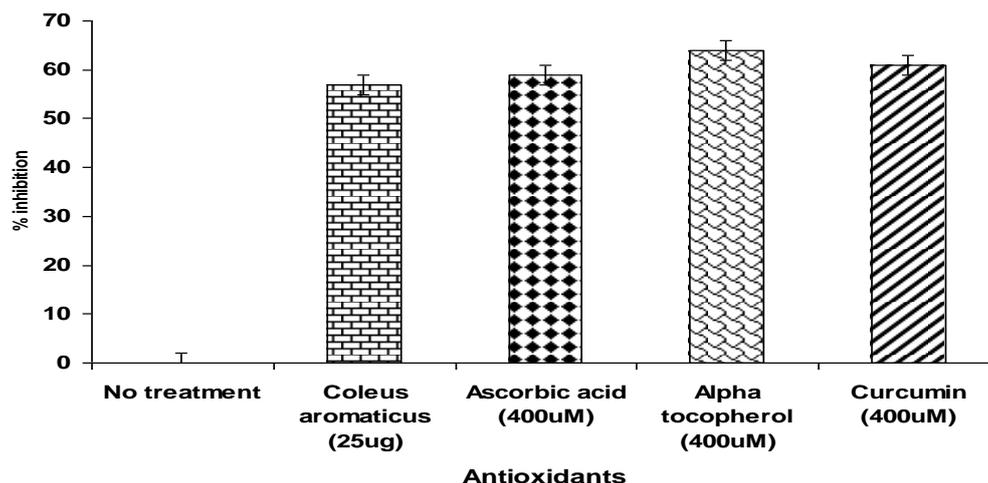


Figure-1: DPPH radical scavenging activity of *Coleus aromaticus* extract

DPPH (0.5mM) + With or without *Coleus aromaticus* extract (25 μ g) / α -tocopherol /Ascorbic acid/Curcumin (400 μ M). Mixture incubated at 37 $^{\circ}$ C for 30 min and the absorbance read at 517 nm using spectrophotometer.

Values are means \pm SD of triplicates.

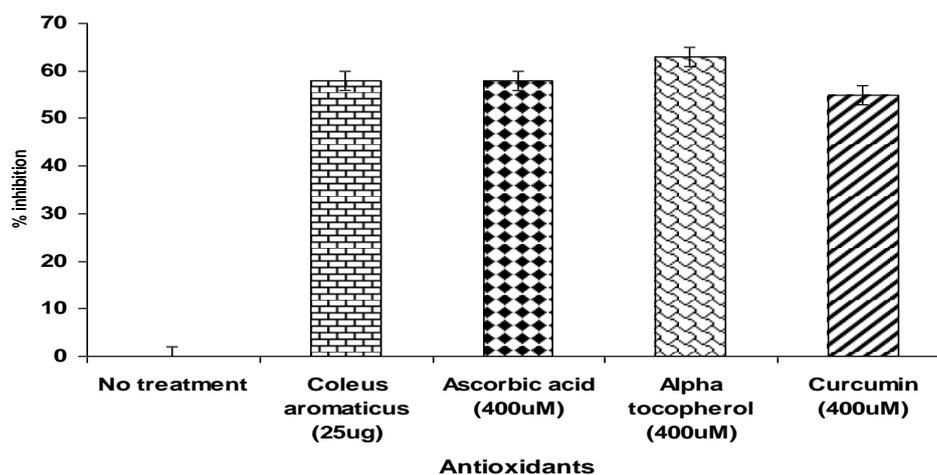


Figure-2: Superoxide scavenging activity of *Coleus aromaticus* extract

Hypoxanthine (30mM) + Nitro blue tetrazolium (NBT) (1.42mM) + With or without *Coleus aromaticus* extract (25 μ g) / α -tocopherol /Ascorbic acid/Curcumin (400 μ M). Mixture incubated at 37 $^{\circ}$ C for 20 min and the absorbance read at 560 nm using spectrophotometer.

Values are means \pm SD of triplicates.

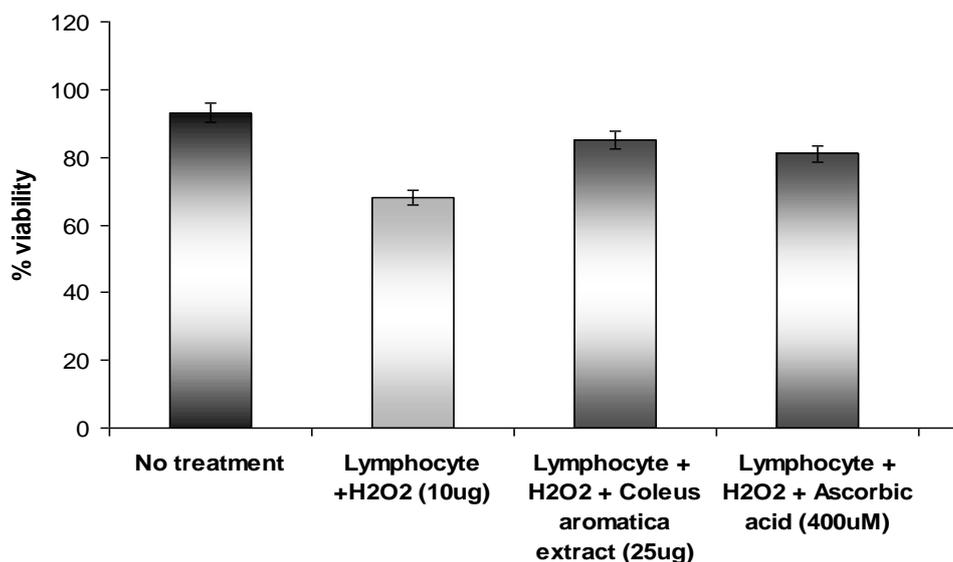


Figure-3: Cytotoxicity study of *Coleus aromaticus* extract

Lymphocytes (10^6 cells) pretreated with or without antioxidants at indicated concentrations in 0.5ml HBSS pH 7.4, incubated at 37°C for 20min., then H₂O₂ (144μM) was added, incubated at 37°C for 60 minutes in final volume of 1ml HBSS, pH 7.4. After the desired incubation time (60 minutes), viability of the cells was determined by Tryphan blue exclusion and the percentage of viable cells was calculated as mentioned in methods. Values are means ± SD of triplicates.

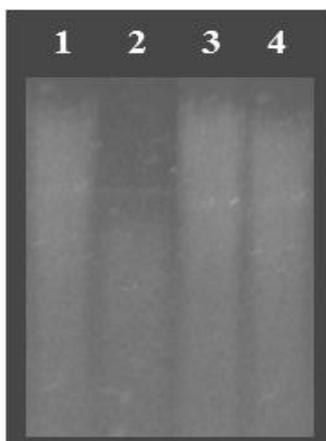


Figure 4: Inhibition of Hydrogen peroxide induced DNA degradation by extracts of Star Anise.

Lane 1: Calf thymus DNA untreated;

Lane 2: As lane (1) + H₂O₂

Lane 3: As lane (2)+ *Coleus aromaticus* extract (15μg)

Lane 4: As lane (2)+ of Ascorbic acid (15μg)

DISCUSSION

Proximate Analysis

The extraction and proximate analysis of the *Coleus aromaticus* extract was done as reported earlier.^[20]

DPPH (1,1-diphenyl-2-picrylhydrazyl) radical scavenging assay

As shown in Figure-1, a direct approach with DPPH radicals - a stable radical used to evaluate the antioxidant activity of extract of *Coleus aromaticus*. It is evidenced that, 57% scavenging activity at 25ug concentration of extract. The *Coleus aromaticus* extract exhibited a statistically significant DPPH radical-scavenging activity when compared to the group without any inhibitor. At the concentration of 400µM standard antioxidants like Ascorbic acid, α-tocopherol and Curcumin showed 59%, 64% and 61% DPPH radicals respectively.

Superoxide scavenging activity

Superoxide anions are the most common free radicals generated in biological systems and the concentration of superoxide anions increases under oxidative stress. The assay was carried to investigate the superoxide radical scavenging activity of extract of *Coleus aromaticus*. As shown in Figure-2, the superoxide scavenging activity of *Coleus aromaticus* extract (25ug) was found to be 58%. Ascorbic acid, α-tocopherol and Curcumin were used as the positive control which offered 58%, 63% and 55% scavenging activity at a concentration of 400µM.

Cytotoxicity study of *Coleus aromaticus* extract

As shown in Figure 3, The study was done to check and confirm the cytotoxic nature of the extract and other standard antioxidant Ascorbic acid. The results confirmed that, the extract of *Coleus aromaticus* and antioxidant Ascorbic acid are not toxic to cells and provide protection to lymphocytes against Hydrogen peroxide.

Protective effect of extracts of *Coleus aromaticus* on H₂O₂ induced DNA damage

As shown in figure 4, the calf thymus DNA was treated with H₂O₂ for 30 min, extensive DNA fragmentation due to oxidation by hydroxyl radicals was seen on submarine agarose gel by the enhanced mobility (Lane 2) as compared to the untreated DNA (Lane 1). The lane 3 showed that, DNA treated with H₂O₂ along with Ascorbic acid (15µg) protected DNA damage in comparison to the untreated DNA (Lane 1). The effectiveness of the extract of *Coleus aromaticus* to prevent H₂O₂ induced DNA damage was associated to its hydroxyl

radical scavenging activity. This suggests that the *Coleus aromaticus* extract could prevent H₂O₂ mediated oxidative DNA damage.

CONCLUSION

The above results indicates that, the boiling water extract of *Coleus aromaticus* acts as an effective antioxidant at a dosage of 25µg and it provide protection to DNA against peroxides. Its antioxidants activities could be mainly due to combined effect of phytochemicals present in it.

REFERENCE

1. Bayani U, Ajay VS, Paolo Z and Mahajan RT. Oxidative Stress and Neurodegenerative Diseases: A Review of Upstream and Downstream Antioxidant Therapeutic Options, *Curr Neuropharmacol*,2009; 7(1): 65–74.
2. Pallavi S, Ambuj BJ, Rama SD. and Mohammad Pessaraki, Reactive Oxygen Species, Oxidative Damage, and Antioxidative Defense Mechanism in Plants under Stressful Conditions, *Journal of Botany*, 2012, pages 26.
3. Vasuki R, Rajeswary H, Samudram P and Geetha A. Antioxidant Activity of Combined Ethanolic Extract of *Eclipta alba* and *Piper longum* Linn, *Journal of Complementary and Integrative Medicine*. 2011, Volume 8, Issue 1.
4. Jayaraman A and Muthukrishnan S. Free radical scavenging activity of ethanolic extract of *Bauhinia tomentosa* leaves collected from Kolli hills (Tamil nadu), *World Journal of Pharmaceutical Research*, 2014, Vol. 3, Issue 3, 4188-4199.
5. Jayaraman A and Muthukrishnan S. Free radical scavenging activity of ethanolic extract of *Bauhinia tomentosa* leaves collected from Kolli hills (Tamil nadu), *World Journal of Pharmaceutical Research*, 2014, Vol. 3, Issue 3, 4188-4199.
6. Dinesha R, Thammannagowda SS, Shwetha KL, Prabhu MSL, Madhu CS, and Leela Srinivas. The antioxidant and DNA protectant activities of Star Anise (*Illicium verum*) aqueous extracts, *Journal of Pharmacognosy and Phytochemistry* 2014; 2 (5): 98-103.
7. Ningappa MB, Dinesha R. and Leela S. Antioxidant and free radical scavenging activities of polyphenol-enriched curry leaf (*Murraya koenigii* L.) extracts, *Food Chemistry* 2008; 106:720-728.
8. Smitha S, Dhananjaya BL, Dinesha R, and Leela S. Purification and characterization of a w34 kDa antioxidant protein (β-turmerin) from turmeric (*Curcuma longa*) waste grits. *Biochemie* 2009; 91(9):1156-62.

9. Kaliappan ND and Viswanathan PK. Pharmacognostical studies on the leaves of *Plectranthus amboinicus Spreng*, 2008, Vol-2, Issue 3,182-184.
10. Smita S, Hemanth K, Thashma, Veena N, Krishnananda P, Priya P, Indu Warriar, Somayaji, Venu Madhav, Bairy KL and Anoop Kishore. Hepatoprotective activity of *Plectranthus amboinicus* against paracetamol hepatotoxicity in rats. International Journal of Pharmacology and Clinical Sciences, 2012, 1(2): 32-8.
11. Subhas Chandrappa M, Shivakumar Hugar, Itgappa M, Nagarajappa K, Antidiabetic and antioxidant potential of *Coleus aromaticus* leaf extracts in Alloxan induced diabetic rats, Pharmacologyonline, 2009, 3: P 1054-1061.
12. Subhas Chandrappa M, Harsha R, Dinesha R and Thammanna Gowda, SS. Antibacterial activity of *coleus aromaticus* leaves .International Journal of Pharmacy and Pharmaceutical Sciences-2010, 2 (3), P. 63-66.
13. Bradford MM. A rapid and sensitive method for the quantification of microgram of protein utilizing the principle dye binding. Annual biochem 1976; 72:245.
14. Dubois M, Gilles KA, Hamilton JK, Rebers PA and Smith F. Colorimetric method for determination of sugars and related substances. Analytical Chemistry 1956; 28:350
15. Kujala TS, Loponen JM, Klika KD and Pihlaja K. Phenolics and betacyanins in red beetroot (*Beta vulgaris*) root Distribution and effect of cold storage on the content of total phenolics and three individual compounds. Journal of Agricultural and Food Chemistry 2000; 48:5338-5342.
16. Woisky R and Salatino A. Analysis of Propolis: some parameters and procedures for chemical quality control. J Apic Res. 1998; 37:99-105.
17. Shimada K, Fujikawa K, Yahara K and Nakamura T. Antioxidative properties of Xanthan on the auto oxidation of soybean oil in cyclodextrin emulsion. Journal of Agricultural and Food Chemistry, 1992; 40:945-948.
18. Lee J, Kwon ES, Kim DW, Cha J and Roe JH. Regulation and the role of Cu, Zn-containing superoxide dismutase in cell cycle progression of *Schizosaccharomyces pombe*. Biochem Biophys Res Commun 2002; 297:854–862.
19. Phillip HJ. Dye exclusion test for cell viability. In: Tissue culture methods (Kruse PF, patterson MK eds). 1973; 406-408.
20. Phillip HJ. Dye exclusion test for cell viability. In: Tissue culture methods (Kruse PF, patterson MK eds). 1973; 406-408.

21. Sultan S, Perwaiz S, Iqbal M and Athar M. Crude extracts of hepatoprotective plants, *Solanum nigrum* and *Cichrium intybus* inhibit free radical mediated DNA damage. *Journal of Ethanopharmacology*, 1995; 45:189-192.
22. Subhas Chandrappa, Ramakrishna Harsha, Ramadas Dinesha and Salekoppal and Sannaswamy Gowda Thammanna Gowda. Antioxidant activity of water (hot and cold) and ethanol extract of *Coleus aromaticus* leaf extract – A comparative study, *Herbal Heritage*, 2009, 1 (4), P 211-217.