

EVALUATION OF SELECTED PLANT EXTRACTS AGAINST *XANTHOMONAS ORYZAE* PV. *ORYZAE* CAUSING BACTERIAL BLIGHT OF RICE

A Yugander, D Ladhakshmi, M Srinivas Prasad and G S Laha*

Department of Plant Pathology, Indian Institute of Rice Research, Hyderabad-500 030,
Telangana State.

Article Received on
26 July 2015,

Revised on 17 Aug 2015,
Accepted on 10 Sep 2015

*Correspondence for
Author

Dr. G. S. Laha

Department of Plant
Pathology, Indian Institute
of Rice Research,
Hyderabad-500 030,
Telangana State.

ABSTRACT

Bacterial blight (BB) of rice caused by *Xanthomonas oryzae* pv. *oryzae* (*Xoo*), remains a major production constraint in rice cultivation. The extent of yield loss could be as high as 50% and depends on the growth stages of the crop, cultivar, season, climatic condition and level of nitrogen fertilizer applied. Chemical control of the disease is not very successful. Use of botanicals in the management of plant diseases is gaining importance as it is safe and eco-friendly. In the present study, the efficacy of 33 plants extracts from different family was tested for their antibacterial activity against *Xoo*. Antimicrobial activity of aqueous plant extract was determined by agar well method on MWA agar medium. Among these extracts, six plant extracts exhibited

significant antibacterial activity against *Xoo*. Out of these five plant extracts were evaluated under field condition for their effectiveness to reduce the BB. Out of these, three plants extract viz., *Curcuma domestica*, *Datura metel* and *Oscimum sanctum* were most effective which is reflected in reduced lesion length. All the treatments significantly increased the yield.

KEYWORDS: Bacterial blight of rice, *Xanthomonas oryzae* pv. *oryzae*, Pathogenicity.

INTRODUCTION

Bacterial blight (BB), caused by *Xanthomonas oryzae* pv. *oryzae* (Ishiyama) Swings *et al.* (*Xoo*) is one of the most important diseases of rice and is a serious production constraint especially in the irrigated and rainfed lowland ecosystems. Major epidemics were reported in north western India and Kerala (Laha *et al.*, 2009). The disease appeared in epidemic form in

about 10,000 acres of rice variety, Samba Mahsuri in Kurnool district of Andhra Pradesh during 2010. Recently, the disease appeared in epidemic proportion in many parts of Andhra Pradesh during 2013 and 2014 (Yugander *et al.*, 2013; 2014). The disease has the potential to reduce the rice yield by 50% or more depending on the rice variety, stage of the plants and the climatic conditions. Chemical control of the disease is not very successful and none of the chemicals including antibiotics were found to provide sufficient level of protection under Indian conditions (Laha *et al.*, 2009). Use of plant based botanical pesticides offer an alternative for the management of plant diseases. There are several reports which show antibacterial and antifungal nature of several plant extracts. Few studies have reported the efficacy of selected plant extracts of against bacterial blight of rice (Kagale *et al.*, 2004; Meena *et al.*, 2013; Kumar *et al.*, 2009; Gurjar *et al.*, 2012), rhizome extract of *Acorus calamus* against brown spot of rice (Devi and Chettry, 2013) and leaf extracts of *Prosopis juliflora* and *Zizyphus jujube* against rice blast (Kamalakaran *et al.*, 2001). In the present study, we tested the antibacterial activity of 33 plant extracts against *Xoo* and efficacy of selected plant extracts in reducing the bacterial blight severity under field condition.

MATERIALS AND METHODS

Isolation and maintenance of *Xoo*: The pathogen, *Xanthomonas oryzae* pv. *oryzae* (*Xoo*) was isolated from infected rice leaf sample collected from experimental fields of Indian Institute of Rice Research following standard isolation procedure (Yugander *et al.*, 2014). The pure culture of *Xoo* was maintained in modified Wakimoto's slants at 4⁰C for short term storage and in 15% glycerol at -80⁰C for long term storage. The pathogenicity of the isolate was confirmed by inoculating on TN1 plants.

Preparation of Plant Extracts: Different plant parts (leaf, root, stem, flower buds, rhizome, bark, seeds, bulbs and fruits) of 33 different plant species belonging to different family were collected from fields in and around Indian Institute of Rice Research, Rajendranagar, Hyderabad or commercially (Table 1). The plant parts were thoroughly washed 2-3 times in running tap water followed by sterilized distilled water. The crude extracts of different plant parts were prepared in a sterile mortar and pestle separately using sterile distilled water (1:1 w/v) and filtered through muslin cloth. The filtrate was centrifuged at 15000 rpm for 10 min and the clear supernatant was collected. These extracts were then passed through bacteria proof sterile syringe filters (Axiva filters, SFPV13R). This pure extract was considered as

100% (concentrated) and was directly tested for its antimicrobial effect against *Xoo* under *in vitro* condition.

Testing of plant extracts for antimicrobial activity: Antimicrobial activity of different aqueous plant extracts was determined by agar well method on modified Wakimoto's agar (MWA) medium. Freshly prepared sterile MWA medium was uniformly poured in sterile plastic Petri plates (15 ml/plate). Once the medium was solidified, each plate was uniformly smeared with culture of *Xoo* by putting 100 μ L bacterial suspensions (3 days old) and then spreading with a sterile spreader. In each plate, 4 wells (5 mm diameter) were made in with the help of sterile cork borer and in each well 50 μ L individual plant extract was added. The plates were then incubated in undisturbed condition at 27⁰C for 72 h and the diameter of inhibition zones was measured.

Field evaluation of selected plant extracts: The effect of selected plant extracts on bacterial blight of rice was tested under field condition (cultivar TN1). The variety was raised in 3x2 m² plots following recommended package of practices. In each plot, one square meter area (block) was marked with wooden sticks when the plants were in maximum tillering stage. Three blocks (replications) were kept for each treatment. The selected plant extracts prepared in sterile distilled water (1:1) were sprayed in each marked blocks one day prior to bacterial inoculation (@ 330 ml/block). *Xoo* was grown in modified Wakimoto's medium at 27 \pm 1⁰C. Using 3-days old bacterial suspension (0.1 OD), plants were clip inoculated (Kauffman *et al.*, 1973). Eight days after *Xoo* inoculation, another spray of respective plant extracts was given. Observations were taken on 8th and 16th day, after bacterial inoculation by measuring the lesion length. In each block, observations were taken on 10 individual leaves. Observations were also taken on any phytotoxic effects of these plant extracts. Plants from each one square meter area from each treatment and control treatments were harvested and yield per sq. mt was calculated.

RESULTS

Xoo strain isolated and used in the present study produced typical susceptible reaction when inoculated on TN1 plants under glass house condition. Out of 33 plant extracts evaluated for their antibacterial activity, 6 plant species showed inhibitory effect against *Xoo* under *in vitro* condition (Table 1). These include leaf extracts of *Eucalyptus globulus*, *Datura metel* and *Ocimum sanctum*, modified stem extracts of *Allium sativum* and *Curcuma domestica* and extract of flower buds of *Syzygium aromaticum*. The mean diameter of inhibition zone varied

from 1.33 cm in case of *Eucalyptus globulus* to 3.73 cm in case of *Allium sativum* (Table 1). In case of *Eucalyptus globulus*, the bacterial growth around the inhibition zone turned blackish in colour. However, when such bacterial growth was cultured on fresh MWA medium, they showed normal yellow growth. Out of the six plant extracts which showed inhibitory effects against *Xoo*, five plant extracts were evaluated under field condition for their effectiveness to reduce the bacterial blight disease intensity. All the plant extracts significantly reduced the lesion length (Table 2). Of these five plant extracts, *Curcuma domestica*, *Datura metel* and *Oscimum sanctum* were most effective which is reflected in reduced lesion length on both dates of observation (Table 2). All the treatments also significantly increased the yield (Table 2). No phytotoxic symptoms were observed with any of the plant extracts.

Table 1: Antibacterial effect of extracts of different plant species against *Xanthomonas oryzae* pv. *Oryzae*.

Plant Name	Common names	Family	Part Used	Mean inhibition zone (cm) ± SE
<i>Allium sativum</i>	Garlic	Amaryllidaceae	stem	3.73±0.16
<i>Curcuma domestica</i>	Turmeric	Zingiberaceae	Stem	2.18 ±0.27
<i>Syzygium aromaticum</i>	Clove	Myrtaceae	Flower buds	1.48 ±0.22
<i>Eucalyptus globulus</i>	Eucalyptus	Myrtaceae	Leaf	1.33 ± 0.14
<i>Datura metel</i>	Thorn apple	Solanaceae	Leaf	1.8 ± 0.21
<i>Ocimum sanctum</i>	Holy Basil	Lamiaceae	Leaf	2.23 ± 0.07
<i>Tamarindus indica</i>	Tamarind	Fabaceae	Leaf	0
<i>Mangifera indica</i>	Mango	Anacardiaceae	Leaf	0
<i>Azadirachta indica</i>	Neem	Meliaceae	Leaf	0
<i>Zingiber officinale</i>	Ginger	Zingiberaceae	Rhizome	0
<i>Cinnamomum camphorum</i>	Camphor	Lauraceae	Bark	0
<i>Aloe vera</i>	Indian Aloe	Liliaceae	Succulent	0
<i>Piper nigrum</i>	Black pepper	Piperaceae	Seeds	0
<i>Ricinus communis</i>	Castor	Euphorbiaceae	Leaf	0
<i>Allium cepa</i>	Onion	Amaryllidaceae	Bulb	0
<i>Lantana camara</i>	Red Sage	Verbanaceae	Leaf	0
<i>Tridax procumbens</i>	Coat button	Asteraceae	Leaf	0
<i>Tagetes patula</i>	Marigold	Asteraceae	Leaf	0
<i>Parthenium hysterophorus</i>	Congress Grass	Asteraceae	Leaf	0
<i>Catharanthus rosea</i>	Periwinkle	Apocynaceae	Leaf	0
<i>Calotropis gigantea</i>	Crown flower	Asclepidaceae	Leaf	0
<i>Ageratum conyzoides</i>	Goat weeds	Asteraceae	Leaf	0
<i>Manilkara zapota</i>	Sapota	Rutaceae	Leaf	0
<i>Ipomea batatas</i>	Sweet potato	Convolvulaceae	Leaf	0
<i>Hybiscus rosa sinensis</i>	China rose	Malvaceae	Leaf	0
<i>Carica papaya</i>	Papaya	Caricaceae	Leaf	0

<i>Euphorbia hirta</i>	Lal Dudhi	Euphorbiaceae	Leaf	0
<i>Zigiphys jujuba</i>	Red date	Rhamnaceae	Leaf	0
<i>Cassia auriculata</i>	Avaram	Fabaceae	Leaf	0
<i>Withania somnifera</i>	Ashwagandha	Solanaceae	Root	0
<i>Aegle marmelos</i>	Bael	Rutaceae	Leaf	0
<i>Pongamia pinnata</i>	Pongam	Fabaceae	Leaf	0
<i>Capsicum annuum</i>	Red pepper	Solanaceae	Fruit	0

Table 2: Effect of selected plant extracts on lesion length caused by *Xanthomonas oryzae* pv. *Oryzae*.

Plant species	Mean Lesion Length (cm)		Yield (gm/m ²)
	8th day	16th day	
<i>Allium sativum</i>	2.5b	9.4b	513.3a
<i>Syzygium aromaticum</i>	2.5b	9.7b	457.0b
<i>Datura metel</i>	2.4b	8.1c	436.3bc
<i>Ocimum sanctum</i>	2.3bc	8.2c	392d
<i>Curcuma domestica</i>	2.1c	7.9c	406.7cd
Unsprayed Control	4.9a	14.1a	315.3e
CV (%)	8.46	5.13	4.48
LSD (p=0.05)	0.43	0.89	34.22

DISCUSSION

There are reports which show antibacterial nature of extracts of *Datura metel*, *Curcuma longa* and *Eucalyptus grandifolia* against rice diseases (Kagale *et al.*, 2004; Meena *et al.*, 2013; Devi and Chetry, 2013). Meena *et al.* (2013) reported presence of the antimicrobial compound withametalin B in *Datura metel*. Maximum inhibition observed with *Allium sativum* may be due to the presence of sulphur compounds and allicin (Kuruchev and Padmavathi, 1998; Singh and Singh, 2005). Many research reports indicated possible use of plant extracts for management of plant diseases. Colpas *et al.* (2009) reported that aqueous extract of leaves of *Ocimum gratissimum* induced systemic resistance in cucumber against *Colletotrichum lagenarium* as reflected by reduction in disease incidence and an increase in chitinase production. Antibacterial nature of *Datura metel*, *Curcuma longa* and *Eucalyptus grandifolia* have been demonstrated in several studies (Kagale *et al.*, 2004; Meena *et al.*, 2013; Kumar *et al.*, 2009). The antimicrobial properties of these plants has been attributed to the presence of chemical compounds viz., curcumin (in *Curcuma longa*), hyoscyamine Scopolamine and withametalin B (in *Datura stramonium* and *D. metel*) and allicin (in *Allium sativum*) (Gurjar *et al.*, 2012; Meena *et al.*, 2013).

CONCLUSIONS

The present study shows that some of these plant extracts can be used for the management of bacterial blight of rice under field condition. As the chemical control of this disease is not very successful, management of the disease with the locally available plant materials like *Datura* and *Curcuma* can be the best alternative.

ACKNOWLEDGEMENT

The authors wish to thanks and I am grateful to the following for supporting my research; Department of Plant Pathology and Project Director, Indian Institute of Rice Research (ICAR), Hyderabad.

REFERENCES

1. Colpas FT, Estrada KRFS, Stangarlin JR, Ferrarese MDL, Scapim CA, Bonaldo SA. Induction of plant defense responses by *Ocimum gratissimum* L. (*Lamiaceae*) leaf extracts . *Summa Phytopathol., Botucatu*, 2009; 35: 191-195.
2. Devi OJ, Chhetry GKN. Evaluation of Antifungal Properties of Certain Plants against *DrechsleraOryzae* Causing Brown Leaf Spot of Rice in Manipur Valley, *International Journal of Scientific and Research Publications.*, 2013; 3: 1-3.
3. Gurjar MS, Shahid A, Masood A,d Singh KS. Efficacy of plant extracts in plant disease management. *Agricultural Scences.*, 2012; 3: 425-433.
4. Kagale S, Marimuthu T, Thayumanavan B, Nandakumar R, Samiyappan R. Antibacterial activity and induction of resistance in rice by leaf extract of *D. metel* against *Rhizoctonia solani* and *Xanthomonas oryzae* pv. *oryzae*. *Physiol. Mol. Plant Pathol*, 2004; 65, 91100.
5. Kamalakannan A, Shanmugan V, Suhendran S, Srinivasan R. Antifungal Properties of plant extracts Against *Pyricularia oryzae*, Rice Blast Phatogen. *Indian Pytopath*, 2001; 5; 4(4): 490-492.
6. Kauffman HE, Reddy APK, Hsieh SPY, Merca SD. An improved technique for evaluating resistance of rice varieties to *Xanthomonas oryzae*. *Plant Disease Rep*, 1973; 56: 537-541.
7. Kumar M, Parate RL, Ninawe BNN. Effect of Botanicals, Bioagents and some chemicals against *Xanthomonas oryzae* pv. *oryzae*. *J.Pl.Dis.Sci*, 2009; 4(1): 60 - 63
8. Kurucheve V, Padmavathi R. Management of damping off of chillies with plant products. *Indian Phytopathology*, 1998; 51: 379-81.

9. Laha GS, Reddy CS, Krishnaveni D, Sundaram RM, Srinivas Prasad M, Ram T, Muralidharan K, Viraktamath BC. Bacterial blight of rice and its management. Technical Bulletin No.41. Directorate of Rice Research (ICAR), Rajendranagar, Hyderabad-500 030, A.P., India., 2009; 37.
10. Meena C, Gopalakrishnan J, Dureja P. Antibacterial Withametelin B from *Datura metel* against *Xanthomonas oryzae* pv. *oryzae*. *Biopestic. Int*, 2013; 9(1): 31-37.
11. Singh M, Singh RP. Management of mushroom pathogens through botanicals. *Indian Phytopathology*, 2005; 57: 189-93.
12. Yugander A, Sundaram RM, Srinivas Prasad M, Laha G S. Outbreak of Bacterial Blight of Rice in Guntur District of Andhrapradesh. *DRR Newsletter*, 2013; 11: 5.
13. Yugander A, Ershad Md, Mutturaman P, Sundaram RM, Sheshu Madav M, Laha GS. Severe outbreak of bacterial blight of rice in East and West Godavari districts of Andhra Pradesh. *DRR News letter*, 2014; 12: 5.
14. Yugander A, Sundaram RM, Ladhakshmi D, Hajira Shaik, Sheshu Madhav M, Srinivas Prasad M, Viraktamath BC, Laha GS. Pathogenic and genetic profile of *Xanthomonas oryzae* pv. *oryzae* isolates from Andhra Pradesh. *Indian j. Plant Protection*, 2014; 42(2): 149-155.