

IMMUNOPHENOTYPING OF HIV PATIENTS CO-INFECTED WITH *MYCOBACTERIUM TUBERCULOSIS*

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ABSTRACT

Human immunodeficiency virus and *Mycobacterium tuberculosis* have been linked with immune suppression leading to an impaired immune response to pathogenic infections. Several studies on HIV, TB, HIV/TB co-infection were previously focused on the estimation of immunological damage to various human tissues. This present study attempt to immunophenotype the CD4⁺ and CD8⁺ cells counts of HIV, TB and HIV/TB subgroups bearing in mind age and gender

relationships of the subjects. A cross-sectional random sampling method was used for 400 subjects. Flow cytometer was used for immunophenotyping of CD4⁺ and CD8⁺ cell counts. HIV diagnosis was performed using the National Algorithm utilizing Determine, Stat Pak and Uni-Gold rapid test kits. while gene-Xpert was used for identification *Mycobacterium tuberculosis*. The result observed showed a significant variation in the ages of HIV, TB and HIV/TB categories and also in their gender (P < 0.001 and P < 0.002 respectively). Chronologically, Older HIV patients from (30 - 59) had significantly lower CD4⁺ count with the least counts found within (60 - 69) age group. A higher significant decrease was also observed between the mean CD4⁺ and CD8⁺ counts of HIV, TB, HIV/TB categories (P < 0.005 and P < 0.012 respectively). A Post Hoc test of a homogeneous subset was used to ascertain which of the group is responsible for the significant difference in the mean CD4⁺ and CD8⁺ counts of HIV, TB, HIV/TB categories. Result revealed that the mean CD4⁺ counts of (HIV and HIV/TB, TB and HIV/TB) groups and mean CD8⁺ counts of (HIV and TB, HIV and HIV/TB) groups contribute higher to the significant decrease (P < 0.001) observed that

the mean CD4⁺ categories of (HIV and TB) and the mean CD8⁺ of (TB and HIV/TB) groups which was not significant ($P > 0.0603$). Since older HIV patients from (30 - 69) showed lower CD4⁺ count and the least count found within 60 - 69 age group and since the CD4⁺ cell counts decrease with increasing age of the study participants. Developing program targeting older subject from 30 - 69 age group and applying methods with high productivity, sensitivity and specificity will help in faster medical treatment in the countries with high TB, HIV/AIDS and HIV/TB prevalence and in general population.

KEYWORDS: Immunophenotyping, HIV, TB, HIV/TB, CD4⁺ and CD8.

INTRODUCTION

Human Immunodeficiency Virus (HIV) and *Mycobacterium tuberculosis* (MTB, TB) are common medical disasters ravaging human race (WHO, 2016). HIV/TB co-infection is one of the major public health burdens globally. According to the World Health Organization (WHO), nearly 15% of individuals diagnosed with MTB were HIV positive and a total of 360,000 deaths were attributed to MTB-HIV co-infection alone (Maria *et al.*, 2015). Clinical evidence suggests that MTB is the most common opportunistic infection causing exacerbation of viral load and diminished CD4⁺ count in HIV patients. The possibility of acquiring *Mycobacterium tuberculosis* infection is 20 - 37 times greater in HIV positive patients than non-infected individuals (Guinn and Rubin, 2017), and HIV remains a significant risk factor for *M. tuberculosis* (Rohan and Indra 2015). *Mycobacterium tuberculosis* remains a leading cause of death in people living with HIV (Daria *et al.*, 2014). The World Health Organization (WHO) estimates that there are about 10.4 million new cases and 1.8 million deaths from TB each year. One-third of these new cases (about 3 million) remain unknown to the health system, and many are not receiving proper treatment (Bloom *et al.*, 2017). About 13% of tuberculosis cases worldwide were co-infected with HIV, and approximately 430,000 deaths in HIV-infected persons in 2011 were due to tuberculosis (Agneta *et al.*, 2014). HIV co-infection with TB remains a major problem in Sub-Saharan Africa (Agneta *et al.*, 2014). The incidence of *M. tuberculosis* remains high in Africa and Asia and sub-Saharan Africa (Mukhtar *et al.*, 2014). *Mycobacterium tuberculosis* stimulates conventional and unconventional T-cell subsets (Teresa *et al.*, 2014). The predominant response is mediated by naturally restricted, peptide-specific Th1 type CD4⁺ T cells and CD8⁺ cytotoxic T lymphocytes (CTLs), which are crucial for protective immunity in murine models of tuberculosis (Yang *et al.*, 2018). T-cell deficiency increases susceptibility to

disease in animal models (Marcos *et al.*, 2015). The severity of *Mycobacterium tuberculosis* in HIV positive patients is a good indication of the importance of CD4⁺ T-cells in control of tuberculosis. CD8⁺ T-lymphocytes play an important role in the effectiveness of T-cell immune reaction (Yang *et al.*, 2018), by excreting cytokines such as IFN- γ and IL-4 which modulate the balance of Th1 and Th2 cells in the lungs of patients with pulmonary tuberculosis (Patrizia *et al.*, 2017). CD4⁺ and CD8⁺ T-cells have many effector functions that facilitate anti mycobacterial activities and CD8⁺ T-cells respond by stimulating immune cells to provide protective immunity to the system (Yang *et al.*, 2018).

Functional CD4⁺ and CD8⁺ T-cell subsets have been describe based on single-cell cytokine (interferon γ [IFN- γ]/interleukin 2 [IL-2]/tumor necrosis factor α [TNF- α]) signatures. These are differentially impacted by disease stage, Mycobacterial load, and treatment (Katrina *et al.*, 2013). This means that certain subsets may serve as biomarkers of disease activity, pathogen burden, or treatment response. However, till date there are evidence of scarcity of data on the various cell surface marker phenotype of these subsets of *M. tuberculosis* infection, which is integral in the characterization of T cells, denoting memory status, disease site homing, survival, and activation of cellular reaction (Drain *et al.*, 2018). Changes in bacteria main function defined memory cell response associated with varying antigen load in other disease models (Prezzemolo *et al.*, 2014), and many studies insinuate that *M. tuberculosis* specific cells existing in active tuberculosis are mainly of effector memory phenotype (Chiacchio *et al.*, 2014).

Several reports have suggested measuring CD4⁺ cells of the *M. tuberculosis*-specific TNF- α -only secreting cells serving as a precise biomarker of active tuberculosis (Chiacchio *et al.*, 2014). It was recently found that primate CD8⁺ T cells and their effector function are critical for controlling *M. tuberculosis* infection and TB lesions (Philana *et al.*, 2015), but sustaining CD8⁺ T-cell responses relies on helper function of CD4⁺ T cells (Yao *et al.*, 2014). Consistently, the depletion of TNF- α + CD8⁺ T cells by anti-TNF- α immunotherapy may contribute to reactivation of TB in humans (Shao *et al.*, 2016). Thus, immunological studies of correlation between *M. tuberculosis* co-infection and CD4⁺ and CD8⁺ T cells in HIV-infected humans will ultimately help define anti-TB immunity and mechanisms of the T-cell population. This study will help us explain the biological and metabolic difference in disease management. The disparity in age and gender differences has not been elucidated, investigating the specific activity will highlight the age levels of HIV, TB HIV/TB. There is a

conflicting result background patterns of gender disparities in HIV, HIV and TB co-infection observed from previous studies. These form the basis of this study.

MATERIALS AND METHODS

STUDY AREA

This study was carried out at the Federal Teaching Hospital (FETHI), IdoEkiti, Nigeria. The Hospital serves an estimated population of 107,000 and also cover a large geographical area boarding the northern part with an estimated area of 6353 km²2453 square miles. FETHI was upgraded in 2006 to serve as a referral hospital for HIV/AIDS, diagnosis and treatment and had remained a center of excellence for *M. tuberculosis* and also offers a free treatment and management of subjects with HIV and those on antiretroviral therapy in Ekiti State. The hospital is currently serving neighboring states such as Osun, Kogi, and Kwara etc.

STUDY DESIGN

A total of four hundred (400) subjects were recruited and their samples analyzed in this study. These subjects were grouped into three categories; 150 HIV infected subjects, 150 individuals infected with *M. tuberculosis* and 100 HIV individuals co-infected with *M. tuberculosis*, all the subjects that participated in this research were on HAART, and anti-Tb Drugs. All patients seen were included in this work.

SAMPLE COLLECTION

Six milliliter (6mls) of whole blood were collected from each subject for CD4⁺ and CD8⁺ cell count. The remaining sample was used for HIV status screening on the subjects.

Sputum Sample Collection Procedures

Sputum samples were collected from the individual subjects early in the morning before eating or drinking. The subjects were instructed to breathe in and out deeply 2 to 4 times, to give a series of low deep coughs to raise sputum from the lungs and deposit material into the sterile container. The containers was immediately covered with a screw cap cover to prevent leakage, and then labeled with the date and time of collection. The sputum specimens was also collected from subjects for three days and each morning with three containers respectively.

Identification of *Mycobacterium tuberculosis* (MTB) Genexpert techniques as described by Somily *et al.*, (2016).

Principle of the PCR

The gene expert identify DNA sequence specific for *M. tuberculosis* and rifampicin resistance by polymerase chain reaction. The test is based on the Cepheid geneXpert system were a platform for rapid and simple-to-use nucleic acid amplification tests (NAAT). The Xpert MTB/RIF purifies and presents *M. tuberculosis* bacilli in sputum samples, isolates genomic material from the captured bacteria by sonication and successively amplifies the genomic DNA by PCR and identifies all the clinically relevant rifampicin resistance-inducing mutations in the RNA polymerase beta (rpoB) gene in the *M. tuberculosis* genome in a real-time setup using fluorescent probes called molecular beacons. Results are obtained from unprocessed sputum sample in 2 hours.

Determination of HIV Seropositivity

The method used in this study was the National Algorithm for HIV screening utilization Determine, Uni-Gold and Stak Pak kits (Ogbonnaya *et al.*, 2015).

Principle of Flow Cytometer (Adan *et al.*, 2017)

The principle of flow cytometer is based on the passage of cells in single file to the laser beam so as to be detected, counted and sorted. Cell components are fluorescently labeled and then excited by the laser to emit light at varying degree of wavelengths. The fluorescence is then measured to determine the amount and type of cells present in a sample. Thousands of particles per second can be analyzed as they pass through the liquid stream. A beam of laser light is directed at a hydrodynamically-focused stream of fluid that carries the cells. Many detectors are precisely placed around the stream near the fluid as it passes through the light beam. One of these detectors is in line with the light beam and is used to measure forward scatter (FSC). Another detector is placed perpendicular to the stream and is used to measure Single Scatter (SSC). Human immunodeficiency virus (HIV) infection is characterized by a decrease, and eventually, a depletion of CD4⁺ T-lymphocytes (helper T cells). By using immunophenotyping, patient specimens are tested for the proportion of lymphocytes that are T cells, B cells, natural killer (NK) cells, CD4⁺ T cells (helper T cells), and CD8⁺ T cells (suppressor/inducer T cells). This is done by incubating anti-coagulated whole blood with monoclonal antibodies to the various cellular antigens that identify specific cell populations (phenotypes) and then lysing the blood to remove red blood cells. The antibodies are conjugated to fluorescent tags that emit light at a certain frequency when excited by a laser beam. The specimens are then analyzed in flow cytometer to determine the proportion of

cells with a particular phenotype (that emit light at the right wavelength). The absolute count of a full lymphocyte subset profile CD4⁺ and CD8⁺ was determined in four tubes with TriTEST by calculating the ratio of regional events for each subset to bead events using the BD Biosciences-developed software, A precise quantity of whole blood is added to the tubes, and the lymphocytes are stained with TriTEST monoclonal antibodies (mAb).

STATISTICAL ANALYSIS

Data collected were subjected to SPSS Version 16.0 (IBM, 2016) statistical analysis using the chi-square and students't-test. Data was significant when $P \leq 0.05$

RESULTS

Table 4.1: Age and Gender distributions of Human immunodeficiency virus-seropositive subjects, *M. Tuberculosis* positives, and HIV Subjects Co-infected with *M. Tuberculosis*.

Age group (in years)	HIV (%) N = 150	TB (%) N = 150	HIV/TB (%) N = 100
10 – 19	11 (7.3)	2 (1.3)	0 (0.0)
20 – 29	13 (8.7)	23 (15.3)	2 (2.0)
30 – 39	33 (22.0)	44 (29.3)	38 (38.0)
40 – 49	56 (37.3)	36 (24.0)	40 (40.0)
50 – 59	29 (19.3)	22 (14.7)	20 (20.0)
60 – 69	8 (5.3)	23 (15.3)	0 (0.0)
<i>Chi square = 56.747, p value <0.001</i>			
Gender			
Male	59 (39.3)	87 (58.0)	57 (57.0)
Female	91 (60.7)	63 (42.0)	43 (43.0)
<i>Chi square = 12.539, p = 0.002</i>			

Table 4.2: Age and Gender variations of CD4⁺ levels of Human immunodeficiency virus- seropositive Subjects, *M. Tuberculosis* positives, and HIV Co-infected with *M. Tuberculosis*.

Age group (in years)	HIV N = 150	TB N = 150	HIV/TB N = 100	F	p-value
	{(Mean ± SD) (Cells/μL)}	{(Mean ± SD) (Cells/μL)}	{(Mean ± SD) (Cells/μL)}		
10 – 19	761.91 ± 255.84	237.00 ± 0.000		25.128 ^t	<0.001
20 – 29	672.08 ± 230.54	521.83 ± 234.24	489.00 ± 0.00	31.662	<0.001
30 – 39	284.88 ± 163.18	379.32 ± 197.01	308.21 ± 130.36	12.357	<0.001
40 – 49	263.90 ± 115.73	269.92 ± 110.47	256.70 ± 97.46	0.440	0.644
50 – 59	236.90 ± 105.73	241.00 ± 132.96	168.25 ± 82.74	15.001	<0.001
60 – 69	176.13 ± 56.35	283.13 ± 156.76		7.867 ^t	<0.001
Gender					

Male	285.14 ± 158.25	359.49 ± 185.03	278.88 ± 120.64	10.745	<0.001
Female	365.22 ± 254.88	307.13 ± 200.67	242.49 ± 123.70	10.507	<0.001

t – Independent *t*-test

Table 4.3: Age and Gender variations CD8+ levels of Human immunodeficiency virus-seropositive Subjects, *M. Tuberculosis* positives, and HIV Co-infected with *M. Tuberculosis*.

Age group (in years)	HIV N = 150 {(Mean ± SD) (Cells/μL)}	TB N = 150 {(Mean ± SD) (Cells/μL)}	HIV/TB N = 100 {(Mean ± SD) (Cells/μL)}	F	p-value
10 – 19	358.64 ± 103.91	205.00 ± 0.000		18.109 ^t	<0.001
20 – 29	325.85 ± 64.24	300.09 ± 44.52	213.00 ± 0.00	83.079	<0.001
30 – 39	280.42 ± 75.51	314.18 ± 130.94	262.37 ± 57.82	7.817	<0.001
40 – 49	347.87 ± 211.51	300.53 ± 131.74	339.45 ± 57.82	3.843	0.022
50 – 59	288.41 ± 50.82	295.95 ± 49.47	268.75 ± 56.40	8.474	<0.001
60 – 69	694.38 ± 383.47	266.78 ± 59.57		13.495 ^t	<0.001
Gender					
Male	319.86 ± 138.40	290.10 ± 89.94	301.14 ± 123.78	2.404	0.092
Female	351.24 ± 209.88	307.35 ± 117.96	283.35 ± 68.96	6.600	0.002

t – Independent *t*-test

Table 4.1 shows a higher significant variation in both age and gender of HIV, TB and HIV/TB subjects in the three study categories ($P < 0.001$ and $P < 0.002$ respectively). Also Male subjects had greater percentage of TB and HIV/TB (58.0%), (57.0%) than Female subjects which had (42.0% and 43.0%). Greater number of females were infected with HIV (60.7%) than Male (39.3%) subjects. The preponderance of the subjects infected with HIV, TB, and HIV/TB fell within 30 to 49 years age bracket.

Table 4.2 compared the difference mean CD4⁺ counts of HIV, TB, HIV/TB subjects studied. Results obtained showed higher significant decrease in both age and gender respondents ($P < 0.001$), except subjects within the age bracket of 40 - 49 years which was not significant ($P > 0.644$). Chronologically, Older HIV patients from (30 - 59) had significantly lower CD4⁺ count with the list counts found within (60 - 69) age group.

Table 4.3 also compared the difference mean CD8⁺ counts of HIV, TB, HIV/TB groups and there were higher significant decrease in all the three group and also in female subjects ($P < 0.001, 0.022, 0.002$) except male subjects which was not significant ($P > 0.092$).

Table 4.4: Mean CD4⁺ and CD8⁺ levels of Human immunodeficiency virus-seropositive Subjects, *M. Tuberculosis* Subjects, and HIV Co-infected with *M. Tuberculosis*.

Variable	HIV N = 150 {(Mean ± SD) (Cells/μL)}	TB N = 150 {(Mean ± SD) (Cells/μL)}	HIV/TB N = 100 {(Mean ± SD) (Cells/μL)}	F	p-value
Mean CD4 ⁺	333.72 ± 224.79	337.50 ± 192.84	263.23 ± 122.69	5.371	0.005
Mean CD8 ⁺	338.90 ± 185.20	297.35 ± 102.63	293.49 ± 103.74	4.485	0.012

Table 4.5: Tukey Post Hoc HSD Test for the Mean CD4⁺ and CD8⁺ of Human immunodeficiency virus-seropositive Subjects, *M. tuberculosis* positives, and HIV Subjects Co-infected with *M. tuberculosis*.

(I)	(J)	/(I – J)/	p-value
CD4⁺			
HIV	TB	3.78	0.876
HIV	HIV/TB	70.49	<0.001
TB	HIV/TB	74.27	<0.001
CD8⁺			
HIV	TB	41.55	<0.001
HIV	HIV/TB	45.41	<0.001
TB	HIV/TB	3.86	0.772

Table 4.4 shows the comparison between mean CD4⁺ and CD8⁺ counts of HIV, TB, HIV/TB groups studied. Results obtained showed a higher significant variation in the mean CD4⁺ and CD8⁺ counts of the three study categories (P < 0.005 and P < 0.012 respectively).

In Table 4.5, A Post Hoc test of a homogeneous subset was used to ascertain which of the group is causing the higher significant variation recorded in the mean CD4⁺ and CD8⁺ counts of the HIV, TB, HIV/TB category. Result obtained showed that the mean CD4⁺ counts of (HIV and HIV/TB, TB and HIV/TB) groups and mean CD8⁺ counts of (HIV and TB, HIV and HIV/TB) groups was responsible for higher significant variation (P < 0.001) observed than the mean CD4⁺ categories of (HIV and TB) and the mean CD8⁺ of (TB and HIV/TB) groups was not significant.

DISCUSSION

Several T-lymphocytes are involved in the inflammatory response against TB and HIV such as interleukin 2 [IL-2], tumor necrosis factor α [TNF- α], CD4⁺ T cells, and CD8⁺T cells. Among all, CD4⁺ T cells and CD8⁺ T cells play a critical role in immune response against TB and HIV (Cecilia *et al.*, 2014). In the present study, an attempt has been made to immunophenotype the circulating CD4⁺ and CD8⁺ cells counts of HIV, TB and HIV/TB

subgroups. Result recorded in this study showed a significant variation in the ages of HIV, TB and HIV/TB categories and also in gender ($P < 0.001$). This findings is in agreement with the earlier studies by Genene and Solomon (2016) who recorded significant variations in gender disparity of HIV infection. Greater number of Female were infected with HIV (60.7%) than Male (39.3%) subjects in this study. Female vulnerability to HIV infection might be aggravated by their inability to negotiate safe sex through condom use. A study in South Africa noted that the gender power imbalance significantly affected female ability to suggest condom use to their partners (Taha *et al.*, 2018). In general, the basic background patterns of gender disparities in HIV infection observed from previous studies are generally consistent, but the higher risk of HIV infection among women than men persists, despite men exhibiting multiple sex partners (Hemmige *et al.*, 2012). Women are particularly at risk of rape or sexual assault in conflict situations. Lack of data on gender disaggregated on TB and HIV are not routinely collected or reported, making it difficult or impossible to determine sex differences or gender dynamics in TB. For example, our study revealed that higher number of male subjects were infected with TB (58.0%) and HIV/TB (57.0%) than female subjects which had (42.0% and 43.0%). A predominant increase in TB/HIV patients was reported in male subjects than female with a positive difference of 88.8%, compare to 69% for females (Matilda *et al.*, 2017). In another study in Albania (Balla *et al.*, 2016), (88%) of TB and HIV patients were males. In Albania, more men than women are diagnosed with HIV and TB. This gender difference could result from under-diagnosis among women, differences in sociocultural causes or various issue related with access in healthcare system and services (Balla *et al.*, 2016).

The Mean age CD4⁺ and CD8⁺ counts of HIV, TB and HIV/TB categories also showed a higher significant decrease and also in gender ($P < 0.001$), except subjects within the age bracket of 40 - 49 years and Male Subject which was not significant ($P > 0.644$ and $P > 0.092$) respectively. Chronologically, Older HIV patients from (30 - 59) had significantly lower CD4⁺ count with the least counts found within (60 - 69) age group. Results on investigations of age-associated differences in CD4⁺ T-cell response for HIV-infected adults on HAART have been inconsistent in the context of cART (Silverberg *et al.*, 2007). The progression from Latently infected TB (LTBI) to Pulmonary (PTB) might be due to CD4⁺ T-cell depletion driven by *M. tuberculosis*-specific CD4⁺ and CD8⁺ T-cell function resulting from pathological impairment (Amelio *et al.*, 2018). This shows that HIV-infected individuals might be harboring reduced *M. tuberculosis*-specific CD4⁺ T-cell frequencies

associated with significant changes in *M. tuberculosis*-specific CD4⁺ T-cell cytokine profiles. Although active anti-viral therapy (ART) reduces opportunistic infections in HIV-infected patients (Cingolani *et al.*, 2011), the increased risk of TB conferred by HIV infection does not appear to be significantly diminished by ART. An immune competent TB and HIV co-infected adult men and women on anti-tuberculosis therapy - with or without ART, were found to be associated with age differential rise in CD4⁺ of TB patients on concurrent anti-tuberculosis and antiretroviral therapy. For patients on simultaneous TB and ART therapies Similar results have been reported by Dodd *et al.*, (2016) who found that TB contributes to the decrease in CD4⁺ T-cell count during TB/HIV co-infection. previously Amara *et al.*, (2015) had examined Age and gender relation to CD4⁺ T-cell and reported a higher significant difference in HIV/TB co-infected adults receiving antiretroviral therapy (cART). Our findings is in agreement to the above work which reported higher significant difference in age and gender of TB and HIV/TB subjects. The age-related CD4⁺ recovery rate of subjects is depended on gender and nutritional status (Amara *et al.*, 2015). Previous studies carried out in HIV/TB infected subjects not receiving ART show variable results with regard to change in CD4⁺ counts following ATT (Swaminathan *et al.*, 2008). HIV infection clearly increases susceptibility to TB (Lewinsohn *et al.*, 2003), but relative importance of CD4⁺ T cell depletion in HIV infection or both have not been elucidated. Recent mechanistic studies in non human primates have demonstrated that CD4⁺ T cells are clearly needed to control TB infection and sustain multi-effector function of CD8⁺ T cells and other immune cells (Yao *et al.*, 2014). Although, research that demonstrate CD8⁺ T cells protection against TB is lacking in humans and controversial in mice (Lewinsohn *et al.*, 2003), primate CD8⁺ T cells have shown to be significant for immunity against TB (Chen *et al.*, 2009). The mean CD4⁺ and CD8⁺ counts of HIV, TB, HIV/TB groups was also compared in this study and there were significant variation in the mean CD4⁺ and CD8⁺ counts of study categories (P < 0.005 and P < 0.012 respectively). A Post Hoc test of a homogeneous subset was used to ascertain which of the group is causing the significant variation recorded in the mean CD4⁺ and CD8⁺ counts of the study category. Result obtained showed that the mean CD4⁺ counts of (HIV and HIV/TB, TB and HIV/TB) groups and mean CD8⁺ counts of (HIV and TB, HIV and HIV/TB) groups contributed higher to the significant decrease (P < 0.001) observed than the mean CD4⁺ categories of (HIV and TB) and the mean CD8⁺ of (TB and HIV/TB) groups which was not significant. The pattern of this finding is suggesting a higher advanced immunodeficiency in co-infected patients. this is similar to the work of (Moustapha *et al.*, 2013), who recorded a significantly fewer mean CD4⁺ and CD8⁺ T-cell counts in TB and

HIV co-infected patients compared with single-infected subjects with either TB or HIV alone, Ddo *et al.*, (2003) also reported that TB contribute to the decrease in CD4⁺ T-cell count during TB/HIV co-infection. Several studies have reported an involvement of TB in the acceleration of immunodeficiency and increased virus replication in HIV infection (Rodrigues *et al.*, 2002; Morris *et al.*, 2003).

CONCLUSION

It has remained unknown whether CD8⁺ T cells contribute to anti-TB immunity against *M. tuberculosis* co-infection or active TB in HIV-infected persons. The devastating impact of HIV/TB co-infection is increasing with little supplementary effort toward management and treatment of such individual. Developing program targeting older subject from 30 - 69 age group and applying methods with high productivity, sensitivity and specificity will help in faster medical treatment in the countries with high TB, HIV/AIDS and HIV/TB prevalence and in general population.

REFERENCES

1. Adan A, Alizada G, Kiraz Y, Baran Y, Nalbant A (2017). Flow cytometry: basic principles and applications. *Critical Reviews in Biotechnology*, 37(2): 163-176.
2. Agneta M, Anthony G, Andrea AK, Abraham K, Herman W, John W, Katherine R, Tom O, William KM, Timothy AK, Kevin MD (2014). Tuberculosis and HIV at the National Level in Kenya: Results From the Second Kenya AIDS Indicator Survey. *Journal of Acquired Immune Deficiency Syndrome*, 1; 66(Suppl 1): S106–S115.
3. Amara, E.E., Ezekiel, M., James, O., Leonardo, M., Robert, K., Xiaoping, Y., Juliet, N.S. and Christopher, C.W. (2015). Age, sex, and nutritional status modify the CD4⁺ T-cell recovery rate in HIV–tuberculosis co-infected patients on combination antiretroviral therapy. *International Journal of Infectious Diseases*, 35: 73-79.
4. Amelio, P., Portevin, D., Hella, J., Reither, K., Kamwela, L., Lweno, O., Tumbo, A., Geoffrey, L., Ohmiti, K., Ding, S., Pantaleo, G., Daubenberger, C., Perreau, M (2018). HIV Infection Functionally Impairs *Mycobacterium tuberculosis*-specific CD4 and CD8 T-cell responses". *Journal of Virology*, 01728-18.
5. Balla, F., Drevishi, M., Bani, R., Balla, E. and Bino, S. (2016). HIV/TB mortality in Albania during the years 2004-2015, Proceedings of Academics Era International Conference, Zurich, Switzerland. ISBN: 978-93-86083-02-9.

6. Bloom BR, Atun R, Cohen T, Dye C, Fraser H, Gomez GB, Knight G, Murray M, Nardell E, Rubin E, Salomon J, Vassall A, Volchenkov G, White R, Wilson D, Yadav P (2017). Tuberculosis. Major Infectious Diseases. 3rd edition. Washington (DC): The International Bank for Reconstruction and Development / The World Bank; Chapter 11.
7. Cecilia S LA, David L, Alessandro S, Deborah L (2014). Antigens for CD4 and CD8 T Cells in Tuberculosis. *Cold Spring Harbor Perspective Medicine*, 4(7): a018465.
8. Chen, C. Y., Huang, D., Wang, R. C., Shen, L., Zeng, G. and Yao, S. (2009). A critical role for CD8 T cells in a nonhuman primate model of tuberculosis. *PLoS Pathogen*, 5: e1000392.
9. Chiacchio T, Petruccioli E, Vanini V, Cuzzi G, Pinnetti C, Sampaolesi A, Antinori A, Girardi E, Goletti D (2014). Polyfunctional T-cells and effector memory phenotype are associated with active TB in HIV-infected patients. *Journal of Infection*, 69(6): 533-45.
10. Cingolani, A., Cozzi, L.A., Castagna, A., Goletti, D., De-Luca, A. and Scarpellini, P. (2011). Impaired CD4 T-Cell Count Response to Combined Antiretroviral Therapy in Antiretroviral-Naive HIV-Infected Patients Presenting With Tuberculosis as AIDS-Defining Condition. *Clinical Infectious Disease*, 54: 853–861.
11. Daria NP, Alexander MP, Daniel G, Frank AP, Jose MM, Mathias B, Hansjakob F, Niels O, Enrico G, Anna V, Marcelo HL, Alejandro A, Joan C, Aza R, Indra Z, Anne MW, Jens DL, Amanda M, Ole K (2014). *European Respiratory Journal*, 43: 166-177.
12. Ddo, S.R., de Cunha, R.M. and Kallas, E.G. (2003). Distribution of naive and memory/effector CD4⁺ T lymphocytes and expression of CD38 on CD8⁺ T lymphocytes in AIDS patients with tuberculosis. *Brazilian Journal of Infectious Disease*, 7(2): 161–165.
13. Dodd, P.J., Looker, C., Plumb, I., Bond, V., Schaap, A. and Shanaube, K. (2016). Age- and sex-specific social contact patterns and incidence of *Mycobacterium tuberculosis* infection. *American Journal of Epidemiology*, 183: 156–166.
14. Drain PK, Bajema KL, Dowdy D, Dheda K, Naidoo K, Schumacher SG, Ma S, Meermeier E, Lewinsohn DM, Sherman DR (2018). Incipient and subclinical tuberculosis: a clinical review of early stages and progression of infection. *Clinical Microbiol Reviews*, 31: e00021-18.
15. Genene T and Solomon G (2016). Treatment outcomes of childhood tuberculosis in Addis Ababa: a five-year retrospective analysis. *BMC Public*, 16: 612.
16. Guinn KM, Rubin EJ (2017). Tuberculosis: just the FAQs. *MBio*, 8: e01910-17.

17. Hemmige V, McFadden R, Cook S, Tang H, Schneider JA (2012). *Journal of General internal Medicine*, 27: 1047.
18. IBM Corp (2016). IBM SPSS Statistics for Windows, Version 24.0. Armonk, NY: IBM Corp.
19. Katrina MP, Hilary SW, Damien JM, Lisa G, Graham SC, Moses SK, Onn MK, Robert DS, Graham PT, Ajit L (2013). T-Cell Immunophenotyping Distinguishes Active From Latent Tuberculosis. *The Journal of Infectious Diseases*, 208(6): 952–968.
20. Lewinsohn D.A., Amy, S. H., James, M. G., LiqingZhu, M.R. and David, M.L. (2003). *Mycobacterium tuberculosis*-specific CD8 T cell preferentially recognize heavily infected cells. *American Journal of Respiratory and Critical Care Medicine*, 168: 1346-1352.
21. Marcos VD, Monique GST, Juliana RM, Djalma AAS, Denise BRR, Virmondes R, Carlo JFO (2015). Complexity and Controversies over the Cytokine Profiles of T Helper Cell Subpopulations in Tuberculosis. *Journal of Immunology Research*, 20(15): 639107.
22. Maria M, Alexandria B, Arun C, Naveen P (2015). (*Mycobacterium tuberculosis* infection in a HIV-positive patient. *Respiratory Medicine Case Repository*, 2015; 16: 160–162.
23. Matilda, G., Jul, B., Arjan, H., Hasan, H. and Pellumb, P. (2017). Tuberculosis in HIV/AIDS Patients. *MedPub Journals*, 3: 16.
24. Morris, L., Martin, D. J., Bredell, H., Nyoka, S.N., Sacks, L. and Pendle, S. (2003). Human Immunodeficiency virus-1 RNA levels and CD4 lymphocyte counts, during treatment for active tuberculosis in South African patients. *Journal of Infectious Disease*, 187: 1967-1971.
25. Moustapha, M., Ndèye, S. S., Makhtar, C, Awa, Ba, Aliou, N., Géraldine, D., Djibril, W., Abdou, A., D., Maxim, T., Maïmouna, D., Nafissatou, L., Papa, A., D., Souleymane, M., Luc, K., and Tandakha, N. D. (2013). HIV and Tuberculosis co-infection impacts T-cell activation markers but not the numbers subset of regulatory T-cells in HIV-1 infected patients. *African Journal of Laboratory Medicine*, 2(1): 76.
26. Mukhtar AA, Abdullah AA, Juliana UO (2014). HIV-Associated tuberculosis: A sub-saharan african perspective. *Sub-Saharan African journal of Medicine*, 1(1): 1-14.
27. Ogbonnaya A, Manafa P, Chucks E, Okeke K, Alo M, Godwin O (2015). The prevalence of diabetes mellitus in human immunodeficiency virus-seropositive subject's co-infected with *Mycobacterium tuberculosis*. *Journal of AIDS and HIV Research*, 7(9): 109-116.
28. Patrizia A, Damien P, Klaus R, Francis M, Maxmillian M, Anneth T, Beatrice N, Hanspeter M, Stefanie K, Song D, Adam P, Fatoumatta D, Khalid O, Thomas JS,

- Giuseppe P, Claudia D, Matthieu P (2017). Mixed Th1 and Th2 *Mycobacterium tuberculosis*-specific CD4 T cell responses in patients with active pulmonary tuberculosis from Tanzania. 10.13-71. PLoS neglected tropical diseases, 11(7): e0005817.
29. Philana LL, and JoAnne LF (2015). CD8 T cells and *Mycobacterium tuberculosis* infection. Seminars in Immunopathology, 37(3): 239–249.
30. Prezzemolo T, Guggino G, La Manna MP, Di Liberto D, Dieli F and Caccamo N (2014). Functional signatures of human CD4 and CD8 T cell responses to *Mycobacterium tuberculosis*. Front Immunology, 5: 180.
31. Rodrigues D.S., Medeiro, E.A.S., Weckx, L.Y., Bonnez, W., Salomao, R. and Kallas, E.G. (2002). Immunophenotypic characterization of peripheral T lymphocytes in *Mycobacterium tuberculosis* infection and disease. Clinical and Experimental Immunology, 128(1): 149-154.
32. Rohan S and Indra DS (2015). Clinical Manifestation and Risk Factors of Tuberculosis Infection in Malaysia: Case Study of a Community Clinic. Global Journal of Health Science, 7(4): 110–120.
33. Shao L, Zhang X, Gao Y, Xu Y, Zhang S, Yu S, Weng X, Shen H, Chen ZW, Jiang W, Zhang W (2016). Hierarchy Low CD4+/CD8+ T-Cell Counts and IFN- γ Responses in HIV-1+ Individuals Correlate with Active TB and/or M.tb Co-Infection. PLoS ONE, 11(3): e0150941.
34. Silverberg, M.J., Leyden, W., Horberg, M.A., DeLorenze, G.N., Klein, D. and Quesenberry, C.P. (2007). Older age and the response to and tolerability of antiretroviral therapy. Archives of Internal Medicine, 167: 684–691.
35. Somily AM, Barry MA, Habib HA, Alotaibi FE, Al-Zamil FA, Khan MA, Sarwar MS, Bakhsh, ND, Alrabiaah AA, Shakoor ZA, Senok AC (2016). Evaluation of GeneXpert MTB/RIF for detection of *Mycobacterium tuberculosis* complex and rpo B gene in respiratory and non-respiratory clinical specimens at a tertiary care teaching hospital in Saudi Arabia. Saudi Medical Journal, 37(12): 1404-1407.
36. Swaminathan, S., Deivanayagam, C.N., Rajasekaran, S., Venkatesan, P., Padmapriyadarsini, C. and Menon, P.A. (2008). Long term follow up of HIV-infected patients with tuberculosis treated with 6 month intermittent short course chemotherapy. National Medical Journal of India, 21: 3-8.
37. Taha TE, Yende-Zuma N, Aizire J, Chipato T, Wambuzi OL, Makanani B, Chinula L, Nyati MM, Hanley S., Brummel SS, Fowler MG (2018). The multi-country PROMOTE

- HIV antiretroviral treatment observational cohort in Sub-Saharan Africa: Objectives, design, and baseline findings. *PLoS One*, 13; 13(12): e0208805.
38. Teresa P, Giuliana G, Marco Pio LM, Diana DL, Francesco D, Nadia C (2014). Functional Signatures of Human CD4 and CD8 T Cell Responses to *Mycobacterium tuberculosis*. *Frontiers in Immunology*, 7: 594.
39. World Health Organization (2016). Global tuberculosis report 2016. World Health Organization, Geneva, Switzerland.
40. Yang JD, Mott D, Sutiwisesak R, Lu Y-J, Raso F, Stowell B, Greg HB, Jinhee L, Steve MC, Sing SW, Sarah MF, Samuel MB (2018). *Mycobacterium tuberculosis*-specific CD4⁺ and CD8⁺ T cells differ in their capacity to recognize infected macrophages. *PLoS Pathogens*, 14(5): e1007060.
41. Yao, S., Huang, D., Chen, C. Y., Halliday, L., Wang, R.C. and Chen, Z.W. (2014). CD4⁺ T Cells Contain Early Extrapulmonary Tuberculosis (TB) Dissemination and Rapid TB Progression and Sustain Multieffector Functions of CD8⁺ T and CD3⁻ Lymphocytes: Mechanisms of CD4⁺ T Cell Immunity. *Journal of Immunology*, 192: 2120–2132.