

INVESTIGATION OF NEUROPROTECTIVE EFFECT OF *SAPINDUS MUKROOSI* EXTRACT ON TYPE1 DIABETE INDUCED NEUROPATHIC PAIN PERCEPTION IN RAT

Vandana Pokhriyal*, Neeraj Kumar, Pragati Bailwal, Vinita Chauhan, Pooja Negi, Subodh Negi and Abhishek Chauhan

*Division of Pharmaceutical Sciences, Shri Guru Ram Rai Institute of Technology and Science, Alpine College of Management and Technology, Dehradun, Uttarakhand, India.

Article Received on
20 Nov. 2019,

Revised on 11 Dec 2019,
Accepted on 01 Jan. 2020

DOI: 10.20959/wjpr20201-16299

*Corresponding Author

Vandana Pokhriyal

Division of Pharmaceutical
Sciences, Shri Guru Ram
Rai Institute of Technology
and Science, Alpine College
of Management and
Technology, Dehradun,
Uttarakhand, India.

ABSTRACT

Diabetic Neuropathy is a most common Microvascular diabetic complication, associated with neuropathic disorder causing nerve injury it mainly depends upon duration of hyperglycemia, elevation in glycosylated haemoglobin. Good Neuropathic pain perception can improve Neuropathy. Single intraperitoneal injection of STZ (60 mg/kg) was administered to the albino wistar rats induced Diabetes, it showed marked hyperglycemia. Hyperglycemia effect Glycated haemoglobin level in blood and reduce and reduce thermal hyperalgesia (Eddy's Hot plate, Tail flick), motor- coordination, as compared to the animals of control group. Effect of test drug Sapindus Mukroosi was assessed by behavioural and biochemical parameters (Serum Glucose Level, Lipid peroxidation, reduced Gluthathione) in the brain tissue of wistar rats. After 28 days treatment of Sapindus

Mukroosi with low (250 mg/kg) and higher (500 mg/kg) dose in diabetic rats showed antidiabetic action by reduction in hyperglycemia. Sapindus showed good result also in behavioural and biochemical parameters such as Thermal hyperalgesia, motor coordination in comparison with diabetic control group. The most promising effect of Sapindus seen with 500mg/kg after treating period. It significantly decrease serum glucose level, Glycated haemoglobin, lipid peroxidation and improving reduced glutathione level as compared to diabetic control rats. Thus Sapindus Mukroosi due to its antioxidant and antidiabetic property can be concluded as a effective or preventive treatment for Diabetic Neuropathy. Streptozotocin induce hyperglycemia and causes increase in oxidative stress and this

conditions are oppose by treatment with Sapindus Mukroosi by inhibit the generation of free radicals and improving self antioxidant GSH level significantly.

KEYWORDS: Diabetic Neuropathy, Glycated Haemoglobin, Neuropathic Pain.

INTRODUCTION

Diabetes mellitus is a metabolic disorder characterized by hyperglycemia resulting from defects in insulin secretion, insulin action or both. Diabetes mellitus is a serious health problem in developing as well as developed countries. The world health organization stated it as an epidemic disease of the 21st century, due to its rising population of 382 million in 2013, and May projected to rise up to 592 million by the year 2035. Uncontrolled diabetes leads to microvascular and macrovascular complications. The most common type of microvascular complication of diabetes is peripheral neuropathy. The prevalence rate is 7% in one year of freshly diagnosed diabetes and 50% in long standing diabetes history patients aged more than 25 years. In them 12% patients experience painful diabetic neuropathy with generating the symptoms of allodynia-stimulus that normally doesn't provoke pain, hyperalgesia increased the response to a non painful stimulus. Patients explain their symptoms as a sharp electric shock shooting their legs, and feeling of walking on broken glass. Hyperglycemia induces fatal changes in nerve tissue, a verity classical metabolic pathways; polyol pathway resulting in accumulation of sorbitol and glucose, increased hexosamine shunt; excess/inappropriate activation of protein kinase C isoforms; resulting in accumulation of advanced glycation end products; weaken neurotrophic support and disrupt the repair mechanism; and stimulation of poly (ADP-ribose) polymerase (PARP); result in imbalanced nerve myo-inositol. Although all of these pathways fatal in the generation of reactive oxygen species through oxidation and reduction reactions in mitochondria lead to aggravation of neuropathy.^[1,2,3,4]

Several drugs such as tricyclic antidepressants and anticonvulsant drugs are presently available to reduce the neuropathic pain. However, these drugs were reported to exhibit a wide spectrum of adverse effects in the management of painful neuropathy. Hence, there are a limited number of ideal medicines to treat diabetes neuropathy and its generating pain. Researchers, health care professionals and educated people acknowledge their interest towards new alternative medicines to treat diabetes associated neuropathy.^[5,6]

The present chosen trailing herb bearing a Latin name Sapindus Mukroosi (Sapindaecae) is a creeping herb grown in tropical regions of, India. Pharmacological studies on Sapindus fruit

pericarp were proven its antioxidant, antidiabetic, hepatoprotective, antifibrinolytic and anti-inflammatory activities in experimental animals, due to presence of various phytochemicals like flavonoids, amino acids, liriodendrons (lignans), β -sistosterol and stetracosanoic acid, ecosanoic, sterolic and urosolic acid have proven capacity to cure and control disease prognosis. Lack of scientific data on neuroprotective activity of *Sapindus Mukroosi* against diabetic neuropathy was gaining my attention towards this experiment.^[7,8]

MATERIAL AND METHOD

Plant Material

Collection and Authentication

Plant Material were collected from the wild area of Garhwal in the month of and authenticated from Botanical Survey of India (BSI), Dehradun. The specimen authentication No. is BSI\NRC Tech.\Herb (Ident.)\2016-17\830.

Method of Extraction The roots of *Boerhaavia diffusa* were collected and dried under shade and grinded into powder. Ethanolic extract of *Boerhaavia diffusa* roots was done in the department of Pharmacology, Geetanjali Medical College, Udaipur using cold maceration. The Fruits of *Sapindus Mukroosi* were collected and washed under running tap water for removal of dust. Pericarp (separated from fruits) and leaf was shade dried, oven dried at 40–45°C for 2h, and then grinded in mechanical grinder to make coarse powder. Aqueous extract of *Sapindus Mukroosi* pericarp was done in department of pharmacology S.G.R.R.I.T.S Dehradun using hot percolation extraction process.^[11,13]

Drugs and Chemicals: Streptozotocin was obtained from Sisco research laboratories Pvt. Ltd, Mumbai, India and Duloxetine was purchased from Swapnaroop drugs & pharmaceuticals, Aurangabad, Maharashtra, India. All other chemicals and reagents used were of analytical grade.^[15,17]

Procurement of Animals

Wistar rats weight 250-300gm of either sex was procured from the departmental animal house Department of Pharmaceutical Sciences of Shri Guru Ram Rai Institute of Technology and Sciences, Patel Nagar, Dehradun. The Animal were acclimatized in the departmental animal house facility and housed n=6 per cage under standard laboratory conditions 23±2°C with 12hr light dark cycle and had free accesses to water with standard chow diet. Animal care should be taken as per guidelines of the Committee for the Purpose of Control and

Supervision of Experiments on Animals (CPCSEA). Approval was taken from the Institutional Animal Ethics Committee (264/ReBi/2002/CPCSEA) for the study.

METHODOLOGY

Experimental Design

2 Experimental Design

5 groups each comprising of 6 animals

Group 1: Control: Normal Saline was administered as a Vehicle.

Group 2: Diabetic control: Streptozotocin (60 mg/kg) was administered.

Group 3: Active Control: Duloxetine was administered for 28 days in diabetic wistar rat as standard.

Group 4: Test Group: Diabetic neuropathy induced wistar rat + SME (250 mg/kg) was administered for 28 days.

Group 5: Test Group: Diabetic neuropathy induced wistar rat + SME (500 mg/kg) was administered for 28 days.

Streptozotocin induced diabetes diabetic neuropathy

Healthy wistar strain albino rats of either sex weighing about 150-200 grams were taken. Animals were deprived to food for 16 hours but allowed free access to water after that blood sample was collected from tail of rats and measure blood glucose level by using GOD-POD kit method. Then they were injected with streptozotocin dissolved in 0.1M sodium citrate and citric acid at a dose of 55 mg/kg body weight intraperitoneally. Then animals were kept for 21 days during which food and water was allowed. After 21 days of streptozotocin administration blood glucose level, body weight, grip strength and pain sensation measurements were taken. The animals showed fasting blood glucose level above 250 mg/dl considered diabetic rats and after that they were divided into five groups in which each group contained six animals. the blood glucose level, body weight, measurements of each rat were taken at the start and at the end of experiment. After the 4th week of the experiment administered the control, standard and test drug orally daily for 8 weeks.^[20,25]

Physical parameters

I. Grip-strength (Rota-rod) test

Grip strength used for evaluation of muscle strength during Diabetic Neuropathy. The test was being used to assess muscular strength or neuromuscular function with the help of rota rod apparatus in which the rats were placed on a horizontal rod rotating at a speed of 25 rpm.

The rats which were capable of remaining on the top for 25 sec or more, in three successive trials were selected for the study. The selected animals were divided into five groups (n= 6). The time of stay at the rod was calculated.^[26,27]

II. Tail Flick

All groups of animals were experimented for this test. Animals are placed into individual restraining cages leaving the tail hanging out freely. The animals are allowed to adapt to the cages for 30 min before testing. The lower 5 cm portion of the tail is marked and the tail of the rat was immersed in hot water maintained at $55\pm 0.2^{\circ}\text{C}$. The basal tail flick latency (withdrawal response of tail) or signs of struggle were observed. The cut off time was 10 sec.^[28,29]

III. Hot Plate Method

In this Hot Plate Method animals from the each group were placed on the hot plate which is commercially available, consists of an electrically heated surface. Temperature of this hot plate is maintained at $55-56^{\circ}\text{C}$ and observation is done up to the time until paw licking or jumping was noted the cut-off time was 10 sec. Then the average basal reaction time was noted after the oral administration of the drugs and test compounds.^[30,31]

Bio-chemical Parameters

I. Blood Glucose Level

The animal blood was collected from tail of rats for the determination of the blood glucose levels. The plasma was obtained after centrifugation (3000 g for 10 min, 4°C). Blood glucose was estimated by GOD-POD kit method.^[32,33]

II. Lipid Peroxidation Assay

This assay was used to determine thiobarbituric acid-reactive substances as described by Slater and Sawyer (1971). In 2.0 mL of the tissue homogenate (supernatant) was added 2.0 mL of freshly prepared 10% w/v trichloroacetic acid (TCA) and the mixture was allowed to stand in an ice bath for 15 min, followed by centrifugation at 2500 rpm for another 15 min at 4°C . Two milliliter of clear supernatant solution was mixed with 2.0 mL of freshly prepared 0.67% w/v TBA. The resulting solution was heated in a boiling water bath for 10 min. It was then immediately cooled in an ice bath for 5 min. The absorbance of colour developed was measured by UV/VIS double beam spectrophotometer (systraonic Japan) at 532 nm using 1, 1, 3, 3-tetraethoxypropane as a standard.^[34,36]

III. Estimation of Glutathione

The assay of GSH was determined by method described by Moron et al. (1979). One milliliter of tissue homogenate (supernatant) and 1.0 mL of 20% TCA were mixed and centrifuged at 2500 rpm for 15 min at 4°C. In 0.25 mL of supernatant, 2 mL of DTNB (0.6 M) reagent was added. The final volume was made up to 3 mL with phosphate buffer (pH 8.0). The colour developed was read at 412 nm against reagent blank. Different concentrations (10-50 µg) of standard glutathione were processed as mentioned above for constructing standard curve. The amount of reduced glutathione was expressed as µg of GSH /mg of protein.^[40,41]

IV. Estimation of Glycosylated Hemoglobin (Ion Exchange Method)

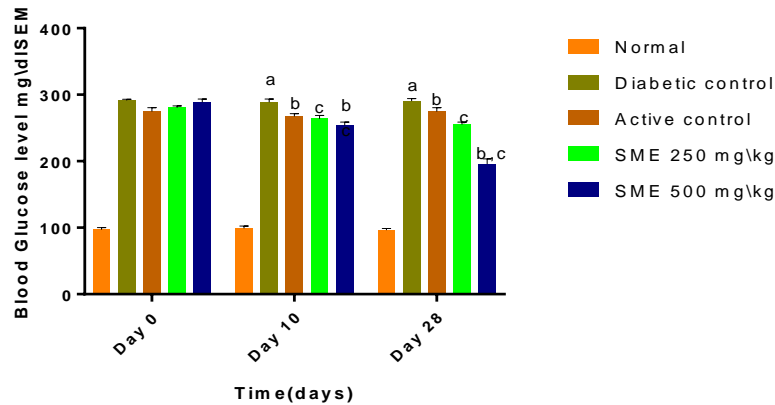
Whole blood is Mixed with lysine reagent to prepare a hemosylate. It then mixed with a weakly binding cation exchange resin. Resin bind with non-glycosylated hemoglobin and forming GHb free in supernatant. This percentage is examined by calculating the absorbance of fraction & of the total Hb level.^[42,44]

Statistical Analysis: The data obtained from the result was analyzed by using twp ways ANOVA followed by Bonferroni's post test using graph pad prism 7 software. All data were expressed as the mean SEM of their parameter.^[45,50]

RESULTS

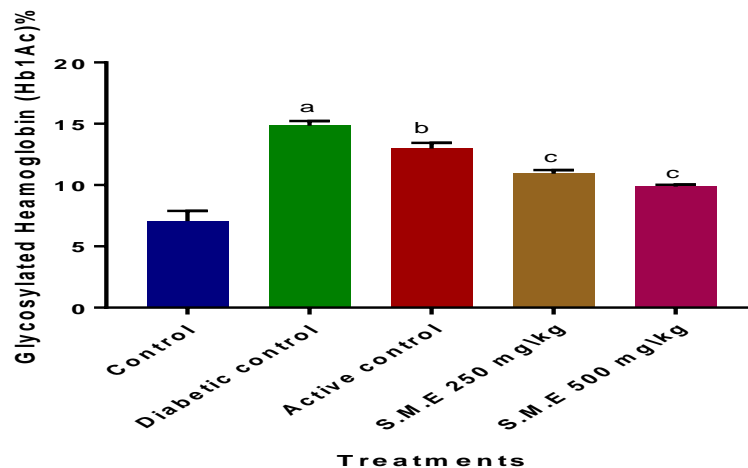
➤ Effect of Sapindus Mukroosi on serum glucose level in STZ induced diabetic rats

Serum glucose level was estimated on 0th, 7th, 14th, 28th, and 75th, days in all STZ treated groups and the result was found significant in comparison to control group. The diabetic rats were selected whose serum glucose level was found more than 250 mg\dl after administration of STZ. Sapindus Mukroosi in two different doses 250 mg\kg, 500 mg\kg was administered to diabetic rat and it was observed that the Serum Glucose level reduced as estimated after 10th and 28th days and effect was found significant.



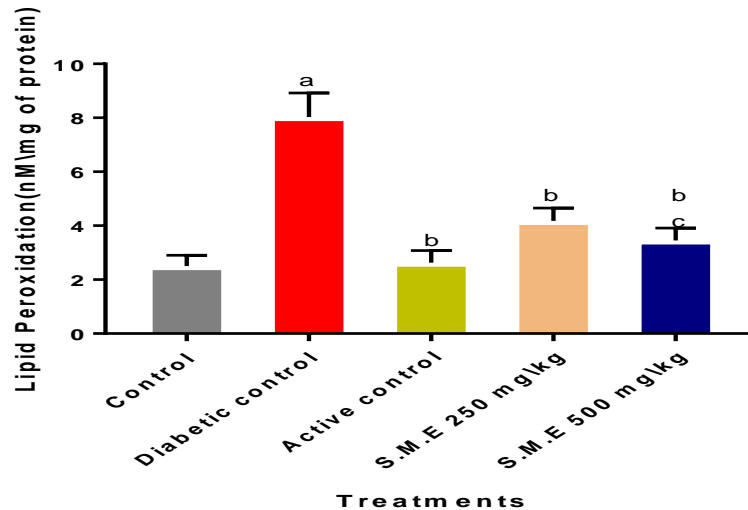
➤ **Effect of Sapindus Mukroosi on Glycosylated Hemoglobin in STZ induced diabetic rats**

Hyperglycemia produces a significant increase in Glycosylated Hemoglobin level in comparison with vehicle control group. However treatment with Sapindus Mukroosi significantly decreases Glycosylated Hemoglobin level and effect was found significant.



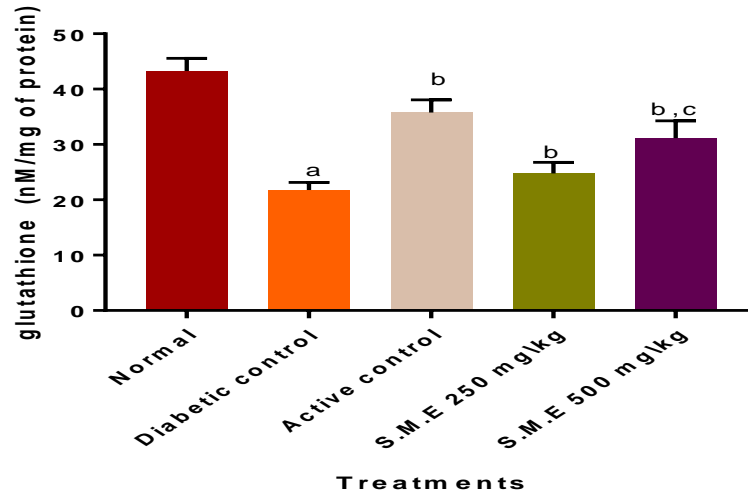
➤ **Effect of Sapindus Mukroosi on level of lipid peroxidation in STZ induced diabetic rats**

When STZ is compared with the control group the levels of MDA during lipid peroxidation in brain produced a significant difference ($p < 0.001$). Administration of SM (250 & 500 mg/kg) significantly reduces the lipid peroxidation level as compare to disease control. Standard treatment with Duloxetine significantly reduces the level of lipid peroxidation in brain.



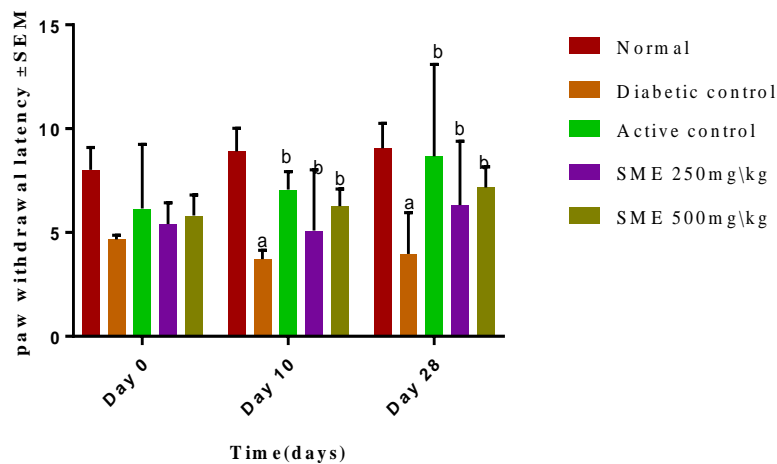
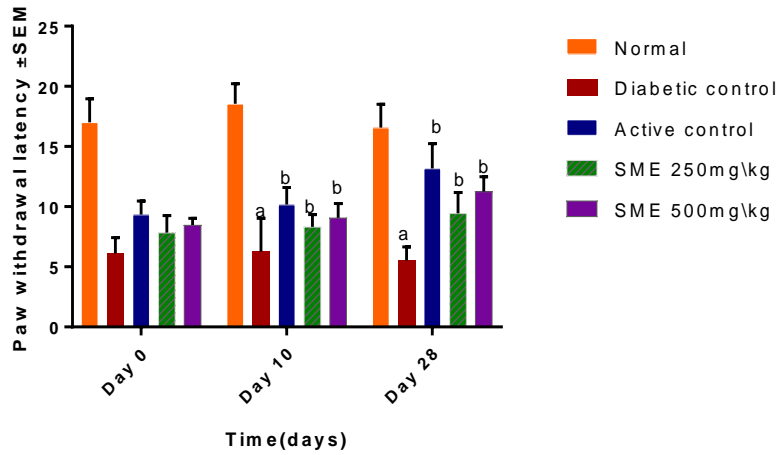
➤ **Effect of Sapindus Mukroosi on level of Gluthathione in STZ induced diabetic rats**

Administration of STZ produces a significant decrease in glutathione level in comparison with vehicle control group. However treatment with Sapindus Mukroosi significantly increases the glutathione level in brain. In other side the Standard treatment with Duloxetine significantly increase the level of glutathione.



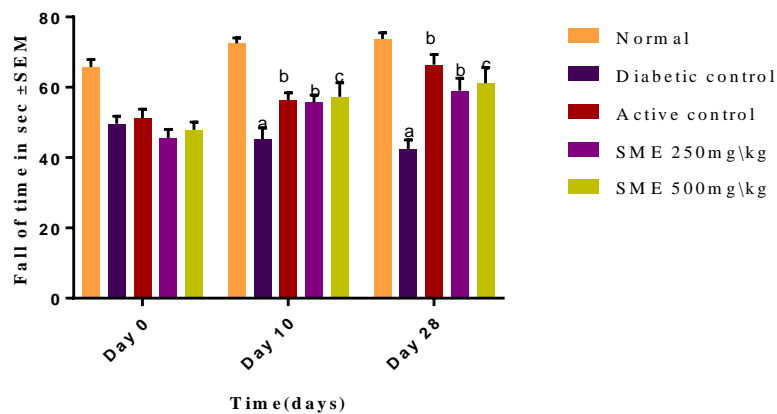
➤ **Effect of Sapindus Mukroosi on thermal allodynia\ hyperalgesia (Hot plate, tail flick) STZ induced diabetic rats**

Thermal Hyperalgesia test were performed 28 days after STZ administration and decrease in tail flick and paw withdrawal latency was observed. After Administration of Sapindus Mukroosi resulted in significant increase in tail flick and paw withdrawal latency on 28 days treatment as compared to Diabetic control group.



➤ **Effect of Sapindus Mukroosi on motor-coordination in STZ induced diabetic rats**

Motor coordination in Streptozotocin treated rat after 28 days STZ administration significantly decrease. Sapindus Mukroosi administration resulted in improved Motor coordination grip strength and positive effect was observed as compared to Standard drug (Duloxetine) treated group.



DISCUSSION

Diabetes mellitus (DM) is a group of metabolic disease which is characterized by persistent hyperglycemia, glycosuria, hyperlipidemia, and negative nitrogen balance and also frequently ketonaemia on account of defects in the insulin secretion and insulin action, or both of these leading to impaired functioning in the carbohydrate, lipid and protein metabolism which furthermore leads to increase fasting and postprandial blood glucose levels. Diabetic neuropathy is a multifaceted and possibly severe complication of diabetes affecting more than 50% of the diabetic individuals.

Present study showed rise in blood glucose level of STZ treated rats. The aqueous root extract of *Sapindus Mukroosi* (250, 500 mg/kg) showed significant reduction in serum glucose level and immobility time in pain perception test of diabetic rats. Antioxidant *Sapindus Mukroosi* extract improved brain glutathione level and diminished the lipid peroxidation and Glycosylated Hemoglobin level as compared to control group. It showed significant improvement in signs of neuropathy. Essentially the most profound effect was observed at the dose of 500 mg/kg of SME.

The experimental study was designed to evaluate the Neuroprotective effect of *Sapindus Mukroosi* in diabetic neuropathy. For this study STZ treated albino wistar rat were used to evaluate the effect of drug in Type 1 diabetes induce neuropathic pain perception.

CONCLUSION

The following finding was revealed in the study.

- *Sapindus Mukroosi* significantly reduces the symptoms of diabetic neuropathy and show the positive effect. Its antioxidant property reduced the blood glucose level as well as Glycated Hemoglobin & increase pain perception in Streptozotocin-induced diabetic rats to prevent progression of diabetic neuropathy.
- *Sapindus Mukroosi* showed most effect at a dose of 500 mg/kg on day 28 of treatment.
- STZ treated rats showed a significant increase in the level of lipid peroxidation which was reversed by the treatment with SME. The activity of antioxidants like GSH in STZ treated group is decreased, the activity is restored after treated with *Sapindus Mukroosi* by reducing generation of free radicals. So it can be concluded that SM used as ideal drug which could offer a better preventive option for diabetic neuropathy. This can act by, either preventing the nerve damage or by providing symptomatic pain control along with good glycemic control.

REFERENCES

1. Dipiro Joseph T. et al; Pharmacotherapy: A pathophysiologic Approach, New Delhi, McGRAW-HILL medical publishing division, sixth edition section eight chapter, 72: 1333 to 1364.
2. Marc Y. Donath¹, Jan A. Eshes¹, Kathrin Meadler¹, Desiree M. and Manfred Reinecke. Mechanisms of beta-cell death in Diabetes, Dec, 2005; 54(2): S108-S113.
3. American Diabetes Associatio. Diagnosis and Classification of Diabetes Melitus, *Diabetes Care.*, 2010; 33(1): 62-69.
4. Miriam Cnop, Nils Welsh, Jean-Christophe Jonas, et al. Eizirik Mechanism of Pancreatic –Cell death in Type 1 and Type 2 Diabetes Many Differences, Few similarities diabetes, 97-107.
5. Type1 Philip JC, Radermecker RP. Type 1 diabetes: from genetic predisposition to hypothetical enviroment tiggers. *Rev Med Liegw*, 2012; 67: 319-325.
6. Typ1 Knip M, SImell O. Envriornmental tiggers of type 1 diabetes. *CSH Perpect Med.*, 2012; 2: 007690.
7. Type 1 Godman gillman A. The Pharmacological basis of therapeutics, McGRAW-HILL ninth edition, 1487.
8. Type 2 American Diabetes Association. Diagnosis and Classification of Diabetes Melitus. *Diabetes Care.*, 2004; 27(1): s5-s10.
9. Type 2 Mogensen CE, Christensen CK, Vittinghus E. The stages in diabetic renal disease. with emphasis on the stage of incipient diabetic nephropathy. *Diabetes*, 1983; 32(2): J64-78.
10. Type 2 Kohei, K., et al. Pathophysiology of type 2 diabetes and its treatment policy. *Journal of Japan Medical Association*, 2010; 53(1): 41-46.
11. Comp Konstantinos, P, Maciej, B, Michael E, Nikolaos P. Complications of diabetes. *Journal of Diabetes Research*. 2015 Article ID 189525, 1-5.
12. Comp Michael J. Fowler, MD. Microvascular and Macrovascular complications of Diabetes. *Clinical Diabetes*, 2008; 26(2): 77-81.
13. Comp Chiarelli F, Marcovecchio ML. The molecular mechanism underlying diabetic complications. *International Journal of pediatric endocrinology*, 1: 14- 17.
14. Micro Kaul K. Inflammation & microvascular complications, 2014; 6(5): 294-303.
15. Micro Vithian K, Hurel S. Microvascular complications: Pathophysiology & Management, *Cilinal Medicine*, 2010; 10(5): 505-509.

16. Macro Khawlani A, et al. Macrovascular complications and their associated risk factors in diabetes patients in yemen. *Estern mediterrasseach health journal*, 2010; 16(8): 851-858.
17. DN. Vinik A, Mitchell I, Leichter S, Wagner AL., Brian J, Georges L.P. Epidemiology of the Complications of Diabetes. In: Leslie RDG, Robbins DC (eds) *Diabetes: Clinical Science in Practice*. Cambridge University Press, Cambridge, 1995; 221287.
18. DN Narayan KM, Boyle JP, Thompson TJ, Sorensen SW, Williamson DF. Lifetime risk for diabetes mellitus in the United States. *JAMA*, 2003; 290: 1884–1890.
19. DN Barrett AM, Lucero MA, Le T, Robinson RL, Dwrkin RH, Chappell AS. Epidemiology, Public Health Burden and treatment of diabetic peripheral neuropathic pain. *A review Pain Med.*, 2007; 8(2): S50-S62.
20. DN Candrilli SD, Davis KL, Kan HJ, et al. Prevalence and associated burden of illness of symptoms of diabetic peripheral neuropathy and diabetic retinopathy. *Journal Diabetes Complications*, 2007; 21: 306-314.
21. clfJames LE, Andrea Mv, Hsinlin TC, Eva L F. Diabetic Neuropathy: Mechanism to management. *Pharmacology and therapeutics*, October, 2008; 120(1): 1-2.
22. clfMark Gostine, et al., Diabetic Neuropathy Study. *Practical Pain Management*.
23. clf Jatinderjot K K, Saini N. Pathogenesis of Diabetic Neuropathy. *International journal of Pharma Profession Research*, 2013; 4(1): 715-733.
24. pn Kanji JN, Anglin RE, Hunt DL, Panju A. Does this patient with diabetes have large-fiber peripheral neuropathy? *JAMA*, 2010; 303: 1526-32.
25. PNWatkins PJ. Pain and diabetic neuropathy. *Br Med journal*, 1984; 288: 168-169.
26. PNSugimoto K, Murakawa Y, Sima AA. Diabetic neuropathy—a continuing enigma. *Diabetes Metab Res Rev.*, 2000; 16: 408-33.
27. PNTesfaye S, Watt J, Benbow SJ, Pang KA Miles J, MacFarlane IA. Electrical spinal cord stimulation for painful diabetic peripheral neuropathy. *Lancet*, 1996; 348: 1696-1701.
28. AN S. Vuckovic- Rebrina, A et al., Diabetic Autonomic Neuropathy. *Diabetologia Croatica*, 2013; 42(3): 73-79.
29. AN Vinik AI, Maser RE, Mitchell BD, Freeman R. Diabetic autonomic neuropathy. *Diabetes Care*, 2003; 26: 155-179.
30. AN Toyry JP, Niskaner LK, Mantysaari MJ. Occurrence predictors and clinical significance of autonomic neuropathy in NIDDM: Ten years follow up from diagnosis. *Diabetes*, 1996; 45: 308-15.

31. Kempler P, Tesfaye S, Chaturvedi N, et al. The prevalence of autonomic neuropathy and potential risk factors: The EURODIAB IDDM Complications study. *Diabetic Med.*, 2002; 19: 900-909.
32. Thomas Frieling et al. Diabetic Autonomic Neuropathy of the gastrointestinal tract. *Wolters Kluwer*, 2015; 2: 10-14.
33. J G Williams. Autonomic Neuropathy in diabetes: A Review. *Journal of Royal Society of Medicine*, 1983; 73: 502-507.
34. PrN Barohn RJ, Sahenk Z, Warmolts JR. The Bruns-Garland syndrome (diabetic amyotrophy). *Arch Neuro*, 1990; 48: 1130.
35. Pr N Diabetic neuropathies: the nerve damage of Diabetes [internet] 2009 [updated 2012] June 25]. Available from <http://diabetes.niddk.nih.gov/dm/pubs/neuropathies>
36. FN Medecin. Consultantly Focal And Multifocal Diabetic Neuropathie. *Neuro-Psiquiatr*, 2007; 65(45b).
37. FN Chance PF. Inherited focal, episodic neuropathies: hereditary neuropathy with liability to pressure palsies and hereditary neuralgic amyotrophy. *Neuromolecular Med.*, 2006; 8: 159-74.
38. CrGoldstein JE, Cogan DG. Diabetic ophthalmoplegia with special reference to the pupil. *Arch Ophthalmol*, 1960; 64: 592-600.
39. CrDreyfuss PM, Hakim S, Adams RD. Diabetic ophthalmoplegia. *Arch Neurol Psychiatry*, 1957; 77: 337-349.
40. RediEllenberg M. Diabetic Truncal mononeuropathy- a new clinical syndrome. *Diabetes Care*, 1978; 1: 10-13.
41. FN RediBoulton AJM, Augur E, Ayyer DR. Diabetic thoracic polyradiculopathy presenting as an abdominal swelling. *BMJ*, 1984; 289: 798-9.
42. Patho Malik ZA, Tabassum N, and Sharma PL. Attenuation of experimentally induced diabetic neuropathy in association with reduced oxidative nitrosative stress by chronic administration of *Momordica charantia*. *Advances in Bioscience and Biotechnology*, 2013; 4: 356-363.
43. Polyol Brownlee M. The pathobiology of diabetic complications. *Diabetes*, 2005; 54(6): 1615-1625.
44. Polyol Alter ML, Ott IM, von Websky K, Tsuprykov O, and Sharkovska Y. DPP-4 inhibition on top of angiotensin receptor blockade offers a new therapeutic approach for diabetic nephropathy. *Kidney Blood Pressure Research*, 2012; 36: 119-130.

45. Polyol Yamagishi S, Maeda S, Matsui T, Ueda S, and Fukami K. Role of advanced glycation end products (AGEs) and oxidative stress in vascular complications in diabetes. *Biochemistry Biophysics*, 2012; 1820: 663-671.
46. Polyol Gabbay KH, Merola LO, Field RA. Sorbitol pathway: presence in nerve and cord with substrate accumulation in diabetes. *Science*, 1996; 151: 209-210.
47. Polyol Ward JD, Baker RWR, Davis B. Effect of blood sugar control on the accumulation of sorbitol and Fructose in nervous tissue. *Diabetes*, 1972; 21: 1173-1178.
48. Advance Edwards JL, Vincent AM, Cheng HT, and Feldman EL. Diabetic neuropathy: mechanisms to management. *Pharmacology Therapeutics*, 2008; 120: 1-34.
49. Advanc Ryle C and Donaghy M. Non-enzymatic glycation of peripheral nerve proteins in human diabetics. *Journal of Neurology Science*, 1995; 129(1): 62-8.
50. Advanc Ahmed N. Advanced glycation end products- role in pathology of diabetic complications. *Diabetes Research Clinical Practice*, 2005; 67: 3-21.
51. Pkc Arikawa E. et al., Effects of insulin replacements, inhibitors of angiotensin, and PKC beta's actions to normalize cardiac gene expression and fuel metabolism in diabetic rats. *Diabetes*, 2007; 56(5): 1410-1420.
52. Proteinkinase Koya D, King GL. Protein kinase C activation and the development of diabetic complications. *Diabetes*, 1998; 47: 859-866.
53. Protein Cameron NE, Cotter MA, Lai K, Hohman TC. Effects of protein kinase C inhibition on nerve function, blood flow and Na, K, ATP defects in diabetic rats. *Diabetes*, 1997; 46(A): 31.
54. Hexo Thornalley PJ. The potential role of thiamine (vitamin B (1)) in diabetic complications. *Current Diabetes Review*, 2005; 1(3): 287-298.
55. Hexo Kolm-Litty V, Sauer U, Nerlich A, Lehmann R, and Schleicher ED. High glucose-induced transforming growth factor beta1 production is mediated by the hexosamine pathway in porcine glomerular mesangial cells. *Journal of Clinical Investigation*, 1998; 101(1): 160-169.
56. ADP Obrosova IG. et al., Oxidative–nitrosative stress and poly (ADP-ribose) polymerase (PARP) activation in experimental diabetic neuropathy: the relation is revisited. *Diabetes*, 2005; 54(12): 3435-3441.
57. ADP Ilnytska O. et al., Poly(ADP-Ribose) Polymerase Inhibition Alleviates Experimental Diabetic Sensory Neuropathy. *Diabetes*, 2006; 55: 1686-1694.

58. Oxidative stress Cameron NE, Cotter MA. The relationship of vascular changes to metabolic factors in diabetes mellitus and their role in the development of peripheral nerve complication.
59. Stress Kitto KF, Haley F, and Wilcox GL. Involvement of Nitric oxide in spinally mediated hyperalgesia in the mouse. *Neuro Sci Lett*, 1992; 77: 227-9.
60. Stress Kim HK. et al., Reactive Oxygen species (ROS) plays an important role in a rat model of neuropathic pain. *Pain*, 2004; 11: 116-124.
61. Stress Naik AK. et al., Role of oxidative stress in pathophysiology of peripheral neuropathy and modulation of N-acetyl-L-cysteine in rats. *Eur J Pain*, 2006; 10: 573-579.
62. TCA Collins SL, Moore A, McQuay HJ, Wiffen P. Antidepressants and anticonvulsants for diabetic neuropathy and postherpetic neuralgia: a quantitative systemic review. *J Pain Symptom Manage*, 2000; 20: 449-458.
63. ED Pirart J. Diabetes mellitus and its degenerative complications: a prospective study of 4,400 patients observed between 1947 -1973. *Diabetes Care*, 1978; 1: 168.
64. QS Khatrak SM. Diabetic peripheral neuropathy. CME. 13th UP Neurocon. *Dept of Neurology. Banaras Hindu University*, 2004: 164-70.
65. Treat Saad Javed, Ioannis N. Petropoulos, Uazman Alam and Rayaz A. Treatment of painful diabetic neuropathy. *Therapeutic Advances in Chronic Disease*, 2015; 6(1): 15 –28.
66. TCA Nash TP. Treatment options in painful diabetic neuropathy. *Acta Neurologica Scand*, 1999; 173: 36-42.
67. TCA Rang HP, Dale MM, Ritter JM, Simmons B, Hotta N, Greene DA, Ward JD. The central nervous system. *Pharmacology, 3rd edition. London: Churchill Livingstone*, 1998: 509-511.
68. Ssri Goldstein, D., Lu, Y., Detke, M., Lee, T. and Iyengar, S. Duloxetine vs. placebo in patients with painful diabetic neuropathy. *Pain*, 2005; 116: 109–118.
69. Ssri Tavakoli, M., Mojaddidi, M., Fadavi, H. and Malik, R. Pathophysiology and treatment of painful diabetic neuropathy. *Curr Pain Headache*, 2008; 12: 192–197.
70. NMDA Sang, C., Booher, S., Gilron, I., Parada, S. and Max, M. Dextromethorphan and memantine in painful diabetic neuropathy and postherpetic neuralgia: efficacy and dose—response trials. *Anesthesiology*, 2002; 96: 1053–1061.
71. NMDA Shaibani, A., Pope, L., Thisted, R. and Hepner, A. Efficacy and safety of dextromethorphan/ quinidine at two dosage levels for diabetic neuropathic pain: a double-blind, placebo-controlled, multicenter study. *Pain Med.*, 2012; 13: 243–254.

72. Mexil Jarvis B., Coukell A. J., Mexiletine. A review of its therapeutic use in painful diabetic neuropathy. *Drugs*, 1998; 56(4): 691-707.
73. Mexel Gimbel J. S., Richards P., Portenoy R. K. Controlled-release oxycodone for pain in diabetic neuropathy: a randomized controlled trial. *Neurology*, 2003; 60(6): 927-923.
74. Anticon Lessar H, Sharma U, Poole RM. Pregabalin relieves symptoms of Painful diabetic neuropathy: *Neurology*, 2004; 63: 2104-2110.
75. Lant Watson C. P., Moulin D., Watt-Watson J., Gordon A., & Eisenhoffer, J. Controlled trial in painful diabetic neuropathy. *Pain*, 2003; 105(2): 71-78.
76. Cpa Zhang, W. and Li Wan Po, A. The effectiveness of topically applied capsaicin. A metaanalysis. *Eur J Clin Pharmacol*, 1994; 46: 517-522.
77. Lidno Barbano, R., Herrmann, D., Hart-Gouveau, S., Pennella-Vaughan, J., Lodewick, P. and Dworkin, R. Effectiveness, tolerability, and impact on quality of life of the 5% lidocaine patch in diabetic polyneuropathy. *Arch Neurol*, 2004; 61: 914-918.
78. Phenolic Huang S.M; Chuang, H.C.; Yen, G.C. Cytoprotective effects of phenolic acids on methylglyoxal-induced apoptosis in Neuro-2A cells. *Mol Nutr Food Res.*, 2008; 52: 940-949.
79. Oil Khali, H. Painful diabetic neuropathy management. *Int. J. Evid. Based Healthc*, 2013; 11: 77-79
80. RF Yadav R, Tiwari P, Dhanraj E, Risk Factors and diabetic complication in diabetes in *Asian. CRIPS*, 9(2): 8-12.
81. Risk factor Tammy AM J.Lindsay, MD, Blakec. Rodgers J. Treating Diabetic Peripheral Neuropathic Pain. *Diabetes*, 2010; (82): 127-155.
82. Risk factor V Bansal, Jkalita and UK Mishra. Diabetic Neuropathy. *Medical Journal*, 2006; 82(964): 95-100.
83. Spindus Mukroosi.com- The Complete Soap nut Guide. 2009 <http://www.spindusmukroosi.com>
84. SM Shital M, Sonawane H. A Review of Recent and Current Research Studies on the Biological and Pharmacological Activities of Sapindus Mukorossi. *International Journal of Interdisciplinary Research and Innovations*, 2015; ISSN 2348-1226; 3(4): 85-95.
85. RL Modi D, Gunvanti B. Rathod K. N, Goswami M. Study of significance of glycosylated hemoglobin in diabetic patient. *International Archives of Integrated Medicine*, 2016; 3(4): 9-15.

86. RL Gangwa AK. Investigation of Neuroprotective effect of rasagiline in diabetic neuropathy in Streptozotocin induced type 2 diabetic rats. *European journal of pharmaceutical and Medical research*, 2016. ISSN 2394-3211: 469-475.
87. RL Ravikant, Verma AK, Shirivastava P. Effect of cocciniaindica leaf extract on diabetic neuropathy pain in rats. Department of pharmacology R.V Northland Institute, Dadri, Greater Noida India *European journal of pharmaceutical and medical research*, 2016. ISSN 3294-3211, 415-420.
88. RL Mishra S, Thakur M. Role of Microwave assisted Extraction for Isolation of Saponins from Sapindus Mukroosai and Synthesis of its stable Biofunctionalized silver Nanoparticles and its Hypolipidaemic Activity. *International Journal of Pharmaceutical Sciences and Reasearch*, 2016; 7(7): 2959-2965.
89. RL Kumar KS, Tahashildar J, Kota K. Neuroprotective effect of ethanolic root extract of Boerhaavia diffusa (Linn.) against Streptozotocin induced Diabetic neuropathy in animal model. *Journal of Chemical and Pharmaceutical Research*, 2016; 8(3): 831-840. Article ISSN NO:0976-6723
90. RL Roman Adhikari, Jyoti Y, Deepika Bora. Combined effect of aqueous extract of curcuma longa linn. With metformin in diabetes induced neuropathic pain in rats. Department of Pharmacology, Krupanidhi College of Pharmacy. *Asian J Pharm Clinical Reasearch*, 2015; 8: ISSN-0974-2441: 166-170.
91. RL Shahbaj Khan, Hardeep Kaur, Gopal Sharma, Sonu Role of Various Mechanisms and Pathways in Diabetic neuropathy: An Overview. *International Journal of Pharmaceutical Sciences Letters*, 2015; 5(1): 495-500.
92. RL Reetika Singh, Nishi Kumari* Comparative determination of phytochemicals and antioxidant activity from leaf and fruit of Sapindus mukorrossi Gaertn. – A valuable medicinal tree. *Elsevier Industrial Crops and Products*, 2015; 73: 1–8.
93. RL Yoseph Charinet Megerssa, Demo Yamane Tesfaye. Glycated Protein: Clinical utility and analytical Approaches. *African journal of Biochemistry Reasearch*, 2015; 9(2): 18-25.
94. RLSaad Javed, Ioannis N. Petropoulos, Uazman Alam and Rayaz A. Malik. Treatment of painful diabetic neuropathy. *Therepeutic Advance Chronic Disease*, 2015; 6(1): 15–2.
95. RL Sachin G, Dileep K, Gopal M, Shivali S, Medicinal Plants of the Genus Sapindus (Sapindaceae)- A Review of their Botany Phytochemistry biological Activity and Traditional Uses. *Journal of Drug Delivery & Therapeutics*, 2014; 4(5): 7-20.

96. RLShilpashree Y. D. et al, Appraisal of oxidative stress markers and antioxidant status in diabetic neuropathy. *International Journal of Recent Trends in Sciences and Technology*. 2014, Volume 11, ISSN 2277-2812 E-ISSN, 2249-8109; 11(3): 313-315.
97. Jatinderjot Kaur Kehal, Saini Neha, PATHOGENESIS OF DIABETIC NEUROPATHY. *International Journal of Pham Professional and Research*, 2013; 4(1): 15-25.
98. RLVerma N, Amresh G, Sahu PK, Neelam Mishra N, Singh AP Ch V Rao3. : Antihyperglycemic activity, antihyperlipedemic activity, haematological effects and histopathological analysis of Sapindus mukorossi Gaerten fruits in streptozotocin induced diabetic rats. *Asian Pacific Journal of Tropical Medicine*, 2012; 518-522.
99. G. Sudhakara, B, Ramesh2, P, Mallaiah. Protective effect of ethanolic extract of commiphoramukul gum resin against oxidative stress in the brain of STZ induced diabetic wistar ras. *Excli journal*, 2012; ISSN 1611-2156: 576-592.
100. Faisal Mohd. The Pharmacological Evaluation of Epigallocatechin-3-Gallate (EGCG) Against Diabetic Neuropathy in Wistar Rats. *International Journal of Scientific Research and Reviews*, 2012; 1(3): 75-87.
101. Sachdev Yadav, Mayank Kulshrestha, Mradul Goswami and Veena Sharma. "Elucination of Analgesic and Antipyretic activities of *Ficus bengalensis* linn. Leaves in Rats" *Journal of applied pharmaceutical science*, 2011; 1(01): 38-41.
102. Heltianu C, Guja C. Role of Nitric Oxide Synthase Family in Diabetic Neuropathy. *Diabetes and metabolism An open excess*, 2011; S-5 3-7.
103. RL TEMBHURNE S.V, SAKARKAR D.M. Effect of Fluoxetine on an Experimental Model of Diabetes-induced Neuropathic Pain Perception in the Rat. *Indian Journal of Pharmaceutical Sciences*, 2011; 73(6): 621-625.
104. RL T Mixcoatl- Zacuati, CG Jolivalt. A Spinal Mechanism of action for Duloxetine in a rat model of Painful Diabetic Neuropathy. *British Journal of Pharmacology*, 2011; 164(1): 159-169.
105. RL Tammy A M, J. Lindasy, MD, Blakec. Rogers J. Treating Diabetic Pheripheral Neuropathic Pain., 2010; 82: 127-155.
106. James W, Russell, alisonberent-spillson, Andrea M, Vincent, Freimann C. Oxidative Injury and Neuropathy in Diabetes and impaired Glucose Tolerance. *NIH Public Access, Neurobiol Dis.*, 2008; 30(3): 420-429.
107. RL Williams, S., Lindau, S.T. Dried Blood Spot Measurement of Glycosylated Hemoglobin (HbA1c). National Social Life, Health & Aging Project (NSHAP), 2008.

108. RL Geeta A Khwaja, Choudhary N. Current and emerging therapy for Diabetic Neuropathies. *Indian Academy of Clinical Medicine*, 2007; 8(1): 53-64.
109. RLThaifa MS, Roshna S. A Review on Diabetes melitus and Diabetic Neuropathy: A Plant based approach. *Journal of Pharmacognosy and Phytochemistry*, 2007; 6(3): 506-510.
110. RL Smith T, Robert A Nicholson. Review of duloxetine in the management of diabetic peripheral neuropathic pain. *Vascular Health Risk Management*, 2007; 3(6): 833–844.
111. Bansal V, Kalita J and Mishra UK. Diabetic Neuropathy. *med Journal*, 2006; 82(964): 95-100.
112. RL Irina G. Obrosova, Viktor R, Drel. et al., Oxidative- Nitrosative Stress and Polyol (ADP- Ribose) Polymerase (PARP) Activation in Experimental Diabetic Neuropathy: *NH Public Access Diabetes*, December, 2005; 54(12): 3435-3441.
113. Booya F, Bandarian F, Larijani B, Lotfi J. Potential risk factors for diabetic Neuropathy: A case control study. *Bio Med Central Neurology*, 2005; 5-24.
114. Davia R, Cornblath M.D. Diabetic Neuropathy: Diagnostic method Advance study in Medicine. *Diabetes*, 2004; 4 8(A): S650-S661.
115. Yamagishi S, Uehara K, Otuski S, Yagihashi S, Differential influence of increased Polyol pathway on protein kinase C expressions between endoneurial and epineurial tissues in diabetic mice. *Journal of neurochem*, 2004; 87(2): 497-507.
116. Ann M. Schmeichel, Schmelzer James D, Philip A. Oxidative injury and Apoptosis of Drosal root Ganglion Neurons in Chronic Experimental Diabetic Neuropathy, *American Diabetes Association*, 2003; 52(1): 165-171.
117. RL Thomas P.K. Diabetic Neuropathy: Models, Mechanisms and Mayhem. *LE JOURNAL CANADIEN DES SCIENCES NEUROLOGIQUES*, 1992; 19: 1-7.
118. Citrate Dharendra Singh anti diabetic effect of hydroalcoholic extract of Ficus palmate forsk leaves in Streptozotocin induced diabetic rats. *International Journal of Green Pharmacy*, 2014; 8(4): 240-256.
119. Induc Tulip group, India. Glucose kit, Available at www.tulipgroup.com
120. Indu Himeno T Pathan R Asif, Viswanad Bhoomi, K Swapnil, Chronic administration of pigiltazone attenuates intracerebroventricular Streptozotocin induced- memory impairment in rats. *Life Sciences*, 2006; 79: 2209-2216.
121. STZ Kandhare Amit. D et al., Neuroprotective Effect of Naringin by Modulation of Endogenous Biomarkers in Streptozotocin Induced Painful Diabetic Neuropathy *Fitoterapia*, 2012; 9: 21-34.

122. STZ JR Zurek. et al., Spinal And Supraspinal Components Of Opioid Antinociception In Streptozotocin Induced Diabetic Neuropathy In Rats. *Pain.*, 2001; 90(1): 57-63.
123. BGL Ebuehi O.A.T, Dibie D.C. Hyperglycemic Effect on Brain Cholinergic Functions, Oxidative stress and protein Expression of Brain Derived neuropathic Factor (Bdnf) on Cognitive Functions in Streptozotocin Induced-Diabetic Rats. *Research in Neuroscience*, 2015; 4(1): 1-9.
124. BGL Vareniuk Igor, Ivan A. et al., Nitrosative Stress and Peripheral Diabetic Neuropathy In Leptin Deficient (Ob/Ob) Mice. *Experimental Neurology*, 2007; 205: 425–436.
125. LP Sharma M, Katyal T, Grewel G, and Behera D. & Budhiraja R. D Effect of antioxidants such as carotene, vitamin C and vitamin E, on oxidative stress, thermal hyperalgesia and cold allodynia in stz induced diabetic rats. *The Internet Journal of Pharmacology*, 2009; 6(2): 67-79.
126. LP Ohkawa H, Ohisi N and Yagi K et al. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal Bio-chemistry*, 1979; 95: 351-358.
127. GSH Misra, H.P. Biochemistry edition, 1967; 15: 681.
128. GSH Zafar Ahmad Malik, Nahida Tabassum, Pyare Lal Sharma. Attenuation of experimentally induced diabetic neuropathy in association with reduced oxidative nitrative stress by chronic administration of Momordica charantina. *Advance in Biosciences and Biotechnology*, 2013; 4: 356-363.
129. Cold Boulton AJ, et al., Diabetic Neuropathies: A statement by the American Diabetes Association. *Diabetes Care*, 2005; 28: 956-962.
130. Hot plate Tammy AM, J. Lindsay, MD, Blakec. Rogers J. Treating Diabetic Pheripheral Neuropathic Pain, 2010; 82: 127-155.
131. Hot As Shaikh et al. Animal Models And Biomarkers Of Neuropathy In Diabetic Rodents. *Indian Journal of Pharmacology*, 2010; 42(3): 129-134.
132. Tail flick Yadav S et al. Elucidation of Analgesic and Antipyretic Activities Of Ficus Bengalensis Linn. Leaves in Rats. *Journal of Applied Pharmaceutical Science*, 2011; 1(1): 38-41.
133. Grip JaNosSzolcsaNyi. et al., Analgesic Effect Of Tt-232, A Heptapeptide Somatostatin Analogue, In Acute Pain Models Of The Rat And The Mouse And In Streptozotocin-Induced Diabetic Mechanical Allodynia. *European Journal of Pharmacology*, 2004; 498(3): 103–109.

134. Grip Kelli. A. Sullivan. et al., Mouse Models Of Diabetic Neuropathy. *Neurobiology of Disease*, 2007; 28(1): 276–285.
135. Anne K Schreiber, Carina FM Nones, Mcunha J. Diabetic Neuropathic Pain: Physiopathology & Treatment. *World Journal of Diabetes*, 2015; 1-20.
136. Discussion Bunn H.F., et al. The glycosylation of hemoglobin: Relevance to diabetes mellitus. *Science*, 1978; 200(7): 21-27.
137. Discus Ursula T., et al. Three assays for Glycosylated hemoglobin compared. *Clinical chemistry*, 1995; 41(2): 191-195.
138. Discuspain D'Amour WL, Smith DL. A method for determining loss of pain sensation. *Journal of Pharmacology Experiment Therapeutics*, 1941; 72: 74.
139. Discus hot tail flick Sawynok J, Reid AR, Esser MJ. Peripheral antinociceptive actions of desipramine and fluoxetine in an inflammatory and neuropathic pain test in the rat. *Pain*, 1999; 82: 149-55.
140. Discus neuro symMark AR. Neuropathies associated with diabetes. *Medicinal Clinical North America*, 1993; 27: 111-124.
141. rfPapanas N, Ziegler D. Risk factor and comorbidites in diabetic neuropathy: An update. *Society for Biomedical Diabetes Research*, 2015; 12(1): 48-62.
142. rfJeremiah John Duby, Reith Cambell, Stephen M. Setter, John Raymond White, Kiristin. A Diabetic Neuropathy: An Intensive Review. *Am Journal Health system Pharm.* 2004; 61(2): 2-5.