

TRANSFERSOMES: A NOVEL VESICULAR CARRIER FOR EFFECTIVE TRANSDERMAL DRUG DELIVERY (A REVIEW)

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ABSTRACT

Due to the ability of lipid vesicles to bypass barrier characteristics of the skin, they have been used as topical drugs in recent years. The new transdermal supply network is more favourable than conventional supply systems. Elastic vesicles are transfers that deliver great distortion. They are widely used in bioactive formulations in transdermal supplies. Several methods can be applied, including iontophoreses, electrophoresis, sonophoresis etc., to increase transdermal penetration. Some other types of those systems must include vesicular system and microneedles (liposomes, noisomes etc.)

The thin constriction of the transfer ranges (5-10 times lesser diameter, than their own) distorts and passes with no noticeable loss. The transferable defect function increases the permeation of the medication. These can be a carrier for low or high weight medications like antifungals, anaesthetics, corticosteroids, etc. These can also act as anaesthetics. The inside of the cell lipid of the stratum cornea will easily penetrate the skin. They move on to a deeply hydrated stratum according to an osmotic gradient, following application of Transfersomes on skin. The presence of surfactants in the structure assists to resolve the lipid in the cornea stratum and allows for high vesicular penetration. This study aims to form and evaluate penetration of the medicine and enhances its anti-fungal activity by using Sertaconazole nitrate transfersomes. The results of the study are based on the percentage of surfactants. Dermatophytosis is used for the efficacy test as a model dummy disease.

KEYWORDS: Ultra deformable vesicles; transfersomes; permeability; antifungal activity.

INTRODUCTION

The definition and the word “transfersome” were termed by Gregor Cevc in 1991. A transfersome is, in the widest sense, a dynamic and adaptable aggregate. A twisted vesicle with an aquatic centre and a complex lipid bilayer are ideally designed. This allows the transferee to effectively overcome numerous transport barriers and to be a carrier for the non-invasive delivery of medicines and for the continued release. The vesicle is autostatative and optimizing, based on local composition and the form of the bilayer, to provide medicines. The transdermal path is an interesting alternative since there is a simple and protected transdermal penetration.

[Figure 1: About Here]

This provides various potential benefits over conventional pathways, such as preventing metabolism of first-pass, stable and prolonged activity, reducing unwanted side effects, short-term effectiveness of medicinal products, increasing physiological and medication sensitivity, preventing variation of drug level and, more significantly, it offers a broad range of possible advantages over convulsion.

[Figure 2: About Here]

Actually, various chemical and physical techniques are applied, including liposomes and proliposomes, and non ionic surfactant vesicle (niosomas and pro-niosoms), to increase materially efficient transmission through the skin, using penetration enhancers, iontophoresis, sonophoresis, and colloidal pores. Transfersomes have been designed to benefit from phospholipids as transdermal pharmaceutical carriers. Such self-optimized aggregate systems aid with the extremely versatile membrane structure to penetrate the product either in or through the skin, depending on the way it is applied or used.

Such vesicular transfers are more versatile and thus ideal for skin penetration than regular liposomes in many orders of magnitude. Transfersomes overcome skin penetration problems by rubbing the inside corneal lipid. The entry is permitted in a self-adaptation way due to the mechanical stress of the system because of the high vesicle deformations. The mixture of an appropriate amount of surface active constituents and the polyvalence of the membrane of transfersomes will be in reasonable ratios.

The multiple capability of the vesicle in the skin is reduced, allowing transferomas to track the natural water gradient in nonocclusive conditions over the epidermis. The vesicle can break down completely. The intact stratum can spontaneously penetrate the intracellular lipid via two routes distinguished by the characteristics of its two layers (Schatzlein et al 1995). The following figure illustrates potential drug pathways for human skin intracellular and transcellular.

The transferomes are able to adapt to the environmental stress compared to the typical composite membrane because they are highly and autonomously deformed. When pressed or drawn to a small pore, the membrane structure is changed locally and reverse. The highly adaptable transfersome molecules sustain high membrane distortion and display high accumulation, while at the places of intense stress, the less stable molecular is diluted. This reduces the energy costs of membrane deformation considerably and enables highly flexible particles to enter the pores and then move through them quickly and proficiently.

In order to treat drug molecules with broad solubility, transfersomes have an infrastructure containing hydrophobic and hydrophilic molecules. Transfersomes twist without detectable failure and go through small constraints (5 to 10 times less than their own diameters). This high deformation gives the intact vesicles a greater penetration. It may be a carrier of medications with low molecular weight in addition to high molecular weight medications, such as anaesthetics, pain reliever, corticosteroids, sex-hormones, anti-neoplastic agents, leptins, cross-protein components and albumin etc. Transfersomes are biocompatible and biodegradable, since they are made of liposomes like normal phospholipids. In the case of lipophilic medicines, they have high capturing capacity, (nearly 90 percent more). They guard against the oxidative degradation of the encapsulated medication.

Transfersomes thus act as a warehouse, which display a slow and steady releasing of its contents. These can be used both for systemic and topical drug delivery. They are easy to scale up on a laboratory and industrial level, as the formulation parameters are clear and do not entail lengthy procedures and unnecessary use of unacceptable product additives.

Role of Skin

Skin is a significant body organ. The human skin is the biggest organ in the body and records for around 15 percent of the all-out body weight of adults. This is the most promptly accessible organs in the body with a thickness of only a couple of millimeters (2.97 ± 0.28

mm). Therefore, it is an essential organ in the body. It has several important roles, such as guarding against external-physical, chemical and biological attackers and avoiding unnecessary water loss from the body and moreover assumes a role in temperature control. Along with the mucous layers on the body surface, the skin is consistent. The skin plays an important role in deciding from the outside the underlying blood supply network. This is an obstacle to microbiological, chemical and physical attacks. This functions in body temperature as a thermostat. The skin controls blood level. This also prevents UV rays penetration in the body. The skin of an average healthy adult's body has a floor span of about 2 m², which is about a one-third of the circulating blood through the body. The skin functions as a barrier to the permeability of different chemicals and biological agents for transdermal absorption. The skin's diffusion resistance relies strongly on the anatomy. Certain basic skin characteristics are worth highlighting.

For the purposes of transdermal drug delivery, we are able to study the functions and the structures of four different layer of the human skin. These layers were further divided into the sub-cutaneous fat layer. The composite structure of the skin consists of the following distinct layers.

1. Subcutaneous Fat Layer

This is a fatty substance usually known as hypodermis. It spans between the overlying dermis and the under-lying body constituents. This layer is ordinarily flimsy in many pieces of the body. With a thickness of few millimetres. This fat tissue layer fundamentally is utilized to disconnect the body and give mechanical stun insurance. The subcutaneous greasy layer may likewise give a basic gracefulness of high-vitality atoms, while the fundamental veins and nerves are transmitted to the skin.

2. Dermis

It is a 3–5 mm thick as a rule and is the most critical piece of human skin. It is comprised of a system of connective tissues, for the most part collagen fibrils, upheld and adaptable tissue encased in a muco-polysaccharide gel. Inside it are numerous segments: blood and lymphatic vessels, sensitive spots, pilo-sebaceous units (hair follicles and sebaceous organs) and sweat organs (eccrine and apocrine). It is likewise a piece of a few different segments and the epidermis is ensured physiologically. For transdermal medication gracefulness, this layer is frequently considered as basically gelled water and in this manner gives an insignificant

obstruction to most polar medications, despite the fact that the dermal boundary can be basic when profoundly lipophilic particles are provided.

3. Epidermis

By means of various layers, the epidermis is additionally evaluated. The base layer of the epidermis is the layer germinativum. Over the basal layer is the layer spinosum layer, the layer granulosum layer, and the layer lucidum layer lastly the layer corneum layer.

4. Stratum Corneum

This is an important layer that does not allow the passage of the chemical matter inside and also externally by smoothening keratin cells (for example corneocytes). Stratum corneum is specialised membrane which contains cells that are cornified and evened-out as they enter this layer. The corneocytes would then be slopped off the skin at a pace of around one cell layer daily. The layer corneum is the essential wellspring of protection from entrance and saturation throught the skin. Cytoplasmic protein lattices involving keratin implanted in extracellular lipid are around 15-20 m thick across the majority of the human body. The skin would thus be able to be basically portrayed as a bi-overlaid. The film and the particles infiltrating through the vascular lipophilic (and quick foundational circulation) will pass.

[Figure 3: About Here]

Constitutional structure and Method of action

The carriers are composed of at least one amphipathic (similar to phosphatidyl-choline) compound. The compound sets into lipid bilayers and closes into one lipid vesicle in aqueous solvents. The additional versatility and permeability of lipid bi-laerys (such as a bioconservative or an amphiphile drug) is considerably improved by at least one bilayer softing factor. For improved versatility and permeability, translator vesicles can be updated and restructured.

Therefore, the local concentration of each part of the two layer is altered to the local stress of the bilayer, as shown in the figure. In its basic structure the transfersoma is mostly similar to a liposome, primarily because it is more deformable and flexible because of the more conventional vesicle. Another rise in the affinity to bind and retain water. The positive consequence of strong deformations in two layers.

The ultra-deformable hydrophilic vesicle is designed to avoid dehydration, which may require a similar method of transport but not the same osmosis forward. For example, a transference vesicle such as the non-occluded skin tend to penetrate the barrier and migrate to deeper, water-rich strata to ensure an appropriate level of humidity. Barriers must be penetrated in order to reversibly deform each other. The dignity of the vesicle and the hazard features of the underlying affinity and gradient cannot be undermined unacceptably, however. Therefore, the use of vesicular transfersomes in the delivery of pharmaceutical materials depends on the carrier's capacity to expand or remove pores in the skin or some other obstacle. Through the gradual release of drug carriers, drug molecules can be dispersed and eventually bonded to their goals. Transport to intracellular sites of a drug that often includes fusion of the carrier's lipid bilayer with the cell membrane except when the cell actively absorbs the vesicle during an endocytose cycle.

As it is too big to spread through the skin, the transfers must find their own way through the liver and follow it. Therefore, the usage of the vesicles transfersomes in the delivery of drug contents depends on the capacity of the carrier to expand and dissolve pores in the skin or some other barrier (such as vegetable cuticles). Thus, the gradual release of drug carriers facilitates the dispersal and subsequent binding of drug molecules to their objectives. Transport of medicinal products to a site where the lipid bilayer fusion of the carriers with the cell membrane is also required unless the cell actively absorbs the vesicle during an endocytose.

[Figure 4: About Here]

Methods of formulation of Transfersomes

There are several methods of formulation for transfersomes. Some of the most commonly used ways are as follows:

1. Vortexing sonicative process

Through this method, a phosphate buffer is mixed and turbulently suspended through milk with a mixture of lipids (phosphatidyl-choline, EA and therapeutic agent). Sonicating capacity and extraction by polycarbonate membranes shall be required for the suspension.

2. Suspension homogenisation process

This method prepares the transfersome with the correct amount of edgeactive molecules (for example, sodium cholate), mixing an ethanol soybean phosphatidylcholine solution. This pre

pared suspension is subsequently combined to achieve a total lipid concentration with the Tri ethanolamineHCl buffer. Sunken, froze and thawed 2 o 3 times is the resultant suspension.

3. Transformed hand shaking process

This process, commonly known as 'lipid film/ thin film-hydration,' prepares the transfersomes. Ethanol and chloroform are mixed into a standard mix in a (1:1) ratio. Later on, this mixture was made up of core material, lecithin (PC) and an edge activator. Then the organic solvent was isolated by evaporation during manual shaking over the lipid transition temperature (43 ° C). During continuous rotation, a thin lipid film was formed inside the bottle wall. This thin film was kept overnight for the overall solvent evaporation. In the end, a phosphate buffer (pH 7.4) was allowed to hydrate this film and shake it for 15 minutes at the correct equivalent temperature.

4. Lipid suspension in an aqueous phase

The drug-to-lipid ratio for vehicles in this cycle ranges from 1/4 to 1/9. The composition is preferred depending on the particular type of formulation. In comparison with the regular phosphatidyl choline vesicles, this will guarantee high flexibility of the vesicle membrane in the fluid phase. Specifically, soya-phosphatidyl choline with a standard size deviation (around 30 percent) is used to generate vesicles ranging from 100 to 200 nm. The lipids can be processed in an aqueous environment in which the drug is dissolved.

5. Method of centrifuge

Alcohol was used as the base fluid for phospholipid dissolution, surfactants and the API during this process. The rotary evaporation process is then used to remove the solvent at a low pressure of 40 ° C. The final solvent residue is removed under vacuum. The film hydrogenated at room temperature at 60 rpm for one hour with an appropriate buffer. At the earlier used room temperature the resultant vesicles can swell for over 2 hours. More sounding at room temperature ranging from 26 to 28 is achieved by the multi-lamella lipid vesicles.

Characterizations of Transfersomes

The transfersomes are characterized for the following mentioned parameters which are almost similar to the liposomes, niosomes and micelles.

1. Surface Charge and Charge Density

Malvern Zetasizer is a computerized inspection instrument used to assess vesicle size and zeta potential for surface charging. The Dynamic Light Spreading Method (DLS) is used to determine the charging density of transfersomes.

2. Size and zeta potential

Dynamic Scattering Light (DLS) method for measuring particle size and zeta potential was calculated using the Particel Size Analyzer. The drug was dissolved with a particulate size analyzer in 9 ml of distilled water 1 ml of suspension. Laser: Doppler anemometry was used for measurements using Zetasizer Nano-Z (Malvern Instruments, Malvern, UK). Possibility and/or scale of Zeta.

3. Morphology of the vesicle

For the vesicle diameter calculation, photon correlation spectroscopy or DLS approach is widely used. A 0, 2 mm membrane filter was taken to filter the prepared sample and diluted with filtered saline and then measured in sizes using Photon spectroscopy. Measurement of the correlation or DLS. The integrity of the bladder can be measured by calculating the size and shape of the bladder with regard to time. For scale and institutional change evaluation, DLS and TEM used respectively.

The transfersomal vesicle is also evident by transmission electron microscopy (TEM) with a 100 kV accelerating voltage. Without sonics, an optical microscope may visualize the shape of transfersome vesicles.

4. Efficiency of trapping

The capture efficiency was determined by the first isolation of the untrapped material using a mini column centrifugation method. Upon centrifugation, the vesicles were interfered with with 0, 1% Triton X-100 or 50% npropanol.

Entrapment efficiency = (total sum added) = 100. Entrapment efficiency

5. In vitro drug permeation through the skin

This study of characterisation was conducted in vitro in a phosphate buffer (pH 7.4) solution using fresh goat skin for the drug permeation analysis. The recipient compartment used the modification Franz diffusion cell, with a capacity of 50 ml, and an effective 2.50 cm² diffusion region. Duration tests were carried out using the skin of the abdominal cabbage.

The abdominal skin had hair stripped away and in the normal saline solution skin hydration was performed.

The adipose tissue coating has been removed by rubbing with a cotton swab. In the isopropyl alcohol solution, the skin was treated with 0-40 ° C. To check your skin. The skin was permeated horizontally in a receptor cell's Franz diffusion cell and the stratum corneum in the donor cell facing upward. The receptor portion was supplied with 50 ml (pH 7.4), packed with a 37 ± 0.5 THC and combined with a magnetic bar at 100 RPM.

Formulation was added and the top of the cell stitched (10 mg equivalent drug). At proportionate daily intervals, 1 ml of Aliquots of the receptor medium was taken away and a fresh phosphate buffer (pH 7.4) was rapidly replaced to control the sink in the same amount. The correction factors for each aliquot were taken into account in deciding the release profile. A sample can be tested using instrumental research methods.

6. Evaluation of the drug content

It has been measured with instruments such as an HPLC with a UV detector, column oven, auto check, pump and the device application. This is achieved using instrumental analytical methods. Such more resources differ by type of API.

Applications of Transfersomes

1. Insulin supply

Transfer drugs are an effective tool for the noninvasive and therapeutic use of these massive molecular drugs on the skin. Insulin is usually provided by a subcutaneous route that is painful. Transfersulin (insulin encapsulation) solves these two problems. Transfersulin was applied to the untouched skin and, depending on the specific carrier material, the initial symptoms of systemic hypoglycaemia were observed after 90 to 180 min.

2. Corticosteroid supply

Corticosteroids are also very easily supplied with transfersomes. The transfers improve the location and patient safety of the corticosteroid supply by optimizing epicutaneous dose administration. These doses are multiple times smaller than those currently used for skin disease treatment. Transfersomes are also bioactive.

3. Protein and peptide supply

Transfersomes were very common as a transport carrier for proteins and peptides recently. The proteins and peptides are large biogenic molecules, which are very difficult to absorb in the human body and are completely degraded when given orally in the GI tract. It is also why these peptides and proteins must be introduced into the body in vaccines. 450 different strategies have been created to enhance these circumstances. A bit like a subcutaneous human Bioavailability resulting from transfersomes was the injection of the same protein suspension.

The transfersomal preparations of this protein have induced strong immune response after many dermal challenges and counterpart injecting proteo-transfersomals, after repeated application of epicutaneous agents such as Adjuvant Immunogenic Serum Albumins.

4. Interferons delivery

Transfersomes have been used commonly as interferon carrier (such as INF- α) for an antiviral, antiproliferating, and other immunomodulatory activity, which naturally occurs as a leukocytic protein. Transfersomes can provide controlled release as well as enhance labile drug stability as drug-delivery systems. Hafen *et al.* Investigated the formulation of transfersomes for the potential transdermal application of interleukin-2 and interferone- α and the study indicates that IL-2 and INF- α transfersomes have been stuck with insufficient immunotherapy.

5. Anticancer therapeutics

For anti-cancer drugs such as methotrexate, transdermal delivery with transfersome technology was studied. The outcome was fine. This was a new method of treating cancer of the head.

6. Anesthetic delivery

In the suspension of strongly deforming vesicles, transferable surfaces cause atypical anesthesia at a rate of less than 10 min, under sufficient conditions. The total insensitivity to pain resulting in a subcutaneous bolus injection is almost as high (80 per cent), but is longer.

7. NSAIDS supply

NSAIDS increases the extent of GI side effects. Ultradeflexible vesicles overcome these problems by transdermal delivery. Tests were conducted with diclofenac and ketoprofen. The

medication was approved to be marketed by Swiss Regulatory Agency (Swiss Medic), under the trade name Diractin (in transfersomal formulations). Clinical studies and further therapies based on transfersoma technology have been introduced to IDEA. According to the IDEA AG.

8. Herbal medicines supply

Xiao-Ying et al, which shows greater topical absorption compared to pure capsaicin, have prepared the transfers of capsaicin. Transferomes can penetrate the stratum and provide the nutrients locally for the functions of skin maintenance.

FUTURE PERSPECTIVES

Transfersomes work on number of mechanisms working together to give an incredible transporter framework to the medication transport. The high bearableness and proficiency of these vesicular frameworks open huge expected restorative employments. These nano-transporters can demonstrate to offer propelled nearby and foundational new treatments with operators that are in any case incapable to proficiently infiltrate the layer corneum via passive diffusion. Further, many new therapeutic products based on the transfersosome technology are under constant clinical research and development.

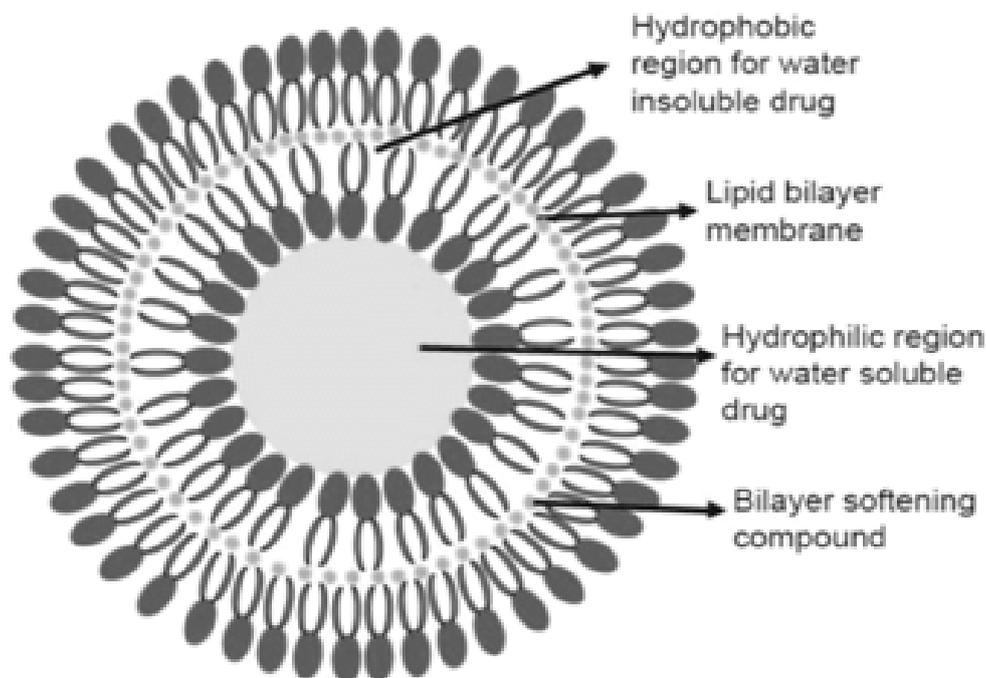


Figure 1: Front View of Transfersome.

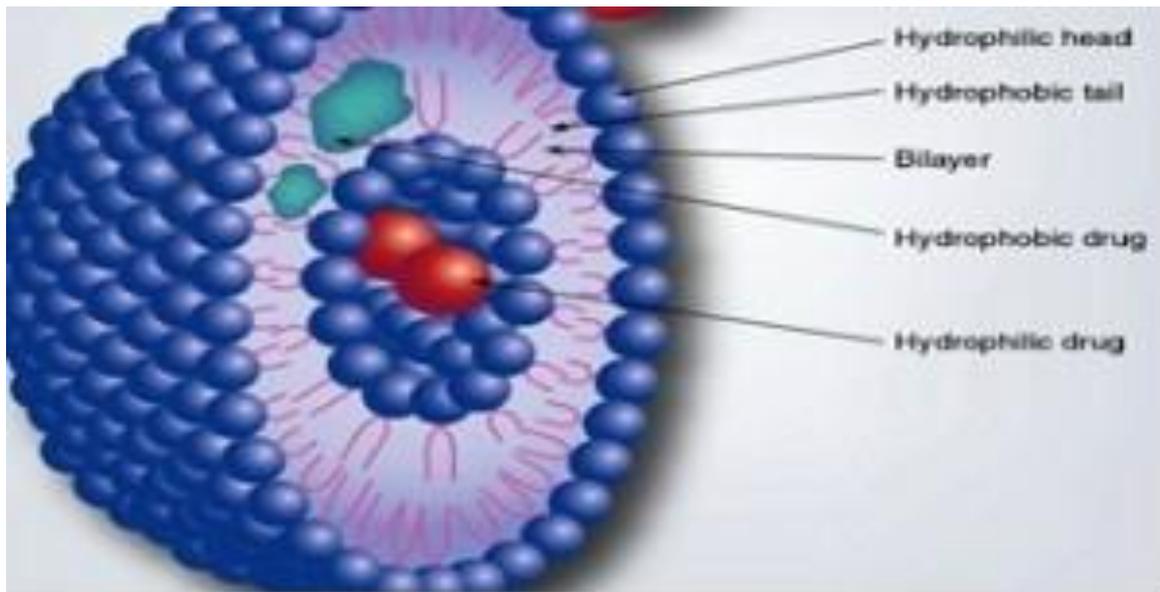


Figure 2: Cross-sectional View of Transfersome.

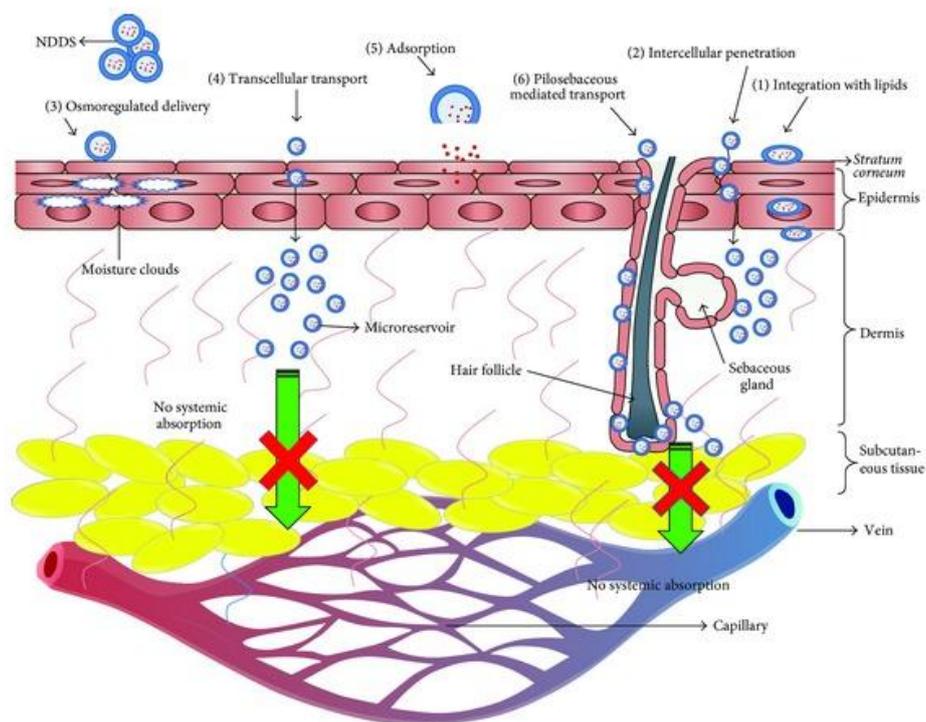


Figure 3: Various mechanisms of penetration of drug loaded NDDS across skin.

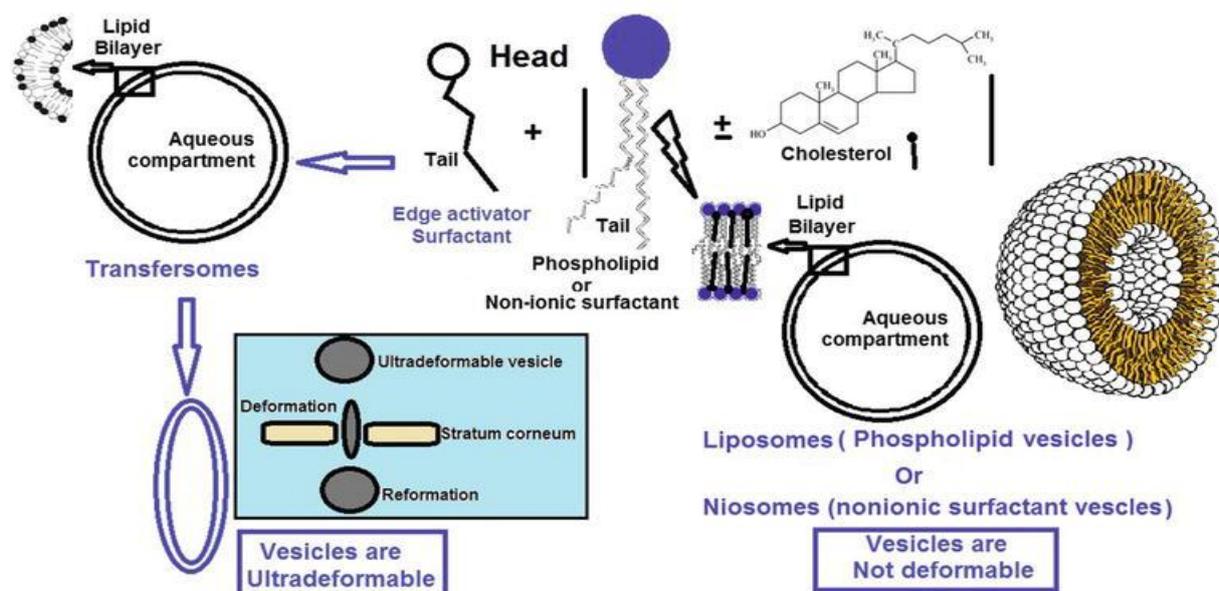


Figure 4: Schematic Representation of the Transfersomal Penetration Pathway.

Table 1: Different additives used in formulation of transfersomes.

S.No	Class	Example	Use
1	Phospholipids	Soya phosphatidyl- choline, egg phosphatidyl-choline, di-palmitoyl phosphatidyl choline	This is a component which forms the vesicles.
2	Surfactants	Sodium cholate, Sodium deoxycholate, Tween-80, Span-80, Tween 20	This is a component which forms the vesicles.
3	Solvents	Ethanol, methanol, isopropyl alcohol, chloroform	This is a component which serves as a solvent.
4	Buffering agent	Saline phosphate buffer (pH 6.4), phosphate buffer pH 7.4	As good medium for providing hydration.
5	Dye	Rhodamine-123 Rhodamine-DHPE Fluorescein-DHPE Nile-red	This is a component for conducting CSLM.

CONCLUSION

Transfersomes constitute small vesicles that are more commonly narrower than those on the skin to respond to external stress and thus contribute to an increased transdermal flux of therapeutic agents. Transfersomes are particularly engineered vesicles which react quickly and energy efficiently to the external stress. In the case of medicinal drugs these deformable particles may also be used to cross the skin's biopermeability barrier. These can move with almost the same efficiency as water 1500 times smaller, if tested in artificial systems and even in small pores (100 nm). The transfersomes are an excellent supplier choice for the supply of drugs on the transdermal layers of the skin. Transfersomes form an important part of a modern drug delivery system because of their small size and efficient carriage characteristics.

In contrast with other vesicular structures such as niosomes and ethosomes, transfersomes have many beneficial aspects. Pass ranges Show high potential skin penetration, increased stability, enhanced systemic drug releases and reduced malformations. Transfersomes thus affect the health system greatly. In future a number of transfersomes for the dermal and transdermal applications are likely to be produced. This new company has a wide range of new opportunities and is an emerging product which can be further explored.

REFERENCES

1. Lade Bipin et al and Shanware Arti et al, 2020, Phytonanofabrication: Methodology and Factors Affecting Biosynthesis of Nanoparticles, DOI: 10.5772/intechopen.90918.
2. Rofida Albas et al, Aly A Abdelbary et al, Hanan Refail et al, 2019, Use of transethosomes for enhancing the transdermal delivery of olmesartan medoxomil: in vitro, ex vivo, and in vivo evaluation, International Journal of Nanomedicine, 1953-68.
3. Chaurasiya Priyanka et al, Ganju Eisha et al, Upmanyu Neeraj et al, 2019; Transfersomes: a novel technique for transdermal drug delivery, Journal of Drug Delivery & Therapeutics, 9(1): 279-285.
4. Das Anirban et al, Sil Amrita et al, Sarkar Tushar et al, 2019, A randomized, double-blind trial of amorolfine 0.25% cream and sertaconazole 2% cream in limited dermatophytosis, | Volume : 85 | Issue : 3 | Page : 276-281.
5. Wick Y. Jeannette et al, 2019, Antifungals Are Key to Fighting Fungal Infections.
6. Narvekar Namita et al, Redkar Mayuresh et al, Bhosale Namita et al, 2019, SELF-ASSEMBLED ULTRADEFORMABLE PHOSPHOLIPID VESICLES WITH EDGE ACTIVATORS FOR DELIVERY OF TRANSCUTANEOUS BIOACTIVES, Indo American Journal of Pharmaceutical Research, ISSN NO: 2231-6876.
7. Kumar Abhay et al*, Nayak Amit et al, Ghatuary Sailesh et al, 2018, TRANSFEROSOME: A RECENT APPROACH FOR TRANSDERMAL DRUG DELIVERY, Journal of Drug Delivery and Therapeutics, pp-100-104.
8. Kanika Sahni et al, Sanjay Singh et al, Sunil Dogra1 et al, 2018, Newer Topical Treatments in Skin and Nail Dermatophyte Infections, Indian Dermatology Online Journal, IP: 47.247.178.162, | Volume 9 | Issue 3 |.
9. Qushawy Mona et al, Nasr Ali et al , Mohammed Abd-Alhaseeb et al, 2018, Design, Optimization and Characterization of a Transfersomal Gel Using Miconazole Nitrate for the Treatment of Candida Skin Infections, 3-18.

10. Mahmood, S. et al; Chatterjee B. et al; Mandal U. et al, 2018, Nano Transfersomes Vesicles of Raloxifene HCl with Sorbitan 80: Formulation and Characterization. *Bioequiv. Bioavailab. Int. J.*, 2: 1–7.
11. Iskandarsyah* et al, Aulia Dwi Rahmi et al, Dwita Medya Pangesti et al, 2018, Comparison of the Characteristics of Transfersomes and Protransfersomes Containing Azelaic Acid, *J Young Pharm*, 2018; 10(2) Suppl: s11-s15, *Journal of Young Pharmacists*, Vol 10, Issue 2 (Suppl).
12. Arora Amanjot et al, Dipankar De et al, Saikia Uma et al, 2018, Co-localization of immunobullous diseases at sites of dermatophytoses: Koebnerisation or a coinci..., *Indian Journal of Dermatology, Venereology, and Leprology*, Volume 84, Issue 6 [p. 712-717], DOI: 10.4103/ijdv.IJDVL_311_17 PMID: 30289119
13. Stolmeier Deirdre et al; Stratman Hannah et al; McIntee Thomas, MD et al; 2018; Utility of Laboratory Test Result Monitoring in Patients Taking Oral Terbinafine or Griseofulvin for Dermatophyte Infections, *JAMA Dermatol*, 154(12): 1409-1416. doi: 10.1001/jamadermatol.2018.3578.
14. Sarah Taylor et al, 2017, Skin Infection: Types, Causes, and Treatment.
15. Chen X, Jiang X, Yang M, et al. 2017, Systemic antifungal therapy for tinea capitis in children: An abridged Cochrane Review. *J Am Acad Dermatol*, 76(2): 368-374. doi: 10.1016/j.jaad.2016.08.061.
16. Satish K Mandlik et al, Shridhar S Siras et al, Kiran R Birajdar et al, 2019, Optimization and Characterization of Sertaconazole Nitrate Flexisomes Embedded in Hydrogel for Improved Antifungal Activity, PMID: 29160732, doi: 10.1080/08982104.2017.1402926. *Epub* 2017 Dec 4. DOI: 10.1080/08982104.2017.1402926; 29(1): 10-20.
17. Shubhra Rai et al, Vikas Pandey et al and Gopal Rai et al, 2017, Transfersomes as versatile and flexible nano-vesicular carriers in skin cancer therapy: the state of the art, *NANO REVIEWS & EXPERIMENTS*, VOL. 8, 1325708.
18. Abdul Ahad et al, Abdulmohsen et al., Al-Saleh et al, 2017, Formulation and characterization of novel soft nanovesicles for enhanced transdermal delivery of eprosartan mesylate, *Saudi Pharmaceutical Journal*, 2017; 25: 1040–1046.
19. Ibrahim Mokhtar et al, Nair Anroop et al, Aldhubiab Bandar et al, 2017, Hydrogels and Their Combination with Liposomes, Niosomes, or Transfersomes for Dermal and Transdermal Drug Delivery, DOI: 10.5772/intechopen.68158.

20. Nimisha et al, Rizvi DA et al, Fatima Z et al. Antipsoriatic and anti-inflammatory studies of *Berberis aristata* extract loaded nanovesicular gels. *Phcog Mag.*, 2017; 13(S3): 587-94.
21. Sugiyati R et al, Iskandarsyah et al, Djajadisastra J et al. 2017, Formulasi dan uji penetrasi *in vitro* sediaan gel transfersom mengandung kofein sebagai antiselulit. *Jurnal Ilmu Kefarmasian Indonesia*, 13(2): 131-6.
22. Abdellatif M et al.; Khalil I et al.; Khalil M et al., 2017, Sertaconazole nitrate loaded nanovesicular systems for targeting skin fungal infection: In-vitro, ex-vivo and in-vivo evaluation. *Int. J. Pharm.*, 527: 1–11.
23. Garg V et al, Singh H et al, Bhatia A et al. 2017, Systematic development of transethosomal gel system of piroxicam: formulation optimization, in vitro evaluation, and ex vivo assessment. *AAPS Pharmscitech*, 18: 58–71.
24. Zhang Y et al, Ng W et al, Feng X et al., 2017, Lipid vesicular nanocarrier: quick encapsulation efficiency determination and transcutaneous application. *Int J Pharm*, 10; 516: 225–230.
25. Hager CL et al, Long L et al, Ghannoum M. et al, 2017; 689 Efficacy of CD101, a novel echinocandin, in the treatment of dermatophytosis using a Guinea Pig (GP) Model. *J Invest Dermatol*, 137: S118.
26. Mohammed H. Elkomy et al, Shahira F. El Menshawe et al, Heba A. Abou-Taleb et al, 2017, Loratadine bioavailability via buccal transferosomal gel: formulation, statistical optimization, invitro/invivo characterization, and pharmacokinetics in human volunteers, *Drug Delivery*, 24: 1, 781-791, DOI: 10.1080/10717544.2017.1321061.
27. Kushwaha AS et al, Sharma P et al, Shivakumar HN et al, 2017; Trans-ungual Delivery of AR-12, a Novel Antifungal Drug. *AAPS PharmSciTec*, PP: 1-4.
28. Poulakos M et al, Grace Y et al, Machin JD et al, 2017; Efinaconazole and Tavaborole. *J Pharm Pract*, PP - 30: 245-55.
29. Novan - SB208 [Internet]. Novan Therapeutics, 2017, Available from: <http://www.novan.com/pipeline/sb208/>. [Last accessed on 2017 Sep 17].
30. Baghi N et al, Shokohi T et al, Badali H et al, 2016; *In vitro* activity of new azoles luliconazole and lanoconazole compared with ten other antifungal drugs against clinical dermatophyte isolates. *Med Mycol*, PP - 54: 757-63.
31. Tabata Y et al, Takei-Masuda N et al, Kubota N et al, 2016; Characterization of Antifungal Activity and Nail Penetration of ME1111, a New Antifungal Agent for Topical Treatment of Onychomycosis. *Antimicrob Agents Chemotherapy*, 60: 1035-9.

32. Evans JM et al, Wang AL et al, Elewski BE et al., 2016; Successful Treatment of *Paecilomyces lilacinus* Onychomycosis with Efinaconazole and Tavaborole. *Skin Appendage Disord*, 1: 169-71.
33. Zane LT et al, Chanda S et al, Coronado D et al, 2016; Antifungal agents for onychomycosis: New treatment strategies to improve safety. *Dermatol Online*, p-22.
34. Laska Amanda et al; Miller William et al; Dominguez Arturo et al, 2016; Purple Crusted Nodules in a Patient With End-stage Liver Disease, *JAMA Dermatol*, 152(8): 941-942. doi:10.1001/jamadermatol.2016.0935.
35. Ali I et al, Satti NK et al, Dutt P et al, 2016; Hydroxychavicol: A phytochemical targeting cutaneous fungal infections. *Sci Rep*, 6: 37867.
36. Hussain A et al, Singh VK et al, Singh OP et al, 2016; Formulation and optimization of nanoemulsion using anti-fungal lipid and surfactant for accentuated topical delivery of Amphotericin B. *Drug Deliv*, 23: 3101-10.
37. Pradhan Sulena et al, Jonas Hedberg et al, Blomberg Eva et al, 2016, Effect of sonication on particle dispersion, administered dose and metal release of non-functionalized, non-inert metal nanoparticles, *Journal of Nanoparticle Research* volume 18, Article number: 285.
38. Mbah et al, Ibrahim MI et al, Builders PF et al, 2015, FORMULATION AND EVALUATION OF A TRANSFERSOMAL VESICULAR CARRIER SYSTEM FOR ENHANCED TOPICAL DELIVERY OF NIPRD-AF1, *Journal of Phytomedicines and Therapeutics*, 1(1): 24-40 Page 24, *JOPAT Vol. 15(1): 23 - 37*, ISSN 1118 – 1028.
39. Ascenso et al, Raposo et al, Batista et al, 2015, Development, characterization, and skin delivery studies of related ultradeformable vesicles: transfersomes, ethosomes, and transethosomes, *International Journal of Nanomedicine*, Dove Press, p 5837-5851.
40. Bseiso EA et al, Nasr M et al, Sammour O et al. 2015; Recent advances in topical formulation carriers of antifungal agents. *Indian J Dermatol Venereol Leprol*, 81: 457.
41. Jinna S et al, Finch J. et al, 2015; Spotlight on tavaborole for the treatment of onychomycosis. *Drug Des Devel Ther*, PP 9: 6185-90.
42. Badali H et al, Mohammadi R et al, Mashedi O et al, 2015; In vitro susceptibility patterns of clinically important Trichophyton and Epidermophyton species against nine antifungal drugs. *Mycoses*, 58: 303-7.

43. Quach Olivia et al; Sylvia Hsu et al, 2016; Perianal Dermatomyiasis during Secukinumab Therapy for Plaque Psoriasis, *JAMA Dermatol*, 152(4): 486-487. doi: 10.1001/jamadermatol.2015.4992.
44. Ardesna Kenit et al, Rohatgi Shaurya et al, Jerajani Hemangi et al, 2015, Successful treatment of recurrent dermatomyiasis with isotretinoin and itraconazole, *Indian Journal of Dermatology, Venereology, and Leprology*, Year, 2016; 82(5): [p. 579-582], DOI: 10.4103/0378-6323.183632 PMID: 27297276.
45. Bseiso Eman et al, Nasr Maha et al, 2015, Sammour Omaima et al, Recent advances in topical formulation carriers of antifungal agents, *Indian Journal of Dermatology, Venereology, and Leprology*, 81(5): [p. 457-463], DOI: 10.4103/0378-6323.162328 PMID: 26261140.
46. Gu J et al, Mao X et al, Li C et al. 2015; A novel therapy for laryngotracheal stenosis: treatment with ethosomes containing 5-fluorouracil. *Ann Otol Rhinol Laryngol*, 124(7): 561-6.
47. Fathi-Azarbayjani A et al, Ng KX et al, Chan YW et al. 2015; Lipid vesicles for the skin delivery of diclofenac: cerosomes vs. other lipid suspensions. *Adv Pharm Bull*, 5(1): 25-33.
48. Garg BJ et al, Garg NK et al, Beg S et al. 2015; Nanosized ethosomes-based hydrogel formulations of methoxsalen for enhanced topical delivery against vitiligo: formulation optimization, in vitro evaluation and preclinical assessment. *J Drug Target*, 12: 1-14.
49. C Abhinav et al, Mahajan Vikram et al, Mehta Karaninder et al, 2015, Allergic contact dermatitis due to clotrimazole with cross-reaction to miconazole, *Indian Journal of Dermatology, Venereology, and Leprology*, 81(1): [p. 80-82], DOI: 10.4103/0378-6323.148592 PMID: 25566914.
50. Balasubramanian Pradeep et al, Jagadeesan Soumya et al, Thomas Jacob et al, 2015, Diaper dermatitis with psoriasiform id eruptions, *Indian Journal of Dermatology, Venereology, and Leprology*, 81(4): [p. 435], DOI: 10.4103/0378-6323.156194 PMID: 25937139.
51. Jachiet Marie et al; Lanternier Fanny et al; Rybojad Michel et al; 2015; Posaconazole Treatment of Extensive Skin and Nail Dermatomyiasis Due to Autosomal Recessive Deficiency of CARD9, free access *JAMA Dermatol*, 151(2): 192-194. doi: 10.1001/jamadermatol.2014.2154.

52. Guan P et al, Lu Y et al, Qi J et al, 2016; Readily restoring freeze-dried pro-biosomes as potential nanocarriers for enhancing oral delivery of cyclosporine A. *Colloids Surf B: Biointerfaces*, 144: 143-51.
53. Iizhar SA et al, Syed IA et al, Satar R et al, 2016; *In vitro* assessment of pharmaceutical potential of ethosomes entrapped with terbinafine hydrochloride. *J Adv Res.*, 7: 453-61.
54. Meng S et al, Zhang C et al, Shi W et al, et al. 2016; Preparation of osthole- loaded nano-vesicles for skin delivery: characterization, *in vitro* skin permeation and preliminary *in vivo* pharmacokinetic studies. *Eur J Pharm Sci.*, 92: 49–54.
55. Shashank J et al, Niketkumar P et al, Mansi KS et al. 2016; Recent advances in lipid-based vesicles and particulate carriers for topical and transdermal application. *J Pharm Sci.*, 116: 1–23.
56. Hassanpour AM et al, Ghanbarzadeh S et al, Javadzadeh Y et al. 2016; Aggregated nano-transfersomal dry powder inhalation of itraconazole for pulmonary drug delivery. *Adv Pharm Bull*, 6: 57–64.
57. González-Rodríguez ML et al, Arroyo CM et al, Cózar-Bernal MJ et al. 2016; Deformability properties of timolol-loaded transfersomes based on the extrusion mechanism, statistical optimization of the process. *Drug Dev Ind Pharm*, 42: 1683–1694.
58. Shreya AB et al, Managuli RS et al, Menon J et al. 2016; Nano-transfersomal formulations for transdermal delivery of asenapine maleate: *in vitro* and *in vivo* performance evaluations. *J Liposome Res.*, 26: 221–232.
59. Seidel EJ et al, Rother M et al, Regenspürger K et al. 2016; A randomized trial comparing the efficacy and safety of topical ketoprofen in Transfersome (®) gel (IDEA-033) with oral ketoprofen and drug-free ultra-deformable Sequeosome™ vesicles (TDT 064) for the treatment of muscle soreness following exercise. *J Sports Sci.*, 34: 88–95.
60. Al Shuwaili AH et al, Rasool BK et al, Abdulrasool AA et al. 2016, Optimization of elastic transfersomes formulations for transdermal delivery of pentoxifylline. *Eur J Pharm Biopharm*, 102: 101-14.
61. Punasiya R, Joshi A, Gupta S, Punasiya J. Transfersomes- A Novel Carrier for Transdermal Drug Delivery. *Research J. Pharma. Dosage Forms and Tech.*, 2010; 2(2): 133-138.
62. P.Sivannarayana, A. Prameela Rani, V. Saikishore, Ch.Venu Babu, V. SriRekha. Transfersomes: Ultra Deformable Vesicular Carrier Systems in Transdermal Drug Delivery System. *Research J. Pharma. Dosage Forms and Tech.*, 2012; 4(5): 243-255.

- A. Santosh Kumar, P. Kavya Deepika, D. Nagasen, D.V. Dakshina Murthy, V. Sai Kishore. Transfersomes: A new vesicular carrier system in topical drug delivery. Res. J. Topical and Cosmetic Sci., Jan. –June 2013; 4(1): 26-31.
63. Madhumitha. V, S. Sangeetha. Transfersomes: A Novel Vesicular Drug Delivery System for Enhanced Permeation through Skin. Research J. Pharm. and Tech, 2020; 13(5): 2493-2501.