

## FORMULATION AND EVALUATION OF MUCOADHESIVE MICROSPHERES OF LORATADINE FOR NASAL DRUG DELIVERY

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### ABSTRACT

Intranasal administrative is an ideal alternative to the parenteral route for systemic drug delivery. Nasal mucosa consists of a rich vasculature and a highly permeable structure for systemic absorption. Drug administration through the nasal cavity is easy and convenient. Avoidance of first pass metabolism is the main advantage of nasal route drug delivery. Bioadhesive polymers are used as drug carriers for the nasal drug delivery. The advantage of using Bioadhesive polymer is increase in residence time of the formulation in the nasal cavity and thereby minimizing rapid mucociliary clearance of the therapeutic agent from the site of deposition.

**KEYWORDS:-** Loratadine, Mucoadhesive, Transdermal, Bioadhesive, Polymers, Nasal Delivery, Microspheres.

### INTRODUCTION

Mucoadhesive microspheres include microparticels and microcapsules (having a core of the drug) of 1-1000um in diameter and consisting either entirely of a mucoadhesive polymer or having an outer coating of it, respectively. Microspheres, in general, have the potential to be used for targeted and controlled release drug delivery; but coupling of mucoadhesive properties to microspheres has additional advantages, e.g. efficient absorption and enhanced bioavailability of the drug due a high surface to volume ratio, a much more intimate contact with the mucus layer, specific targeting of drug to the absorption site achieved by anchoring plant lectins bacterial adhesion and antibodies *etc.* on the surface of the microspheres. Mucoadhesive microsphere can be tailored to adhere to any mucosal tissue including those found in eye, nasal cavity, urinary and gastrointestinal tract, thus offering the possibilities of

localized as well as systemic controlled release of drugs. Application of mucoadhesive microspheres to the mucosal tissues of ocular cavity, gastric and colonic epithelium is used for administration of drugs for localized action. Prolonged release of drug and reduction in frequency of drug administration to the ocular cavity can highly improve the patient compliance. The latter advantage can also be obtained for drug administered intra-nasal due to reduction in mucociliary clearance of drug adhering to nasal mucosa.

## MATERIAL AND METHOD

**Table no. 1: List of material used.**

| S. No. | Chemicals/ Materials  | Manufactures/Suppliers   |
|--------|-----------------------|--|
| 1.     | Loratadine            | Microlab PVT.LTD. Baddhi, Himachal Pradesh                             |
| 2.     | Chitosan              | Thermo Fischer Scientific India PVT.LTD.,B-Wing, Delphi, Mumbai-400076 |
| 3.     | Methanol              | Jiangsu Huaxi International Trade Co. LTD.,<br>Made in China.          |
| 4.     | Acetic acid           | Qualinges Fine Chemicals   |
| 5.     | Glutaraldehyde        | Qualinges Fine Chemicals   |
| 6.     | Light liquid paraffin | Thermo Fischer Scientific India,PVT.LTD.,B-Wing, Delphi, Mumbai-400076 |
| 7.     | Heavy liquid paraffin | Qualinges Fine Chemicals   |
| 8.     | Acetone               | Sd Fine CHEM LIMITED, Mumbai   |
| 9.     | Hexane                | Sd Fine CHEM LIMITED, Mumbai   |
| 10.    | DOSS                  | Thermo Fischer Scientific India PVT.LTD.,B-Wing, Delphi, Mumbai-400076 |
| 11.    | Sodium hydroxide      | Qualinges Fine Chemicals, Thermo Electro, India PVT.LTD.,Navi Mumbai   |

**Table no. 2: List of equipments used.**

| S. No. | Equipments                    | Manufacturers/Suppliers                    |
|--------|-------------------------------|--|
| 1.     | Digital balance               | VibraEssae                                 |
| 2.     | Melting point apparatus       | Scientex                                   |
| 3.     | UV-1700 Spectrophotometer     | Shimadzu Corporation, Japan                |
| 4.     | Magnetic stirrer              | IKA®RW 20 digital, Remi Lab Stirrer        |
| 5.     | Scanning electron microscopy  | Shimadzu Corporation, Japan                |
| 6.     | Dissolution apparatus USPXXII | Electrolab Dissolution tester(USP)-TDT-06L |
| 7.     | Digital oven                  | Alkolite industries Ltd.                   |
| 8.     | Refrigerator                  | LG Refrigerator                            |
| 9.     | pHep® pocket sized pH meter   | Hanna Instruments                          |
| 10.    | FTIR Spectrophotometer        | Shimadzu corporation, Japan                |

## Formulation development

### Spectrophotometric methods for estimation of Loratadine-Spectrophotometric methods or estimation of Loratadine<sup>[16,42]</sup>

#### 3.1.2.1. UV Spectrophotometric studies

##### Calibration curve of Loratadine

A standard curve from the stock solution was obtained in the range of 10-50 $\mu$ g/ml concentration using methanol, distilled water and pH 6.8 phosphate buffer by measuring absorbance at specific range (nm).

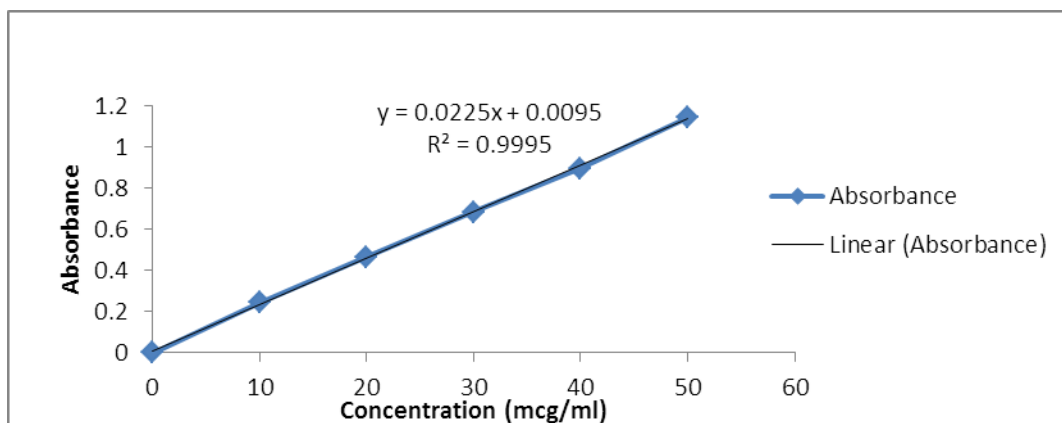
##### a) Calibration curve of Loratadine in Distilled water

Date 20.08.2013

|  |                 |
|--|-----------------|
| Calibration curve for                        | Loratadine      |
| Solvent                                      | Distilled water |
| $\lambda_{\max}$                             | 246             |
| Unit of concentration                        | mcg/ml          |
| Slope of Calibration curve                   | 0.022           |
| Constant of Calibration curve                | 0.009           |
| Coefficient correlation of Calibration curve | 0.999           |

**Table 3: Standard calibration table for loratadine in distilled water.**

| S. NO. | Concentration | Absorbance |
|--------|---------------|------------|
| 1.     | 0.0           | 0.00       |
| 2.     | 10.0          | 0.245      |
| 3.     | 20.0          | 0.484      |
| 4.     | 30.0          | 0.680      |
| 5.     | 40.0          | 0.888      |
| 6.     | 50.0          | 1.142      |



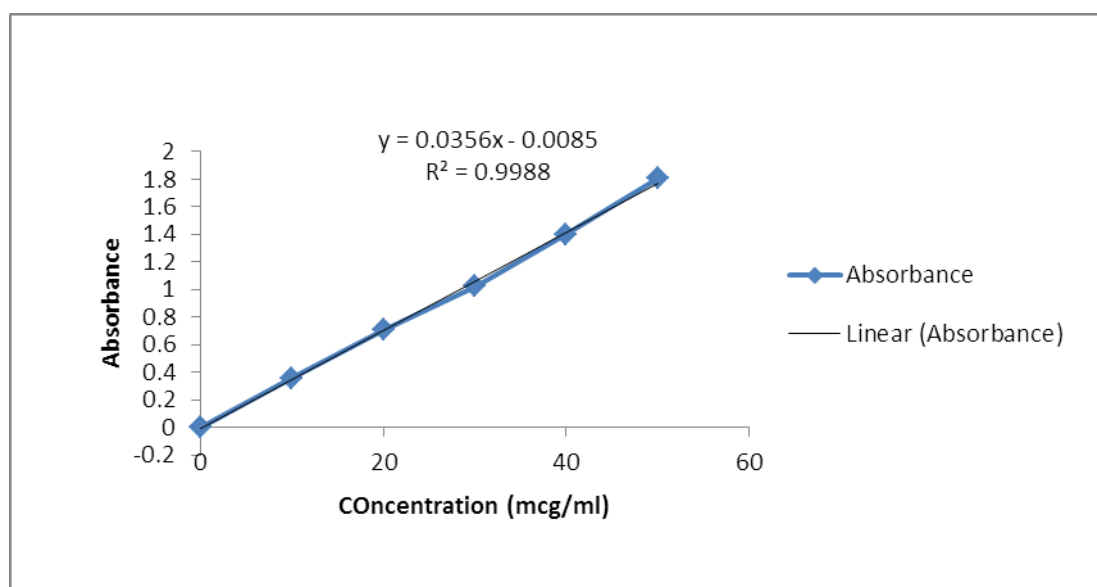
**Fig. no. 1 Calibration curve of loratadine in distilled water.**

**b) Calibration curve of Loratadine in methanol**

|   |            |
|---|------------|
| Date  | 19.08.2013 |
| Caliberation curve for                        | Loratadine |
| Solvent                                       | Methanol   |
| $\lambda_{\max}$                              | 247        |
| Unit of concentration                         | mcg/ml     |
| Slope of Caliberation curve                   | 0.0035     |
| Constant of Caliberation curve                | 0.008      |
| Coefficient correlation of Caliberation curve | 0.998      |

**Table 4: Standard calibration table for loratadine in methanol.**

| S. NO. | Concentration | Absorbance |
|--------|---------------|------------|
| 1.     | 0.0           | 0.0        |
| 2.     | 10.0          | 0.356      |
| 3.     | 20.0          | 0.709      |
| 4.     | 30.0          | 1.026      |
| 5.     | 40.0          | 1.399      |
| 6.     | 50.0          | 1.806      |

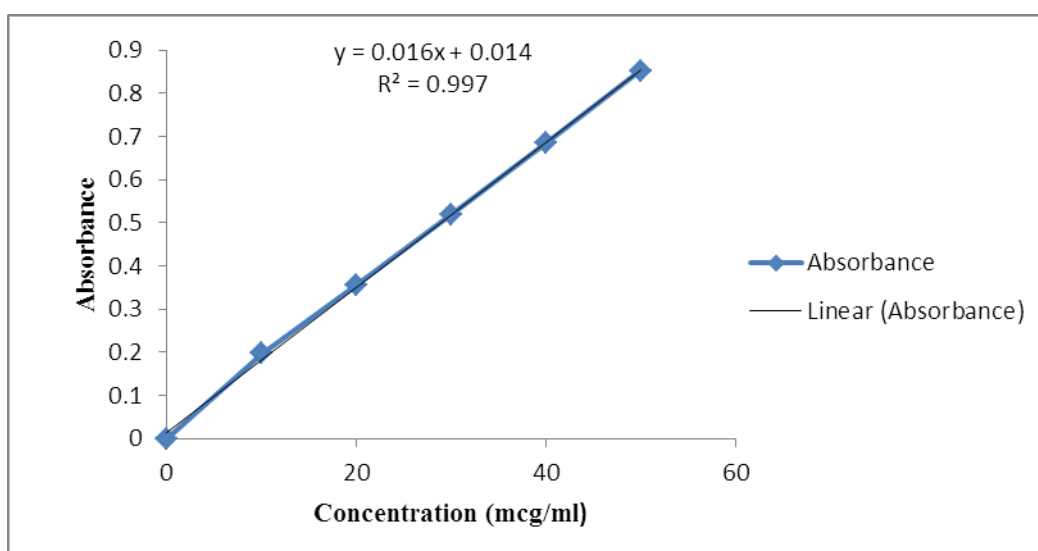
**Fig. no. 2: Calibration curve of loratadine in methanol.****c) Calibration curve of Loratadine in Phosphate buffer pH 6.8**

|                        |                      |
|------------------------|----------------------|
| Date                   | 14.08.2013           |
| Caliberation curve for | Loratadine           |
| Solvent                | phosphate buffer 6.8 |
| $\lambda_{\max}$       | 247                  |

|  |        |
|--|--------|
| Unit of concentration                        | mcg/ml |
| Slope of Calibration curve                   | 0.016  |
| Constant of Calibration curve                | 0.022  |
| Coefficient correlation of Calibration curve | 0.997  |

**Table 5: Standard calibration table for loratadine in phosphate buffer 6.8.**

| S. NO. | Concentration | Absorbance |
|--------|---------------|------------|
| 1.     | 0.0           | 0.0        |
| 2.     | 10.0          | 0.198      |
| 3.     | 20.0          | 0.356      |
| 4.     | 30.0          | 0.520      |
| 5.     | 40.0          | 0.685      |
| 6.     | 50.0          | 0.852      |



**Fig. no. 3: Calibration curve of loratadine in phosphate buffer 6.8.**

## RESULTS AND DISCUSSION

### Characterisation of mucoadhesive microspheres<sup>[52,53,54]</sup>

#### a) Determination of particle size

The mean particle sizes of the formulations is shown in the table 3.8 and mean particle size is depicted in fig No.3.8. The particle size mainly depends on the stirring rate and slow effect of concentration of mucoadhesive polymers, it is clear that stirring rate increases particle size decreases both at higher and lower concentration of polymers while concentration of mucoadhesive polymer had opposite effect on particle size.

| Micromer measurement test report |  |
|----------------------------------|--|
| Name : Shilpi sonkar             | Application:   |
|                                  | Operator:  |
| Sample info id :                 | Microscope obj: Formulation of loratadine and chitosan |

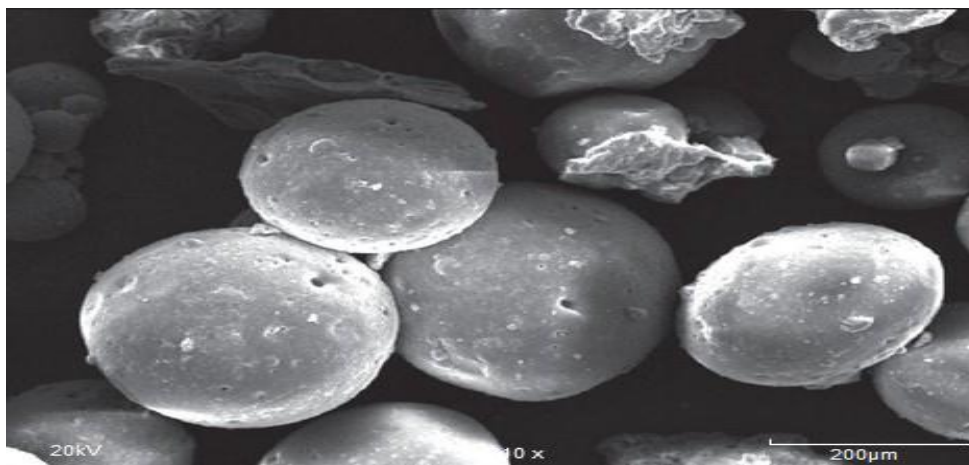


**Fig. no. 4: Optical microscopy of formulation (F3).**

**Table No. 3.8: Optical microscopy of formulation (F6).**

| S. NO.             | 1      | 2      | 3      | 4      |
|--------------------|--------|--------|--------|--------|
| Length (Micron)    | 58.79  | 11.448 | 11.448 | 11.448 |
| Length (Micron)    | 47.352 | 8.119  | 8.119  | 8.119  |
| Area (Micron Sqr.) | 100.00 | 73.046 | 73.046 | 73.046 |
| Asp. Ratio         | 1.242  | 1.41   | 1.41   | 1.41   |
| Roundness          | 100    | 100    | 100    | 100    |
| Shape              | 0.01   | 0.01   | 0.01   | 0.01   |

**b) Determination of particle size distribution by scanning electron microscopy.**



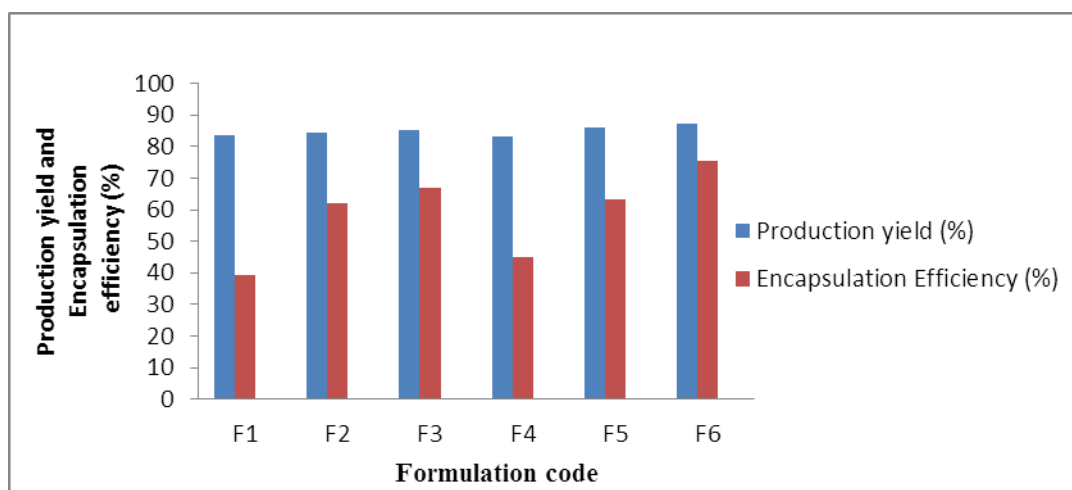
**Fig. no. 5: S. E. M. of Formulation (F3).**

### c) Production yield and Encapsulation efficiency.

The production yields of microspheres prepared by emulsion cross-linking method were found to be between 80-85 as shown in table No.3.9.

**Table 7: Production yield and Encapsulation Efficiency of formulations.**

| Formulation code | Production yield (%) | Encapsulation Efficiency (%) |
|------------------|----------------------|------------------------------|
| F1               | 83.5                 | 39.5                         |
| F2               | 84.5                 | 62.157                       |
| F3               | 85.0                 | 66.921                       |
| F4               | 83.0                 | 45.036                       |
| F5               | 86.0                 | 63.101                       |
| F6               | 87.25                | 75.298                       |



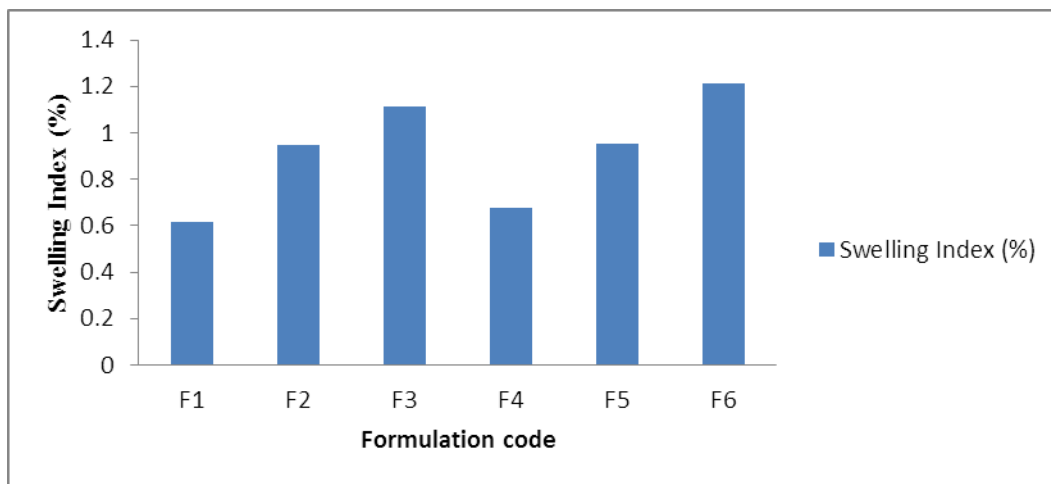
**Fig. no. 6: Bar Diagram to show production yield and encapsulation efficiency of formulations.**

### Swelling index

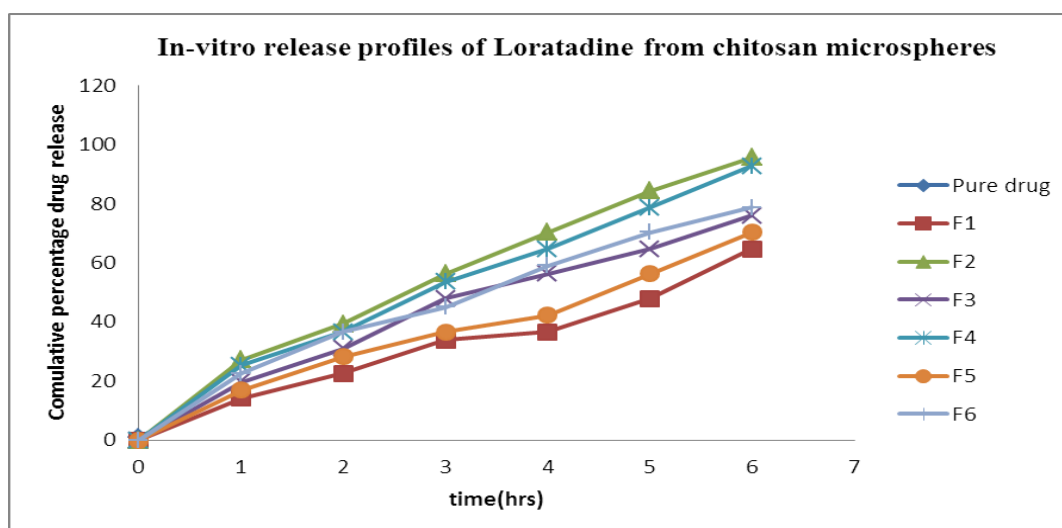
The swelling index of all formulation is shown in Table 3.

**Table 8: Swelling index of formulations.**

| Formulation | Swelling index (%) |
|-------------|--------------------|
| F1          | 0.615              |
| F2          | 0.945              |
| F3          | 1.115              |
| F4          | 0.675              |
| F5          | 0.955              |
| F6          | 1.215              |



**Fig. no. 7: Bar diagram to show swelling index of formulations.**



**Fig. no. 8 In vitro drug release of all formulations.**

#### d) In vitro drug release kinetics studies

The in-vitro drug release data of all the formulations were fit into Zero order, First order, Higuchi Equation and Korsmeyer-Peppas model. The results are shown in table

**Table no. 10: Correlation coefficient 'R' values in the analysis of release data of microspheres as per various kinetics model and 'n' value of Peppas equation.**

| Formulations | Zero order           | First order          | Higuchi matrix       | Peppas plot          |           |
|--------------|----------------------|----------------------|----------------------|----------------------|-----------|
|              | R <sup>2</sup> value | R <sup>2</sup> value | R <sup>2</sup> value | R <sup>2</sup> value | 'n' value |
| F1           | 0.977                | 0.931                | 0.908                | 0.976                | 0.811     |
| F2           | 0.980                | 0.976                | 0.963                | 0.933                | 0.762     |
| F3           | 0.988                | 0.947                | 0.958                | 0.989                | 0.733     |
| F4           | 0.983                | 0.982                | 0.931                | 0.982                | 0.761     |
| F5           | 0.984                | 0.896                | 0.965                | 0.995                | 0.701     |
| F6           | 0.992                | 0.889                | 0.960                | 0.987                | 0.704     |



Table no. 11: Zero order release kinetics of all formulations.

| Time (hrs) | Cumulative percentage drug release |        |        |        |        |        |
|------------|------------------------------------|--------|--------|--------|--------|--------|
|            | F1                                 | F2     | F3     | F4     | F5     | F6     |
| 0          | 0                                  | 0      | 0      | 0      | 0      | 0      |
| 1          | 14.06                              | 19.687 | 25.312 | 16.875 | 22.50  | 27.125 |
| 2          | 22.50                              | 30.937 | 36.56  | 28.125 | 36.562 | 39.37  |
| 3          | 33.75                              | 47.81  | 53.437 | 36.56  | 45.00  | 56.25  |
| 4          | 36.56                              | 56.25  | 64.687 | 42.187 | 59.062 | 70.31  |
| 5          | 47.81                              | 64.687 | 78.75  | 56.25  | 70.312 | 84.375 |
| 6          | 64.687                             | 75.93  | 92.812 | 70.31  | 78.75  | 95.625 |

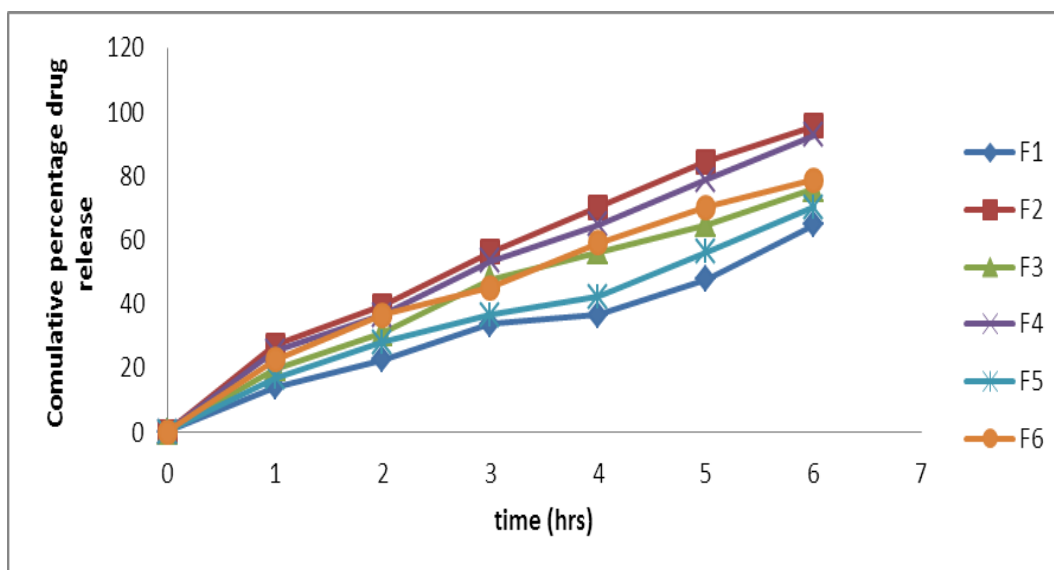


Fig. no. 9: Zero order release kinetics of all formulations.

Table no. 12: First order release kinetics of all formulations.

| Time (hrs) | log Cumulative percentage drug remaining |        |       |        |        |        |
|------------|--|--------|-------|--------|--------|--------|
|            | F1                                       | F2     | F3    | F4     | F5     | F6     |
| 0          | 2  | 2      | 2     | 2      | 2      | 2      |
| 1          | 1.934                                    | 1.904  | 1.873 | 1.919  | 1.889  | 1.856  |
| 2          | 1.889                                    | 1.839  | 1.821 | 1.856  | 1.802  | 1.782  |
| 3          | 1.821                                    | 1.717  | 1.668 | 1.802  | 1.74   | 1.6409 |
| 4          | 1.802                                    | 1.6409 | 1.547 | 1.762  | 1.612  | 1.4752 |
| 5          | 1.717                                    | 1.547  | 1.327 | 1.6409 | 1.4752 | 1.1938 |
| 6          | 1.547                                    | 1.3814 | 0.856 | 1.4752 | 1.327  | 0.64   |

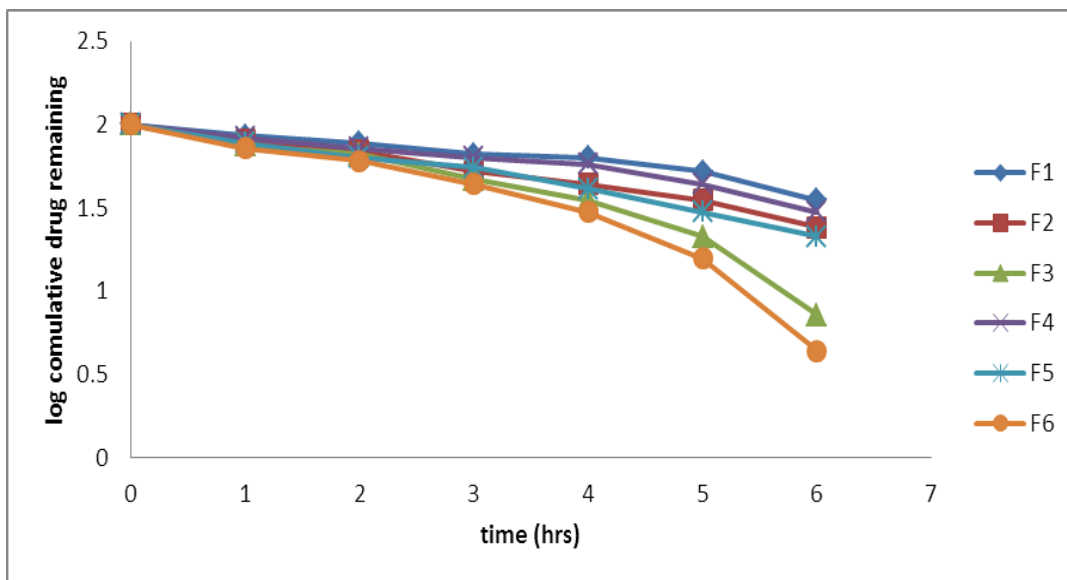


Fig. no. 10: First order release kinetics of all formulation.

Table no. 13: Higuchi matrix release kinetics of all formulations.

| Square root of time | Cumulative percentage drug release |        |        |        |        |        |
|---------------------|------------------------------------|--------|--------|--------|--------|--------|
|                     | F1                                 | F2     | F3     | F4     | F5     | F6     |
| 0                   | 0                                  | 0      | 0      | 0      | 0      | 0      |
| 1                   | 14.06                              | 19.687 | 25.31  | 16.875 | 22.5   | 27.125 |
| 1.414               | 22.5                               | 30.937 | 36.56  | 28.125 | 33.75  | 39.37  |
| 1.732               | 33.75                              | 47.81  | 53.437 | 36.56  | 45     | 56.25  |
| 2                   | 36.56                              | 56.25  | 64.687 | 42.187 | 59.062 | 70.31  |
| 2.236               | 47.81                              | 64.687 | 78.75  | 56.25  | 70.312 | 84.375 |
| 2.449               | 64.687                             | 75.93  | 92.812 | 70.31  | 78.75  | 95.625 |

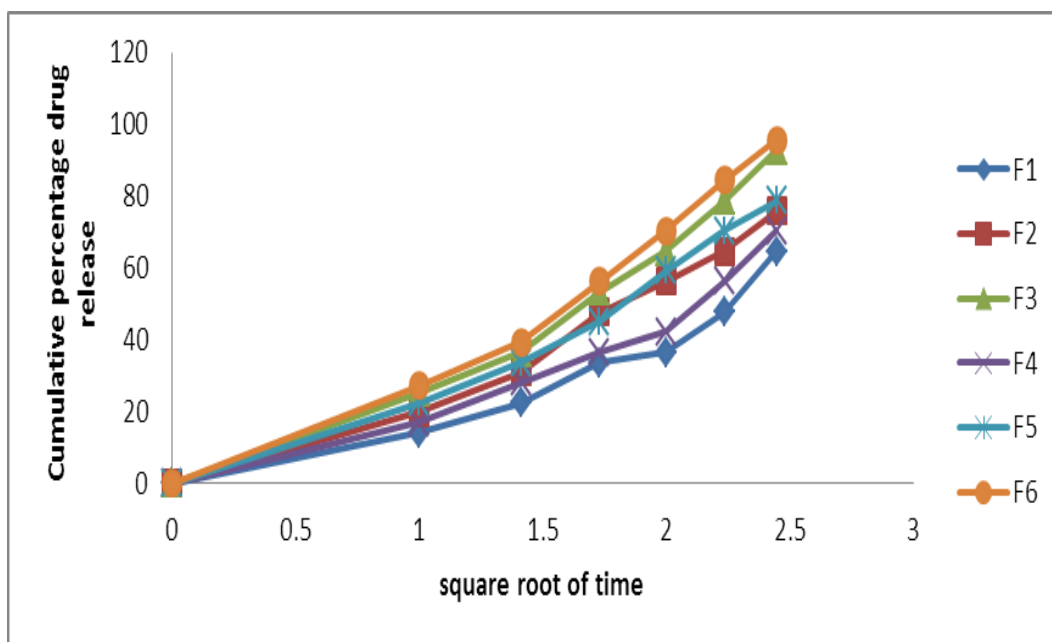


Fig. no. 11: Higuchi matrix release kinetics of all formulations.

Table no. 14: Korsmeyer peppas release kinetics of all formulations.

| Log time | log Cumulative percentage drug release |        |        |       |        |        |
|----------|--|--------|--------|-------|--------|--------|
|          | F1                                     | F2     | F3     | F4    | F5     | F6     |
| 0        | 1.147                                  | 1.294  | 1.4032 | 1.227 | 1.3521 | 1.449  |
| 0.301    | 1.352                                  | 1.4904 | 1.563  | 1.449 | 1.563  | 1.5951 |
| 0.4771   | 1.5282                                 | 1.6795 | 1.727  | 1.563 | 1.6532 | 1.751  |
| 0.602    | 1.563                                  | 1.751  | 1.8108 | 1.625 | 1.771  | 1.847  |
| 0.6989   | 1.6795                                 | 1.8108 | 1.896  | 1.751 | 1.847  | 1.926  |
| 0.7781   | 1.8108                                 | 1.88   | 1.9676 | 1.847 | 1.896  | 1.9805 |

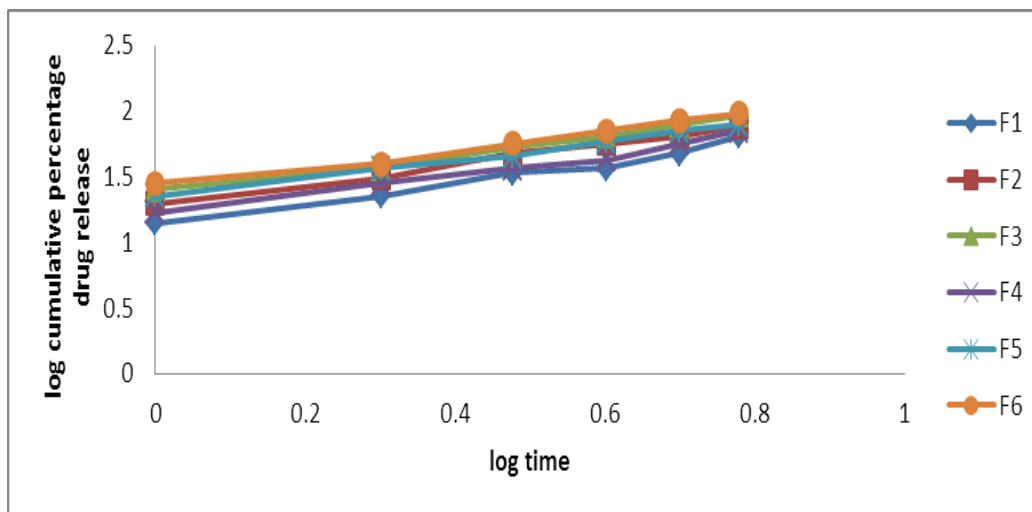


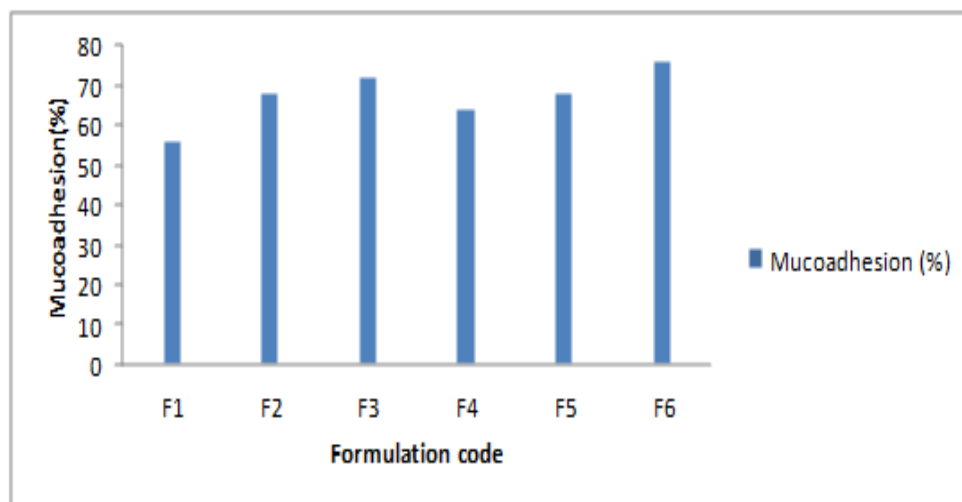
Fig. no. 12: Korsmeyerpeppas release kinetics of all formulations.

#### g) In-Vitro Mucoadhesion test for microspheres

The result of in-vitro mucoadhesion is shown in Table No.3.23. The percent mucoadhesion increased with increase in the concentration of mucoadhesive polymer. Such excellent mucoadhesion of chitosan microspheres was from the electrostatic attraction between chitosan and mucin. Moreover, the linear molecules of chitosan expressed sufficient chain flexibility for interpenetration and entanglement.

Table no. 15: Percentage mucoadhesion of formulations.

| Formulation code | Ratio (Drug : polymer) | Mucoadhesion (%) |
|------------------|------------------------|------------------|
| F1               | 1:1                    | 56               |
| F2               | 1:2                    | 68               |
| F3               | 1:3                    | 72               |
| F4               | 1:1                    | 64               |
| F5               | 1:2                    | 68               |
| F6               | 1:3                    | 76               |



**Fig. no. 13:** Bar diagram to show percentage mucoadhesion of all formulations.

#### h) Stability studies

Stability studies of the prepared Loratadine microspheres were carried out by storing the best formulation F3 & F6 at  $4\pm 1^\circ\text{C}$ ,  $25\pm 2^\circ\text{C}$  &  $60\pm 5\%$  RH and  $37 \pm 2^\circ\text{C}$  &  $65 \pm 5\%$  RH for one month.

**Table no. 16:** Stability studies of formulation (F3 & F6).

| Formulation code | $4\pm 1^\circ\text{C}$    | $25\pm 2^\circ\text{C}$ & $60\pm 5\%$ RH | $37\pm 2^\circ\text{C}$ & $65\pm 5\%$ RH |
|------------------|---------------------------|--|--|
|                  | Entrapment Efficiency (%) |  |  |
| F3               | 65.038                    | 66.758                                   | 60.123                                   |
| F6               | 74.345                    | 75.192                                   | 71.678                                   |

#### SUMMARY AND CONCLUSION

To achieve clinical efficacy is the most important criteria for any novel drug delivery system. A novel biodegradable and bioadhesive microspheres system has been developed for the intranasal delivery. Loratadine is a peripheral histamine H<sub>1</sub> receptors antagonist, which is used in the treatment of allergy & rhinitis. It is generally given by oral & parenteral route. However, it has poor bioavailability (%) by oral route which makes oral treatment unsatisfactory. Intranasal route may be a viable alternative for self-administration where the limitations of oral and parenteral route could be overcome. Conventional dosage forms may be unsatisfactory due to their poor residence time in nasal cavity. Mucoadhesive polymer like chitosan can be employed to increase the residence time of the formulation to enhance the bioavailability.

Hence, in the present work, an attempt was made to formulate and evaluate mucoadhesive microspheres of Loratadine that will increase residence time in the nasal cavity and at the same time increase the absorption.

orption of drug and its bioavailability. The microspheres were prepared by emulsion crosslinking method in different ratios by using mucoadhesive polymer, chitosan. The prepared microspheres were characterized for their drug entrapment efficiency & drug loading, particle size analysis, surface morphology, degree of swelling, in-vitro mucoadhesion, in vitro drug release behavior. Almost all the formulations showed fairly acceptable values for all the parameters evaluated. The results of all parameters are tabulated and depicted graphically in the results and discussion section. Stability studies revealed that the microspheres kept at 25°C showed the maximum stability. Thus, the prepared microspheres proved to be potential candidate as a microparticulate intranasal controlled drug delivery system.

## REFERENCES

1. A. Martinac, J. Filipovic-Grcic, D. Voinovich, B. Perissutti, E. Franceschinis, Development and bioadhesive properties of chitosan-ethylcellulose microspheres for nasal delivery, *International Journal of Pharmaceutics*, 2005; 69-77.
2. Patel J. K., Bodar M. S., Amin A. F., Patel M. M., Formulation and Optimization of Mucoadhesive microspheres of Metoclopramide. *Indian J. Pharm. Sci.*, 2004; 300-305.
3. Chowdary K.P.R. and Rao Y.S., Mucoadhesive Microspheres for Controlled Drug Delivery. *Biol. Pharm. Bull.*, 2004; 1717-1724.
4. Agrawal P., Rajput S., Pathak A., Shrivastava N., Microspheres a magical novel drug delivery system: a review, *World journal of pharmacy and pharmaceutical sciences*, 2012; 1(1): 439-455.
5. Singh P., Prakash D., B Ramesh, Singh N., T Tamizh Mani, Biodegradable Polymeric Microspheres as Drug Carriers ,A Review. *Indian Journal of Novel Drug delivery*, 2011; 70-82.
6. Manivannan R., Kugalur G.P., Recent advances in novel drug delivery system, *International journal of research in Ayurveda & pharmacy*, 2010; 316-326.
7. Garg A., Upadhyay P., Mucoadhesive Microspheres: A short review. *Asian journal of pharmaceutical and clinical research*, 2012; 5: 24-27.
8. Alexander A., Ajazuddin, Tripathi D.K., Verma T., Swarna, Maurya J., Patel S., Mechanism responsible for mucoadhesion of mucoadhesive drug Delivery system: a review. *International journal of applied biology and pharmaceutical technology*, 2011; 2(1): 435-445.

9. Vinod KR, Rohit Reddy T, Sandhya S, David Banji, Venkatram Reddy, Critical Review on Mucoadhesive Drug Delivery Systems. *Hygeia Journal for drugs and medicines*, 2012; 4: 7-28.
10. Singh A., Sharma P.K. and Malviya R., Sustained Drug Delivery Using Mucoadhesive Microspheres: The Basic, Concept, Preparation Methods and Recent Patents, *Recent Patents on Nanomedicine*, 2012; 2(1): 62-77.
11. Carvalho F.C., Bruschi M. L., Evangelista R.C., Mucoadhesive drug delivery systems, *Brazilian Journal of Pharmaceutical Sciences*, 2010; 46(1): 1-17.
12. Shaikh R., Singh T.R.R., Garland M.J., Mucoadhesive drug delivery systems, *Journal of Pharmacy and bioallied science*, 2011; 3(1): 89-100.
13. Sudarshan Singh, Mohan Govind and Sunil B Bothara. A Review on in vitro - in vivo Mucoadhesive Strength Assessment. *PhTechMed*, 2013; 2(1): 221-229.
14. Alagusundaram M, Chengaiah B., Gnanaprakash K., Ramkanth S., C.Madhusudhana Chetty, Dhachinamoorthi D., Nasal drug delivery system-an overview. *Int. J. Res. Pharm. Sci*, 2010; 1(4): 454-465.
15. AnaísaPires, Ana Fortuna, Gilberto Alves, and Amílcar Falcao, Intranasal Drug Delivery: How, Why and What for. *J Pharm PharmaceutSci*, 2009; 288 – 311.
16. Rahisuddin, Sharma P. K., Garg G, and MohdSalim, Review on nasal drug delivery system with recent advancement. *International Journal of Pharmacy and Pharmaceutical sciences*, 2011; 2: 6-11.
17. Yadav V.K., Gupta A.B., Kumar R., Jaideep S. Yadav, Mucoadhesive Polymers: Means of Improving the Mucoadhesive Properties of Drug Delivery System *.Journal of Chemical and Pharmaceutical Research*, 2010; 418-432.
18. Dey S., Mahanti B., Mazumder B., Nasal drug delivery: An approach of drug delivery through nasal route *.Pelagia Research Library Der Pharmacia Sinica*, 2011; 94-106.
19. Dhakar R.C., Maurya S.D., Tilak V.K., Gupta A.K, A review on factors affecting the design of nasal drug delivery system. *International Journal of Drug Delivery*, 2011; 194-208.
20. Choudhary Rakhi, Goswami Lakshmi, Nasal Route: A Novelistic approach for targeted drug delivery to CNS. *International Research journal of Pharmacy*, 2013; 59-62.
21. Ozsoy Y., Gungor S., Cevher E., Nasal Delivery of High Molecular Weight Drugs, *Molecules ISSN*, 2009; 3754-3779.
22. Swamy N G N, Zaheer Abbasb, Mucoadhesive *in situ* gels as nasal drug delivery systems: an overview, *Asian Journal of Pharmaceutical Sciences*, 2012; 168-180.

23. Basu S., Bandyopadhyay A.K., Nasal Drug Delivery: An Overview. *Int J PharmSci Tech*, 2010; 4(1): 1-19.
24. T. Praveen Kumar, Sirisha B., P. NarayanaRaju and G. Nagarjuna Reddy. Nasal drug delivery: a potential route for brain Targeting, *The pharma innovation –journal*, 2013; 2(1): 77-85.
25. Huha Y., Hyun-JongChoa, In-SooYoona, Min-Koo Choia, b, Jung Sun Kimc. Preparation and evaluation of spray-dried hyaluronic acid microspheres for intranasal delivery of fexofenadine hydrochloride. *European Journal of Pharmaceutical Sciences*, 2010; 9-15.
26. Atul S. Pratapwar, Vijay A. Agrawal and Aditya P. Chiddarwar, Formulation and biological factors influencing the absorption of drugs through nasal epithelium and current nasal formulations - An overview. *Pharmacophore (An International Research Journal)*, 2012; 3: 37-43.
27. Akhtar Ali, Prajapati S.K., Singh Devendra, Kumar Brijesh, Shafat Kausar. Enhanced bioavailability of drugs via intranasal drug delivery systems. *International research journal of pharmacy*, 2013; 68-74.
28. Tyagi S., Sharma N., Sharma P.K., A review on application of natural bioadhesive Polysaccharides for intranasal drug delivery. *ISSN*, 2012; 1(2): 80-94.
29. Kisan R. Jadhav, Manoj N. Gambhire, Ishaque M. Shaikh, Vilarsrao J. Kadam and Sambjahi S. Pisal, Nasal Drug Delivery System-Factors Affecting and Applications. *Current Drug Therapy*, 2007; 27-38.
30. Swatantra K.S. Kushwaha, Ravi Kumar Keshari and A.K. Rai, Advances in nasal trans-mucosal drug delivery. *Journal of Applied Pharmaceutical Science*, 2011; 21-28.
31. Saraswathi B., Balaji Annaand Umashankar M.S., polymers in mucoadhesive drug delivery system-Latest updates. *International journal of pharmacy and pharmaceutical sciences*, 2013; 5: 423-430.
32. Bhowmik D., Khare R., Jaiswal J., Chiranjib, Biswajit and K. P.SampathKumar. Innovative approaches for nasal drug delivery system and its challenges and opportunities, *Scholars Research Library Annals of Biological Research*, 2010; 21-26.
33. Sipai Altafbhai, Vandana Yadav, Mamatha Y, Prasanth V.V. Mucoadhesive Microspheres an Overview, *American journal of Pharmatech Research*, 2012; 237-258.
34. Bhanja S., Panigrahi B.B., Shukla N. Formulation and in-vitro evaluation of nicardipine hydrochloride microcapsules. *Asian Journal of Pharmaceutical and Clinical Research*, 2012; 5: 60-63.

35. The United state of Pharmacopeia, NF. The official compendia of standards. United state Pharmacopeial convention, INC, 2004; 656: 2492.
36. Martindale. The complete drug reference, Pharmaceutical press Lambeth high street London, 2007; 1: 527.
37. The Merck Index An encyclopedia of chemicals, Drugs and biological. Merck Research Laboratories division of Merck & Co., INC. Whitehouse station NJ, 1997; 5599.
38. Raymond C Rowe, Paul J Sheskey and Marian E Quinn. Handbook of Pharmaceutical Excipients. Pharmaceutical press An imprint of RPS, Lambert high street, London, 2006; 6: 159-162, 503-504.
39. Gilman's, Goodman. The Pharmacological basis of Therapeutics. International edition, Mc Graw- Hill Medical, New York, 654.
40. Jain N.K., Controlled and Novel drug delivery. CBS Publishers & Distributors, New Delhi, 2011; 236-251.
41. Banker S. Glibert, Rhodes T. Christopher. Modern Pharmaceutics. Informa healthcare, New York, 121: 268.
42. Chakraborty S., Nayak P., Krishna B.M., Khandai M., Ghosh A.K. Preparation and preclinical evaluation of aceclofenac loaded pectinate mucoadhesive microspheres. *Drugs and Therapy Studies*, 2012; 2: 36-42.
43. Naga Phani J.V.V, Mohan Varma M, Development and Evaluation of Novel Mucoadhesive Multiparticulate Drug Delivery System of Simvastatin. *Indian Journal of Pharmaceutical Education and Research*, 2013; 47(1): 62-70.
44. Brahmaiah B., Sasikanth K, Sreekanth Nama, Siva Sankara Sumar GV, Krishna Chaitanya, 2013. Formulation and evaluation of extended release mucoadhesive microspheres of rosuvastatin, *international journal of biological & pharmaceutical research*, 271-281.
45. Mangesh R. Bhalekar, Mangesh R. Bhalekar, Kalpesh P. Patil, Ashwini R. Madgulkar, Formulation Optimization and evaluation of mucoadhesive microspheres of Xyloglucan, *Int. J. Pharm. Health Sci.*, 2007; 1(1): 23-31.
46. Abdul Jaleel, Omar W1, Abdulrasool, Alaa A, Ghareeb, Mowafaq M., Preparation and Characterization of Orally Disintegrating Loratadine Tablets from PVP Solid Dispersion *International Journal of Pharmaceutical Sciences*, 2010; 759-768.
47. Ilango van Ponnilarasan, Chebrolu. Sunil, Kumar N., and P. Asha et al., Simultaneous estimation of ambroxol hydrochloride and Loratadine in tablet dosage form by using UV Spectrophotometric method, *International journal of pharma and bio sciences*, 2011; 2(2): 332-344.



48. Radha G. V., N. Lakshmi Sarvanthi, Swetha P, Formulation and Evaluation of Mucoadhesive Microspheres of Nifedipine. *Journal of Pharmaceutical scientific innovation*, 2012; 39-43.
49. Sandra Guerrero, Teijon C, Characterization and *in vivo* evaluation of ketotifen-loaded chitosan microspheres, *European Journal of Pharmaceutical Sciences*, 2010; 79(4): 17, 1006–1013.
50. HardeniaS.S., Jain A., Patel R., Kaushal A. Formulation and Evaluation of Mucoadhesive Microspheres of Ciprofloxacin. *Journal of Advanced Pharmacy Education & Research*, 2011; 214-22.